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Shimazu et al.

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(54) MUTATED ALKALINE CELLULASE

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(52) **U.S. Cl.**

CPC *C11D 3/38645* (2013.01); *C12N 9/2437* (2013.01); *C12Y 302/01004* (2013.01)

(58) Field of Classification Search

None

See application file for complete search history.

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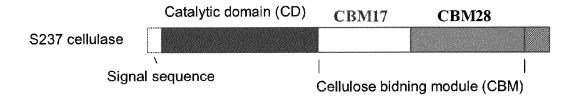
Primary Examiner — Iqbal H Chowdhury (74) Attorney, Agent, or Firm — Sterne, Kessler, Goldstein & Fox P.L.L.C.

(57) ABSTRACT

The present invention provides a mutant alkaline cellulase having enhanced anti-redeposition ability. The present invention provides a mutant alkaline cellulase having an amino acid sequence in which at least amino acid residue selected from the amino acid residues at the positions corresponding to positions 58, 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, 175, 176, 178, 179, 181, 193, 194, 195, 196, 197, 199, 202, 203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 312, 318, 324, 345, 354, 356, 357, 360, 363, 368, 418, 419, 420, 421, 422, 453, 454, 455, 457, 458, 459, 494, 495, 496, 500, 501, 502, 503, 504, 550, 551, 552, 604, 605, 606, 607, 608, 640, 641, 642, 644, 645, 646, 683, 684, 685, 690, 691, 692, 693, 694, 739, 740, and 741 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of the amino acid sequence of an alkaline cellulase having at least 70% identity with the amino acid sequence of SEQ ID NO: 2, is substituted with other amino acid residue.

11 Claims, 4 Drawing Sheets

FIG. 1



В

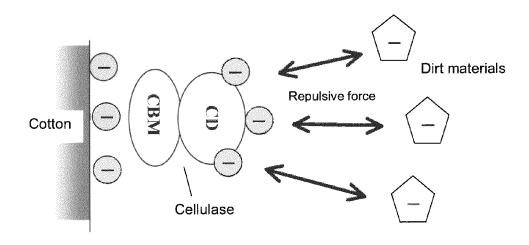
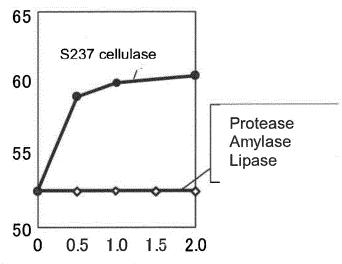


FIG. 2

A

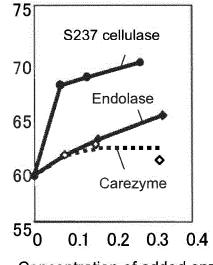
Anti-redeposition ratio (%)



Amount of enzyme incorporation (% detergent)

B





Concentration of added enzyme (mg/L)

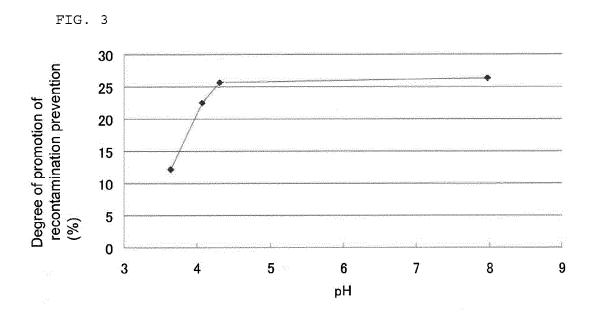
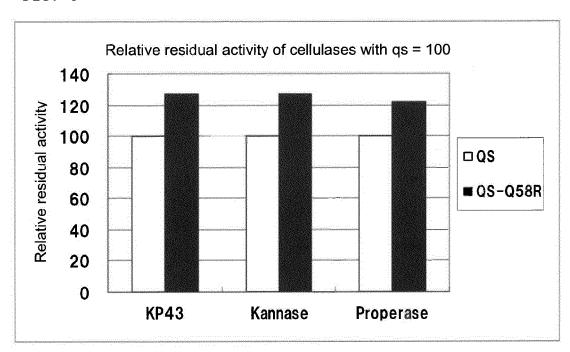


FIG. 4



MUTATED ALKALINE CELLULASE

FIELD OF THE INVENTION

The present invention relates to a mutant alkaline cellulase 5 having enhanced anti-redeposition ability, and a mutant alkaline cellulase having both enhanced anti-redeposition ability and enhanced protease resistance.

BACKGROUND OF THE INVENTION

For effective washing of clothing products and the like, it is important to sufficiently detach dirt materials from the object to be washed or to rapidly remove dirt materials through enzymatic degradation or the like, as well as seriously impor- 15 tant to prevent reattachment of dirt materials that have been once detached from the object to be washed (preventing of redeposition). Particularly, with regard to minute dirt materials such as soot, it is known that if the dirt materials are once diffused in washing water and then are reattached to the 20 object to be washed, it is very difficult to remove the dirt materials. Accordingly, various anti-redeposition agents have been conventionally incorporated into, for example, detergents for clothing products. Examples of the anti-redeposition agents that may be used include cellulose-based com- 25 pounds such as carboxymethyl hydroxypropylmethyl cellulose, and hydroxybutylmethyl cellulose; nonionic polymers such as polyethylene glycol, polyvinyl alcohol, and polyvinylpyrrolidone; and amino acid polymers. However, there have been needs for development 30 of the anti-redeposition agent having a further enhanced

Meanwhile, it has been a traditional practice to incorporate hydrolases such as proteases, lipases and amylases as cleaning aids, into cleaning agents. Cellulases, which constitute 35 one class of hydrolases, are originally known as enzymes that function in neutral to acidic condition, and it has been conventionally considered that cellulases are not suitable to be incorporated into alkaline cleaning agents for clothing. However, in recent years, alkaline cellulases or variants thereof are 40 obtained from plural organisms including thermophilic or alkalophilic bacteria of the genus *Bacillus*, and the cellulases or variants thereof may also be incorporated into alkaline detergents for clothing (for example, Patent Documents 1 to

Cellulases are enzymes which hydrolyze glycoside bond within β-1.4-glucans including cellulose. Cellulases are mainly produced by bacteria or plants and are available in various kingdoms. Cellulases are classified into endoglucanases (EC3.2.1.4) that degrade cellulose from the interior of 50 the molecule, and exoglucanases (cellobiohydrolases) (EC3.2.1.91) that degrade cellulose from the reduced ends or non-reduced ends of the sugar chain, and release cellobiose. On the other hand, glycoside hydrolases including cellulases are classified into one glycoside hydrolase family, and that 55 item [1], wherein the at last one amino acid residue is a family is currently further classified into subfamily 1 to subfamily 108. Cellulases are classified, based on their structures, into families 5, 6, 7, 8, 9, 10, 12, 44, 45, 48, 51, 61, and 74 of the glycoside hydrolase family, and it is also known that the amino acid sequence identity between those families is 60

However, in recent years, it has been found by analyses of amino acid sequences or three-dimensional structures that cellulases have a common domain having catalytic activity (catalytic domain; CD) and another common functional 65 domain that are linked to each other through a linker. A representative example of the other functional domain is a

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cellulose binding region having cellulose binding properties (also referred to as cellulose binding module: CBM) (see FIG. 1A). Cellulases usually have plural CBMs. Since cellulose is basically water-insoluble, a cellulase binds to the cellulose surface through the CBMs, and thereby relatively increases the substrate concentration. CBMs are also classified into 40 or more families based on the amino acid sequence identity. In regard to these CBMs, identification of amino acid residues that directly participate in binding to the cellulose has also been carried out (Non-Patent Document 1).

Patent Document 1 discloses a Bacillus sp. strain KSM-S237-derived alkaline cellulase having heat resistance, Patent Document 2 discloses a mutant alkaline cellulase having an optimum pH increased to near pH 10.5, and Patent Document 3 discloses a mutant alkaline cellulase having enhanced productivity. Furthermore, Patent Documents 4 and 6 describe that some of these cellulases show anti-redeposition activity.

PRIOR ART DOCUMENTS

Patent Document

Patent Document 1: JP-B-3512981 Patent Document 2: JP-A-2003-310270 Patent Document 3: JP-A-2004-140 Patent Document 4: JP-A-2004-536593 Patent Document 5: JP-A-2006-509850 Patent Document 6: JP-A-2002-265998

Non-Patent Document

Non-Patent Document 1: Boraston A. B. et al., Biochem. J., (2002) 361, p. 35-40

SUMMARY OF THE INVENTION

The present invention includes the following:

[1] A mutant alkaline cellulase having an amino acid sequence in which at least one amino acid residue selected from the amino acid residues at the positions corresponding to positions 58, 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, 175, 176, 178, 179, 181, 193, 194, 195, 196, 197, 199, 202, 203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 312, 318, 324, 345, 354, 356, 357, 360, 363, 368, 418, 419, 420, 421, 422, 453, 454, 455, 457, 458, 459, 494, 495, 496, 500, 501, 502, 503, 504, 550, 551, 552, 604, 605, 606, 607, 608, 640, 641, 642, 644, 645, 646, 683, 684, 685, 690, 691, 692, 693, 694, 739, 740, and 741 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of the amino acid sequence of an alkaline cellulase having at least 70% identity with the amino acid sequence of SEQ ID NO: 2, is substituted with other amino acid residue.

- [2] The mutant alkaline cellulase according to the above glutamine residue at the position corresponding to position 58 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of the amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2.
- [3] The mutant alkaline cellulase according to the above item [2], wherein the other amino acid residue is arginine or glutamic acid residue.
- [4] The mutant alkaline cellulase according to the above item [1], wherein the at least one amino acid residue is selected from non-charged amino acid residues at the positions corresponding to positions 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, 175,

176, 178, 179, 181, 193, 194, 195, 196, 197, 199, 202, 203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 312, 318, 324, 345, 354, 356, 357, 360, 363, and 368 of the amino acid sequence set forth in SEQ ID NO: 2, and the other amino acid residue is a charged amino acid residue.

- [5] The mutant alkaline cellulase according to the above item [4], wherein the charged amino acid residue is selected from the group consisting of glutamic acid, aspartic acid, lysine, arginine and histidine residue.
- [6] The mutant alkaline cellulase according to the above item [4], wherein the non-charged amino acid residue is alanine, serine, glutamine or asparagine residue, and the charged amino acid residue is glutamic acid, aspartic acid or arginine residue.
- [7] The mutant alkaline cellulase according to the above item [4], wherein the non-charged amino acid residue is at least one amino acid residue selected from the non-charged amino acid residues at the positions corresponding to positions 71 and 193 of the amino acid sequence set forth in SEQ 20 ID NO: 2, and the charged amino acid residue is an acidic amino acid residue.
- [8] The mutant alkaline cellulase according to the above item [1], wherein the at least one amino acid residue is selected from the amino acid residues at the positions corresponding to positions 418, 419, 420, 421, 422, 453, 454, 455, 457, 458, 459, 494, 495, 496, 500, 501, 502, 503, 504, 550, 551, 552, 604, 605, 606, 607, 608, 640, 641, 642, 644, 645, 646, 683, 684, 685, 690, 691, 692, 693, 694, 739, 740, and 741 of the amino acid sequence set forth in SEQ ID NO: 2.
- [9] The mutant alkaline cellulase according to the above item [8], wherein the at least one amino acid residue is selected from the amino acid residues at the positions corresponding to positions 419, 421, 454, and 501 of the amino acid sequence set forth in SEQ ID NO: 2.
- [10] The mutant alkaline cellulase according to the above item [8], wherein the at least one amino acid residue is tryptophan residue while the other amino acid residue is tyrosine residue, or the at least one amino acid residue is an amino acid other than tryptophan residue while the other amino acid 40 residue is alanine residue.
- [11] The mutant alkaline cellulase according to the above items [1] to [10], wherein a signal sequence consisting of the amino acid residues at the positions 1 to 30 of the amino acid sequence set forth in SEQ ID NO: 2, or of amino acid residues 45 equivalent to the foregoing amino acid residues, is deleted.
- [12] A gene encoding the mutant alkaline cellulase according to any one of the above items [1] to [11].
- [13] A recombinant vector including the gene according to the above item [12].
- [14] A transformant containing the recombinant vector according to the above item [13].
- [15] The transformant according to the above item [14], being a microorganism.
- [16] An anti-redeposition agent including the mutant alkaline cellulase according to any one of the above items [1] to [11].
- [17] An enzyme composition including the mutant alkaline cellulase according to any one of the above items [1] to [11].
- [18] The enzyme composition according to the above item [17], further including one or more enzymes selected from the group consisting of a protease, a cellulase, a β -glucanase, a hemicellulase, a lipase, a peroxidase, a laccase, an α -amylase, a glucoamylase, a cutinase, a pectinase, a reductase, an oxidase, a phenoloxidase, a ligninase, a pullulanase, a pectate 65 lyase, a xyloglucanase, a xylanase, a pectin acetylesterase, a polygalacturonase, a rhamnogalacturonase, a pectin lyase,

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another mannanase, a pectin methylesterase, a cellobiohydrolase, a transglutaminase, and mixtures thereof.

- [19] A cleaning agent composition including the mutant alkaline cellulase according to the above items [1] to [11], the anti-redeposition agent according to the above item [16], or the enzyme composition according to the above item [17] or [18].
- [20] A method for producing a mutant alkaline cellulase, the method including expressing the mutant alkaline cellulase from the gene according to the above item [12].
- [21] A method for enhancing the anti-redeposition ability of an alkaline cellulase, by substituting at least one amino acid residue selected from the amino acid residues at the positions corresponding to positions 58, 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, 175, 176, 178, 179, 181, 193, 194, 195, 196, 197, 199, 202, 203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 312, 318, 324, 345, 354, 356, 357, 360, 363, 368, 418, 419, 420, 421, 422, 453, 454, 455, 457, 458, 459, 494, 495, 496, 500, 501, 502, 503, 504, 550, 551, 552, 604, 605, 606, 607, 608, 640, 641, 642, 644, 645, 646, 683, 684, 685, 690, 691, 692, 693, 694, 739, 740, and 741 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of the amino acid sequence of an alkaline cellulase having at least 70% identity with the amino acid sequence of SEQ ID NO: 2, with other amino acid residue.
- [22] The method according to the above item [21], wherein the at least one amino acid residue is a glutamine residue at the position according to the position 58 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of the amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2.
- [23] The method according to the above item [21], wherein the at least one amino acid residue is selected from non-charged amino acid residues at the positions corresponding to positions 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, 175, 176, 178, 179, 181, 193, 194, 195, 196, 197, 199, 202, 203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 312, 318, 324, 345, 354, 356, 357, 360, 363, and 368 of the amino acid sequence set forth in SEQ ID NO: 2.
- [24] The method according to the above item [21], wherein the at least one amino acid residue is selected from the amino acid residues at the positions corresponding to positions 418, 419, 420, 421, 422, 453, 454, 455, 457, 458, 459, 494, 495, 496, 500, 501, 502, 503, 504, 550, 551, 552, 604, 605, 606, 607, 608, 640, 641, 642, 644, 645, 646, 683, 684, 685, 690, 691, 692, 693, 694, 739, 740, and 741 of the amino acid sequence set forth in SEQ ID NO: 2.
- [25] A method for enhancing the anti-redeposition ability of an alkaline cellulase, or method for both enhancing the anti-redeposition ability and protease resistance of an alkaline cellulase, wherein a glutamine residue at the position corresponding to position 58 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of the amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2, is substituted with glutamic acid or arginine.

BRIEF DESCRIPTION OF DRAWINGS

FIG. 1 is a schematic diagram of (A) the structure of S237 cellulase, and (B) a speculated mechanism of the effect of preventing redeposition by a cellulase. The signal sequence illustrated in FIG. 1A is usually cut and removed in a mature protein of S237 cellulase.

FIG. 2 is a diagram illustrating the effect for promoting preventing of redeposition provided by the addition of S237 cellulase in comparison with other enzymes. FIG. 2A illustrates the results of comparing S237 cellulase with other hydrolases, and FIG. 2B illustrates the results of comparing S237 cellulase with other cellulases.

FIG. 3 is a diagram illustrating the pH dependency of the anti-redeposition effect provided by the addition of S237 cellulase.

FIG. 4 is a diagram illustrating the results of a comparison of the residual activity (when the activity of $S237_Q242S$ (QS) is 100) of a cellulase in the presence of various proteases.

DESCRIPTION OF EMBODIMENTS

The details of the mechanism for prevention of redeposition by cellulases are not clearly known, and in regard to a method for obtaining a cellulase having higher anti-redeposition ability is not yet known. Furthermore, a problem of cellulases being susceptible to degradation by co-existing proteases is known. Thus, further improvement is needed.

The present invention relates to provision of a mutant alkaline cellulase having enhanced anti-redeposition ability, and a 25 mutant alkaline cellulase having enhanced anti-redeposition ability and protease resistance.

The inventors of the present invention conducted a thorough investigation in order to address the problems described above, and as a result, the inventors found that when an amino acid residue at a predetermined position in a *Bacillus* sp. strain KSM-S237-derived alkaline cellulase is substituted with other amino acid residue, the anti-redeposition ability can be enhanced, and in some mutant alkaline cellulases, both anti-redeposition ability and protease resistance can be enhanced.

Furthermore, the inventors of the present invention found that when a non-charged amino acid residue at a predetermined position exposed to the enzyme surface in the catalytic domain of an alkaline cellulase is substituted with a charged amino acid residue so as to increase hydrophilicity of the enzyme surface of the relevant alkaline cellulase, the antiredeposition ability of the alkaline cellulase can be remarkably enhanced.

Furthermore, the inventors of the present invention found that when an amino acid residue in a region that participates in cellulose binding or the vicinity thereof in the cellulose binding module of an alkaline cellulase is substituted so as to weaken the cellulose binding properties, the anti-redeposition ability of the alkaline cellulase can be further enhanced.

The mutant alkaline cellulase having enhanced anti-redeposition ability according to the present invention, and an anti-redeposition agent containing the mutant alkaline cellulase can bring about a high anti-redeposition promoting effect 55 in, for example, a washing process. The enzyme composition according to the present invention can provide an anti-redeposition effect to a cleaning agent composition and the like when incorporated into the cleaning agent composition and the like. The cleaning agent composition according to the 60 present invention can exhibit a high anti-redeposition effect at the time of washing. When the method for producing a mutant alkaline cellulase according to the present invention is used, production of an alkaline cellulase having enhanced antiredeposition ability is possible. Furthermore, according to the 65 present invention, an alkaline cellulase which has a high anti-redeposition activity, has both a high anti-redeposition

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activity and high protease resistance, and is useful as an enzyme for incorporation into a cleaning agent, can be provided.

Hereinafter, the present invention will be described in detail.

1. Mutant Alkaline Cellulase and Production Thereof

In the mutant alkaline cellulase of the present invention, at least one amino acid residue selected from the amino acid residues at the positions corresponding to positions 58, 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, 175, 176, 178, 179, 181, 193, 194, 195, 196, 197, 199, 202, 203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 312, 318, 324, 345, 354, 356, 357, 360, 363, 368, 418, 419, 420, 421, 422, 453, 454, 455, 457, 15 458, 459, 494, 495, 496, 500, 501, 502, 503, 504, 550, 551, 552, 604, 605, 606, 607, 608, 640, 641, 642, 644, 645, 646, 683, 684, 685, 690, 691, 692, 693, 694, 739, 740, and 741 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of the amino acid sequence of an alkaline cellulase having at least 70% identity with the amino acid sequence of SEQ ID NO: 2, is substituted with other amino acid residue. The mutant alkaline cellulase of the present invention has enhanced anti-redeposition ability, or has both enhanced anti-redeposition ability and protease resistance, as compared with the alkaline cellulase before mutation (parent alkaline cellulase).

One embodiment of the present invention is a mutant alkaline cellulase which acquires enhanced anti-redeposition ability, or acquires enhanced anti-redeposition ability and protease resistance, by substituting a glutamine residue at the position corresponding to position 58 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of the amino acid sequence of an alkaline cellulase having at least 90% identity with the amino acid sequence of SEQ ID NO: 2, with other amino acid.

More specifically, the mutant alkaline cellulase of the present invention acquires enhanced anti-redeposition ability, or acquires both enhanced anti-redeposition ability and protease resistance, by substituting a glutamine residue at the position corresponding to position 58 of the amino acid sequence set forth in SEQ ID NO: 2, with glutamic acid or arginine residue.

Another embodiment of the present invention is a mutant alkaline cellulase which acquires enhanced anti-redeposition ability by substituting a non-charged amino acid residue at a particular position in the catalytic domain of any parent alkaline cellulase, with a charged amino acid residue.

More specifically, the mutant alkaline cellulase according to the present invention is a mutant alkaline cellulase obtainable by introducing an amino acid substitution into the amino acid sequence of an available alkaline cellulase, which may be an alkaline cellulase derived from a bacterium of the genus *Bacillus*, such as *Bacillus* sp. strain KSM-S237, or an alkaline cellulase having a high amino acid sequence identity with the foregoing alkaline cellulase, which acquires enhanced antiredeposition ability by substituting a non-charged amino acid residue that is particularly selected among the non-charged amino acid residues exposed to the enzyme surface in the catalytic domain of the alkaline cellulase with a charged amino acid residue.

Even more specifically, the mutant alkaline cellulase according to the present invention acquires enhanced antiredeposition ability, by substituting at least one amino acid residue selected from the non-charged amino acid residues at the positions corresponding to positions 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, 175, 176, 178, 179, 181, 193, 194, 195, 196, 197, 199, 202,

203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 312, 318, 324, 345, 354, 356, 357, 360, 363, and 368 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of an amino acid of an alkaline cellulase having at least 70% identity with the amino acid sequence of 5 SEQ ID NO: 2, with a charged amino acid residue.

Another embodiment of the present invention is a mutant alkaline cellulase which has enhanced anti-redeposition ability, by substituting an amino acid residue in the region that directly participates in cellulose binding, or a region adjacent thereto, in the cellulose binding module of an available parent alkaline cellulase.

More specifically, the mutant alkaline cellulase according to the present invention is a mutant alkaline cellulase obtainable by introducing an amino acid substitution into the amino 15 acid sequence of an available alkaline cellulase, which may be an alkaline cellulase derived from a bacterium of the genus Bacillus, such as Bacillus sp. strain KSM-S237, or an alkaline cellulase having high identity of amino acid sequence with that of the foregoing alkaline cellulase, which acquires 20 enhanced anti-redeposition ability by substituting an amino acid residue in the region that directly participates in cellulose binding or a region in the vicinity thereof in the cellulose binding module of the alkaline cellulase, with other amino acid residue.

Even more specifically, the mutant alkaline cellulase according to the present invention acquires enhanced antiredeposition ability by substituting at least one amino acid residue selected from the amino acid residues at the positions corresponding to positions 418, 419, 420, 421, 422, 453, 454, 30 455, 457, 458, 459, 494, 495, 496, 500, 501, 502, 503, 504, 550, 551, 552, 604, 605, 606, 607, 608, 640, 641, 642, 644, 645, 646, 683, 684, 685, 690, 691, 692, 693, 694, 739, 740, and 741 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of the amino acid sequence of an 35 alkaline cellulase having at least 70% identity with the amino acid sequence of SEQ ID NO: 2, with other amino acid

In the present specification, an original alkaline cellulase, stitution, is referred to as a "parent alkaline cellulase", the amino acid sequence thereof is referred to as a "parent amino acid sequence", and the gene encoding the alkaline cellulase is referred to as a "parent alkaline cellulase gene".

The term amino acid residue as used in the present inven- 45 tion means twenty kinds of amino acid residues constituting proteins, namely, Ala, Arg, Asn, Asp, Cys, Gln, Glu, Gly, His, Ile, Leu, Lys, Met, Phe, Pro, Ser, Thr, Trp, Tyr and Val.

The term non-charged amino acid as used in the present invention means amino acids that do not have a charge among 50 the twenty kinds of amino acids constituting proteins, that is, non-polar amino acids (valine, alanine, leucine, isoleucine, methionine, phenylalanine, tryptophan, and proline) and polar amino acids (glycine, asparagine, cysteine, glutamine, serine, threonine, and tyrosine). The non-charged amino acid 55 residue that may be subjected to an amino acid substitution in the present invention may be any non-polar amino acid or polar amino acid, and the non-charged amino acid residue is more preferably alanine, serine, glutamine or asparagine.

Furthermore, the term charged amino acid as used in the 60 present invention means amino acids having a charge among the twenty amino acids constituting proteins, that is, acidic amino acids (aspartic acid and glutamic acid), and basic amino acids (arginine, histidine and lysine). The charged amino acid residue that is introduced into the mutant alkaline 65 cellulase by substituting a non-charged amino acid residue in the present invention may be any acidic amino acid or basic

amino acid, and the charged amino acid residue is preferably glutamic acid, aspartic acid or arginine.

According to the present invention, the identity between amino acid sequences refers to the proportion (%) of the number of positions at which amino acid residues are identical between two aligned amino acid sequences, relative to the number of full-length amino acid residues. Specifically, the identity is calculated by the Lipman-Pearson method; Science, 227, 1435 (1985)), and can be computed by performing an analysis using a homology analysis program (Search homology) of the genetic information processing software, Genetyx-Win (Ver. 5.1.1; Software Development, Inc.) by setting the unit size to compare (ktup) at 2.

The amino acid sequence of the mutant alkaline cellulase according to the present invention can be designed by using the amino acid sequence of any available alkaline cellulase as a parent. Examples of such a parent amino acid sequence include the amino acid sequences of the various alkaline cellulases belonging to family 5 of the glycoside hydrolase family. Furthermore, examples of the parent amino acid sequence include the amino acid sequences of alkaline cellulases derived from any available organisms producing alkaline cellulases, such as bacteria of the genus Bacillus, bacteria of the genus Clostridium, and bacteria of the genus Acidothermus. Among these, an amino acid sequence of an alkaline cellulase derived from a bacterium of the genus Bacillus can be favorably used.

A suitable example of the alkaline cellulase of a bacterium of the genus Bacillus includes Bacillus sp. strain KSM-S237derived alkaline cellulase [S237 cellulase]. The *Bacillus* sp. strain KSM-S237-derived alkaline cellulase, as a precursor protein containing an N-terminal signal sequence (positions 1 to 30 of SEQ ID NO: 2; MMLRKKTKQLISSILILV-LLLSLFPAALAA [one-character notation of amino acids]), has the amino acid sequence set forth in SEQ ID NO: 2, and as a mature protein has an amino acid sequence obtained by removing the signal sequence from the amino acid sequence of SEQ ID NO: 2.

Accordingly, examples of the parent amino acid sequence that is, an available alkaline cellulase before amino acid sub- 40 of the mutant alkaline cellulase according to the present invention include the amino acid sequence set forth in SEQ ID NO: 2, as well as amino acid sequences having at least 70% identity, preferably at least 80% identity, more preferably at least 90% identity, even more preferably at least 95% identity, even more preferably at least 96% identity, and even more preferably at least 98% identity, with the amino acid sequence set forth in SEO ID NO: 2. These amino acid sequences may be amino acid sequences encoded in the open reading frame (ORF), or may be amino acid sequences obtained by removing the signal sequence from the foregoing amino acid sequences.

> Examples of the alkaline cellulase of a bacterium of the genus Bacillus other than S237 cellulase include alkaline cellulases derived from various Bacillus bacterial strains, such as Bacillus sp. strain DSM12648-derived alkaline cellulase (SEQ ID NO: 4) [the identity of amino acid sequence with that of S237 cellulase (including the signal sequence) is about 98.2%], Bacillus sp. strain 1139-derived alkaline cellulase (SEQ ID NO: 6) [the identity of amino acid sequence with that of S237 cellulase (including the signal sequence) is about 91.3%], Bacillus sp. strain KSM-64-derived alkaline cellulase (SEQ ID NO: 8) [the identity of amino acid sequence with that of S237 cellulase (including the signal sequence) is about 91.6%], Bacillus sp. strain KSM-635derived alkaline cellulase (SEQ ID NO: 10) [the identity of amino acid sequence with that of S237 cellulase (including the signal sequence) is about 71.0%], and Bacillus sp. strain

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N-4-derived alkaline cellulase (SEQ ID NO: 12) [the identity of amino acid sequence with that of S237 cellulase (including the signal sequence) is about 64.0%], as well as two kinds of alkaline cellulases such as N131a and N131b derived from *Bacillus* sp. strain KSM-N131 (JP-A No. 2001-231569) [the 5 identity of amino acid sequence with that of S237 cellulase (including the signal sequence) is 87.9% and 97.1%, respectively]. These alkaline cellulases have high mutual amino acid sequence identity and similarity.

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Therefore, examples of the parent amino acid sequence of the mutant alkaline cellulase according to the present invention include the amino acid sequences of *Bacillus* sp. strain DSM12648-derived alkaline cellulase (SEQ ID NO: 4), *Bacillus* sp. strain 1139-derived alkaline cellulase (SEQ ID NO: 6), *Bacillus* sp. strain KSM-64-derived alkaline cellulase (SEQ ID NO: 8), *Bacillus* sp. strain KSM-635-derived alkaline cellulase (SEQ ID NO: 10), and *Bacillus* sp. strain N-4-derived alkaline cellulase (SEQ ID NO: 12), as well as amino acid sequences having at least 90% identity, preferably at least 95% identity, and 20 even more preferably at least 98% identity, with the aforementioned amino acid sequences.

Alternatively, when the mutant alkaline cellulase according to the present invention is a variant in which the amino acid residue at the position corresponding to position 58 has 25 been substituted (58th position variant), examples of the parent amino acid sequence of the variant include the amino acid sequence of *Bacillus* sp. strain KSM-S237-derived alkaline cellulase as set forth in SEQ ID NO: 2, as well as amino acid sequences having at least 90% identity, more preferably at least 95% identity, even more preferably at least 96% identity, and even more preferably at least 98% identity, with the amino acid sequence set forth in SEQ ID NO: 2.

Examples of a parent alkaline cellulase derived from a bacterium of the genus *Bacillus* other than S237 cellulase, 35 which has an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2, include the alkaline cellulases having the amino acid sequences set forth in SEQ ID NO: 4, SEQ ID NO: 6 and SEQ ID NO: 8, and the alkaline cellulases derived from various *Bacillus* bacterial 40 strains, such as *Bacillus* sp. strain KSM-N131-derived alkaline cellulase N131b.

The parent amino acid sequence of position 58 variant may be an amino acid sequence encoded in the open reading frame (ORF), or may also be an amino acid sequence having the 45 signal sequence (positions 1 to 30 of SEQ ID NO: 2) removed from the foregoing amino acid sequence.

Furthermore, the parent amino acid sequence according to the present invention may also include amino acid sequences having a deletion, substitution or addition of one or several 50 amino acids with respect to each of the amino acid sequences of SEQ ID NOs: 2, 4, 6, 8, 10 and 12. The "one or several" refers to, hereinafter for example, 1 to 50 amino acids, preferably 1 to 30 amino acids, more preferably 1 to 20 amino acids, even more preferably 1 to 10 amino acids, and even 55 more preferably 1 to 5 amino acids. Examples of such a parent amino acid sequence include, but are not limited to, the amino acid sequences of a mutant alkaline cellulase in which the amino acid residue at the position corresponding to position 10 of the amino acid sequence set forth in SEQ ID NO: 2 has 60 been substituted with glutamine, alanine, proline or methionine; a mutant alkaline cellulase in which the amino acid residue at the position corresponding to position 16 has been substituted with asparagine or arginine; a mutant alkaline cellulase in which the amino acid residue at the position 65 corresponding to position 22 has been substituted with proline; a mutant alkaline cellulase in which the amino acid

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residue at the position corresponding to position 33 has been substituted with histidine; a mutant alkaline cellulase in which the amino acid residue at the position corresponding to position 39 has been substituted with alanine, threonine or tyrosine; a mutant alkaline cellulase in which the amino acid residue at the position corresponding to position 76 has been substituted with histidine, methionine, valine, threonine or alanine; a mutant alkaline cellulase in which the amino acid residue at the position corresponding to position 109 has been substituted with isoleucine, leucine, serine or valine; a mutant alkaline cellulase in which the amino acid residue at the position corresponding to position 242 has been substituted with alanine, phenylalanine, valine, serine, aspartic acid, glutamic acid, leucine, isoleucine, tyrosine, threonine, methionine or glycine; a mutant alkaline cellulase in which the amino acid residue at the position corresponding to position 263 has been substituted with isoleucine, leucine, proline or valine; a mutant alkaline cellulase in which the amino acid residue at the position corresponding to position 308 has been substituted with alanine, serine, glycine or valine; a mutant alkaline cellulase in which the amino acid residue at the position corresponding to position 462 has been substituted with threonine, leucine, phenylalanine or arginine; a mutant alkaline cellulase in which the amino acid residue at the position corresponding to position 466 has been substituted with leucine, alanine or serine; a mutant alkaline cellulase in which the amino acid residue at the position corresponding to position 468 has been substituted with alanine, aspartic acid, glycine or lysine; a mutant alkaline cellulase in which the amino acid residue at the position corresponding to position 552 has been substituted with methionine; a mutant alkaline cellulase in which the amino acid residue at the position corresponding to position 564 has been substituted with valine, threonine or leucine; a mutant alkaline cellulase in which the amino acid residue at the position corresponding to position 608 has been substituted with isoleucine or arginine; and the mutant alkaline cellulases disclosed in Patent Documents 2 and 3, such as mutant alkaline cellulases having combinations of one or more of the substitutions of amino acid residues described above. Among the amino acid sequences described above, preferred examples include the amino acid sequences of alkaline cellulases having a mutation in which the amino acid residue at position 242 of the amino acid sequence set forth in SEQ ID NO: 2 has been substituted with other amino acid residues. Amore preferred example includes the amino acid sequence of an alkaline cellulase having a mutation in which the glutamine residue at position 242 of the amino acid sequence set forth in SEQ ID NO: 2 has been substituted with a serine residue.

According to the present invention, in the parent amino acid sequences such as described above, one or more amino acid residues selected from the amino acid residues at the positions corresponding to positions 58, 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, $175,\,176,\,178,\,179,\,181,\,193,\,194,\,195,\,196,\,197,\,199,\,202,$ 203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 312, 318, 324, 345, 354, 356, 357, 360, 363, 368, 418, 419, 420, 421, 422, 453, 454, 455, 457, 458, 459, 494, 495, 496, 500, 501, 502, 503, 504, 550, 551, 552, 604, 605, 606, 607, 608, 640, 641, 642, 644, 645, 646, 683, 684, 685, 690, 691, 692, 693, 694, 739, 740, and 741 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of the amino acid sequence of an alkaline cellulase having at least 70% identity with the amino acid sequence of SEQ ID NO: 2, can be substituted with other amino acid residues.

According to an exemplary embodiment of the present invention, in the parent amino acid sequences such as

described above, the glutamine residue at the position corresponding to position 58 of the amino acid sequence SEQ ID NO: 2 is substituted with other amino acid.

Preferably, the glutamine residue at the position corresponding to position 58 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of the amino acid sequence of an alkaline cellulase having an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2, may be substituted with glutamic acid or arginine residue.

Furthermore, preferably, the mutant alkaline cellulase of the present invention may have an amino acid sequence in which the glutamine residue at the position corresponding to position 58 of the amino acid sequence set forth in SEQ ID NO: 2 in the amino acid sequence of an alkaline cellulase having a substitution of one or several amino acid residues of the amino acid sequence set forth in SEQ ID NO: 2 by other amino acid residues, is substituted with other amino acid residue. For example, the mutant alkaline cellulase of the 20 present invention may have an amino acid sequence in which the glutamine residue at the position corresponding to position 58 of the amino acid sequence set forth in SEQ ID NO: 2 in the amino acid sequence of a mutant alkaline cellulase having a mutation of substitution of 1 to 20 amino acid resi- 25 dues, preferably 1 to 10 amino acid residues, and more preferably 1 to 5 amino acid residues, by other amino acid residues with respect to the amino acid residues of an alkaline cellulase having the amino acid sequence set forth in SEQ ID NO: 2, is substituted with glutamic acid or arginine residue. 30

More preferably, the mutant alkaline cellulase of the present invention may have an amino acid sequence in which the glutamine residue at the position corresponding to position 58 of the amino acid sequence set forth in SEQ ID NO: 2 in the amino acid sequence of a mutant alkaline cellulase 35 having a mutation of substitution of the amino acid residue at position 242 in an alkaline cellulase having the amino acid sequence set forth in SEQ ID NO: 2 by other amino acid residue, is substituted with other amino acid residue.

Even more preferably, the mutant alkaline cellulase of the 40 present invention may have an amino acid sequence in which the glutamine residue at position 58 of the amino acid sequence set forth in SEQ ID NO: 2 in the amino acid sequence of a mutant alkaline cellulase having a mutation of substitution of the glutamine residue at position 242 of the 45 amino acid sequence set forth in SEQ ID NO: 2 by a serine residue, is substituted with glutamic acid or arginine.

According to another exemplary embodiment of the present invention, in the parent amino acid sequences such as described above, one or more amino acid residues selected 50 from the non-charged amino acid residues at the positions corresponding to positions 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, 175, 176, 178, 179, 181, 193, 194, 195, 196, 197, 199, 202, 203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 55 312, 318, 324, 345, 354, 356, 357, 360, 363, and 368 of the amino acid sequence set forth in SEQ ID NO: 2, may be subjected to a substitution by charged amino acid residues.

Preferably, one or more amino acid residues selected from the non-charged amino acid residues at the positions corresponding to positions 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, 175, 176, 178, 179, 181, 193, 194, 195, 196, 197, 199, 202, 203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 312, 318, 324, 345, 354, 356, 357, 360, 363, and 368 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of the amino acid sequence of an alkaline cellulase having at

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least 70% identity with the amino acid sequence of SEQ ID NO: 2, may be subjected to a substitution by charged amino acid residues.

Furthermore, preferably, the mutant alkaline cellulase of the present invention may have an amino acid sequence in which one or more amino acid residues selected from the non-charged amino acid residues at the positions corresponding to positions 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, 175, 176, 178, 179, 181, 193, 194, 195, 196, 197, 199, 202, 203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 312, 318, 324, 345, 354, 356, 357, 360, 363, and 368 of the amino acid sequence set forth in SEQ ID NO: 2 in the amino acid sequence of a mutant alkaline cellulase having a substitution of one or several amino acid residues by other amino acid residues with respect to an alkaline cellulase having the amino acid sequence set forth in SEQ ID NO: 2, are substituted with charged amino acid residues. For example, the mutant alkaline cellulase of the present invention may have an amino acid sequence in which one or more amino acid residues selected from the non-charged amino acid residues at the positions corresponding to positions 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, 175, 176, 178, 179, 181, 193, 194, 195, 196, 197, 199, 202, 203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 312, 318, 324, 345, 354, 356, 357, 360, 363, and 368 of the amino acid sequence set forth in SEQ ID NO: 2 in the amino acid sequence of a mutant alkaline cellulase having a mutation of substitution of 1 to 20 amino acid residues, preferably 1 to 10 amino acid residues, and more preferably 1 to 5 amino acid residues, by other amino acids with respect to the amino acid residues of an alkaline cellulase having the amino acid sequence set forth in SEQ ID NO: 2, are substituted with charged amino acid residues.

More preferably, the mutant alkaline cellulase of the present invention may have an amino acid sequence in which one or more amino acid residues selected from the non-charged amino acid residues at the positions corresponding to positions 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, 175, 176, 178, 179, 181, 193, 194, 195, 196, 197, 199, 202, 203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 312, 318, 324, 345, 354, 356, 357, 360, 363, and 368 of the amino acid sequence set forth in SEQ ID NO: 2 in the amino acid sequence of a mutant alkaline cellulase having a mutation of substitution of the amino acid residue at position 242 in an alkaline cellulase having the amino acid sequence set forth in SEQ ID NO: 2 by other amino acid residue, are substituted with charged amino acid residues.

Even more preferably, the mutant alkaline cellulase of the present invention may have an amino acid sequence in which one or more amino acid residues selected from the non-charged amino acid residues at the positions 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, 175, 176, 178, 179, 181, 193, 194, 195, 196, 197, 199, 202, 203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 312, 318, 324, 345, 354, 356, 357, 360, 363, and 368 of the amino acid sequence set forth in SEQ ID NO: 2 in the amino acid sequence of a mutant alkaline cellulase having a mutation of substitution of the glutamine residue at position 242 of the amino acid sequence set forth in SEQ ID NO: 2 by a serine residue, are substituted with charged amino acid residues.

According to another exemplary embodiment of the present invention, the amino acid residues in the region that directly participates in cellulose binding or a region adjacent

thereto in the cellulose binding module in a parent amino acid sequence such as described above, are subjected to substitu-

Preferably, one or more of the amino acid residues at the positions corresponding to positions 418, 419, 420, 421, 422, 453, 454, 455, 457, 458, 459, 494, 495, 496, 500, 501, 502, 503, 504, 550, 551, 552, 604, 605, 606, 607, 608, 640, 641, 642, 644, 645, 646, 683, 684, 685, 690, 691, 692, 693, 694, 739, 740, and 741 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of the amino acid sequence of an alkaline cellulase having at least 70% identity with the amino acid sequence of SEQ ID NO: 2, may be subjected to amino acid substitution.

Furthermore, preferably, the mutant alkaline cellulase of 15 the present invention may have an amino acid sequence in which one or more amino acid residues selected from the amino acid residues at the positions corresponding to positions 418, 419, 420, 421, 422, 453, 454, 455, 457, 458, 459, 494, 495, 496, 500, 501, 502, 503, 504, 550, 551, 552, 604, 20 605, 606, 607, 608, 640, 641, 642, 644, 645, 646, 683, 684, 685, 690, 691, 692, 693, 694, 739, 740, and 741 of the amino acid sequence set forth in SEQ ID NO: 2 in the amino acid sequence of a mutant alkaline cellulase having a substitution of one or several amino acid residues by other amino acid 25 residues with respect to the amino acid residues of an alkaline cellulase having the amino acid sequence set forth in SEQ ID NO: 2, are substituted with other amino acid residues. For example, the mutant alkaline cellulase of the present invention may have an amino acid sequence in which one or more amino acid residues selected from the amino acid residues at the positions corresponding to positions 418, 419, 420, 421, 422, 453, 454, 455, 457, 458, 459, 494, 495, 496, 500, 501, 502, 503, 504, 550, 551, 552, 604, 605, 606, 607, 608, 640, 641, 642, 644, 645, 646, 683, 684, 685, 690, 691, 692, 693, 694, 739, 740, and 741 of the amino acid sequence set forth in SEO ID NO: 2 in the amino acid sequence of a mutant alkaline cellulase having a mutation of substitution of 1 to 20 and more preferably 1 to 5 amino acid residues, by other amino acids with respect to the amino acid residues of an alkaline cellulase having the amino acid sequence set forth in SEQID NO: 2, are substituted with other amino acid residues.

More preferably, the mutant alkaline cellulase of the 45 present invention may have an amino acid sequence in which one or more amino acid residues selected from the amino acid residues at the positions corresponding to positions 418, 419, 420, 421, 422, 453, 454, 455, 457, 458, 459, 494, 495, 496, 500, 501, 502, 503, 504, 550, 551, 552, 604, 605, 606, 607, 50 608, 640, 641, 642, 644, 645, 646, 683, 684, 685, 690, 691, 692, 693, 694, 739, 740, and 741 of the amino acid sequence set forth in SEQ ID NO: 2 in the amino acid sequence of a mutant alkaline cellulase having a mutation of substitution of the amino acid residue at position 242 of the amino acid 55 sequence set forth in SEQ ID NO: 2 by other amino acid residue, are substituted with other amino acid residues.

Even more preferably, the mutant alkaline cellulase of the present invention may have an amino acid sequence in which one or more amino acid residues selected from the amino acid 60 residues at positions 418, 419, 420, 421, 422, 453, 454, 455, 457, 458, 459, 494, 495, 496, 500, 501, 502, 503, 504, 550, 551, 552, 604, 605, 606, 607, 608, 640, 641, 642, 644, 645, 646, 683, 684, 685, 690, 691, 692, 693, 694, 739, 740, and 741 of the amino acid sequence set forth in SEQ ID NO: 2 in the amino acid sequence of a mutant alkaline cellulase having a mutation of substitution of the glutamine residue at position

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242 of the amino acid sequence set forth in SEQ ID NO: 2 by a serine residue, are substituted with other amino acid resi-

The phrase "the position corresponding to position "Y" of the amino acid sequence set forth in SEO ID NO: "X"" is used to assign a predetermined position of amino acid residue in the amino acid sequence of an available alkaline cellulase when the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: "X" (for example, the amino acid sequence of S237 cellulase set forth in SEQ ID NO: 2) is used as a reference sequence. For example, the "amino acid residue at the position corresponding to position "Y" of the amino acid sequence set forth in SEQ ID NO: "X"" means the amino acid residue appearing in the Y-th position counted from the first amino acid residue of the amino acid sequence set forth in SEQ ID NO: "X".

On the other hand, in the case of the amino acid sequence ("Z") of an alkaline cellulase other than SEQ ID NO: "X", the "amino acid residue at the position corresponding to position "Y" of the amino acid sequence set forth in SEQ ID NO: "X"" means the amino acid residue in the amino acid sequence "Z", which is aligned with the Y-th amino acid residue counted from the 1st amino acid residue of the amino acid sequence of SEQ ID NO: "X" when the amino acid sequence "Z" is aligned with the amino acid sequence of SEQ ID NO: "X" (that is, aligned in the same column with respect to the alignment). Furthermore, the alignment of the amino acid sequence of SEQ ID NO: "X" and other amino acid sequence may be carried out manually, or the alignment can also be obtained by, for example, using Clustal W multiple alignment program (Thompson, J. D. et al., (1994) Nucleic Acids Res. 22, p. 4673-4680) at a default setting. Clustal W can be used at, for example, the websites of European Bioinformatics Institute (EBI, http://www.ebi.ac.uk/index.html) or the DNA Data Bank of Japan operated by the National Institute of Genetics in Japan (DDBJ, http://www.ddbj.nig.ac.jp/Welcome-j.html).

A person ordinary skilled in the art can further adjust the amino acid residues, preferably 1 to 10 amino acid residues, 40 alignment thus obtained, if necessary, so as to obtain the optimal alignment. It is preferable to determine such an optimal alignment by taking into consideration of the similarity of amino acid sequences or the frequency of the gap that is inserted. Hereinafter, the similarity of amino acid sequences refers to the proportion (%) of the number of positions at which identical or similar amino acid residues are present in both of two amino acid sequences when the amino acid sequences are aligned, relative to the number of full-length amino acid residues. The term similar amino acid residue means an amino acid residue having properties that are similar to each other in terms of polarity or charge and used for so-called conservative substitution, among the twenty kinds of amino acids constituting proteins. Groups composed of such similar amino acid residues are well known to those ordinary skilled in the art, and examples include, but not limited to, arginine and lysine; glutamic acid and aspartic acid; serine and threonine; glutamine and asparagine; and valine, leucine and isoleucine.

When alignment is carried out according to the method described above, the glutamine residue at position 29 in the sequence of SEQ ID NO: 4 (derived from strain DSM12648), the glutamine residue at position 58 in the sequence of SEQ ID NO: 6 (derived from strain 1139), and the glutamine residue at position 57 in the sequence of SEQ ID NO: 8 (derived from strain KSM-64) respectively correspond to the "amino acid residue at the position corresponding to position 58 of the amino acid sequence set forth in SEQ ID NO: 2".

Furthermore, as a reference, the amino acid residues respectively corresponding to the positions described above of the sequence of SEQ ID NO: 2 in the respective amino acid sequences of SEQ ID NO: 2 (derived from *Bacillus* sp. strain KSM-S237), SEQ ID NO: 4 (derived from strain 5 DSM12648), SEQ ID NO: 6 (derived from strain 1139), SEQ ID NO: 8 (derived from strain KSM-64), SEQ ID NO: 10 (derived from strain KSM-635), and SEQ ID NO: 12 (derived from strain N-4), which can be used as the parent amino acid sequences, are shown in Table 5 and Table 6 illustrated below.

According to a preferred exemplary embodiment, the mutant alkaline cellulase according to the present invention includes, for example, a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting the glutamine residue at the position corresponding to position 58 15 of the amino acid sequence of SEQ ID NO: 2 and of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2, with glutamic acid or arginine residue; and having an amino acid sequence that is obtained by further removing a signal sequence (correspond- 20 ing to a sequence including the positions 1 to 30 of SEQ ID NO: 2) from the foregoing sequence. The substitution described above provides remarkably high anti-redeposition ability, or provides both remarkably high anti-redeposition ability and high protease resistance, to the mutant alkaline 25 cellulase.

A preferred example of the mutant alkaline cellulase of the present invention according to this exemplary embodiment includes a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting the glutamine residue at position 58 of the amino acid sequence set forth in SEQ ID NO: 2 with glutamic acid and arginine; and having an amino acid sequence that is obtained by further removing the signal sequence from the foregoing sequence.

Another preferred example includes a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting the glutamine residue at position 58 in the amino acid sequence of a mutant alkaline cellulase having a mutation of substitution of the glutamine residue at position 242 of the amino acid sequence set forth in SEQ ID NO: 2 by a serine 40 residue, with glutamic acid or arginine residue; and having an amino acid sequence that is obtained by further removing the signal sequence from the foregoing sequence.

Another preferred example includes a mutant alkaline cellulase having an amino acid sequence that is obtained by 45 substituting the glutamine residue at position 29 of the amino acid sequence of SEQ ID NO: 4 in the amino acid sequence set forth in SEQ ID NO: 4, with glutamic acid or arginine residue; and having an amino acid sequence that is obtained by further removing the signal sequence from the foregoing 50 sequence.

Another preferred example includes a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting the glutamine residue at position 58 of the amino acid sequence of SEQ ID NO: 4 in the amino acid sequence 55 set forth in SEQ ID NO: 6, with glutamic acid or arginine residue; and an amino acid sequence that is obtainable by further removing the signal sequence from the foregoing sequence.

Another preferred example includes a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting the glutamine residue at position 57 of the amino acid sequence of SEQ ID NO: 4 in the amino acid sequence set forth in SEQ ID NO: 8, with glutamic acid or arginine residue; and having an amino acid sequence that is obtained 65 by further removing the signal sequence from the foregoing sequence.

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According to another preferred exemplary embodiment, the mutant alkaline cellulase related to the present invention includes, for example, in a mutant alkaline cellulase having the amino acid sequence set forth in SEQ ID NO: 2 or an amino acid sequence having at least 70% identity with the amino acid sequence of SEQ ID NO: 2, the mutant alkaline cellulase having an amino acid sequence that is obtained by substituting a non-charged amino acid residue at a particular position in the catalytic domain with a charged amino acid residue, and having an amino acid sequence that is obtained by further removing a signal sequence (corresponding to a sequence including the positions 1 to 30 of SEQ ID NO: 2) from the foregoing sequence.

The mutant alkaline cellulase according to the above exemplary embodiment exhibits anti-redeposition ability based on the high hydrophilicity of the enzyme surface and the overall structure of the enzyme. Therefore, in the case of using the mutant alkaline cellulase simply for the purpose of obtaining an anti-redeposition effect, it makes no difference whether the mutant alkaline cellulase has cellulase activity or not. However, from the viewpoint that a glycolytic activity against saccharides including cellulose can also be simultaneously obtained, it is more preferable that the mutant alkaline cellulase related to the present invention has cellulase activity. The relevant mutant alkaline cellulase may have a signal sequence at the N-terminal, or may be in the form of a mature protein having the signal sequence removed therefrom.

In the mutant alkaline cellulase according to the exemplary embodiment described above, the substitution of a non-charged amino acid residue in the catalytic domain with a charged amino acid residue is not limited, and for example, the substitution may be a substitution of alanine, serine, glutamine or asparagine with glutamic acid, aspartic acid or arginine. Such a substitution provides remarkably high anti-redeposition ability to the mutant alkaline cellulase according to the present invention.

A preferred example of the mutant alkaline cellulase in this exemplary embodiment may be a mutant alkaline cellulase in which the non-charged amino acid residue at position 71 in the amino acid sequence set forth in SEQ ID NO: 2 (this is the position of an amino acid residue adjacent to an acidic amino acid) is substituted with an acidic amino acid residue. Such a substitution particularly efficiently increases the hydrophilicity in the vicinity of the substituted amino acid residue, and provides remarkably high anti-redeposition ability to the resulting mutant alkaline cellulase.

Another preferred example may be a mutant alkaline cellulase in which the non-charged amino acid residue of position 71 (this is the position of an amino acid residue adjacent to an acidic amino acid) in the amino acid sequence of a mutant alkaline cellulase having a mutation of substitution of the glutamine residue at position 242 of the amino acid sequence set forth in SEQ ID NO: 2 by a serine residue, with an acidic amino acid residue. Such a substitution particularly efficiently increases the hydrophilicity in the vicinity of the substituted amino acid residue, and provides remarkably high anti-redeposition ability to the resulting mutant alkaline cellulase.

Another preferred example may be a mutant alkaline cellulase in which the non-charged amino acid residue at position 193 in the amino acid sequence set forth in SEQ ID NO: 2 (this is the position where there is an amino acid residue which is not adjacent to an acidic amino acid) is substituted with a basic amino acid residue. Such substitution particularly efficiently increases the hydrophilicity in the vicinity of

the substituted amino acid residue, and provides remarkably high anti-redeposition ability to the resulting mutant alkaline cellulase

Another preferred example may be a mutant alkaline cellulase in which the non-charged amino acid residue at position 193 (this is the position where there is an amino acid residue which is not adjacent to an acidic amino acid) in the amino acid sequence of a mutant alkaline cellulase having a mutation of substitution of the glutamine residue at position 242 of the amino acid sequence set forth in SEQ ID NO: 2 with a serine residue, is substituted with a basic amino acid residue. Such a substitution particularly efficiently increases the hydrophilicity in the vicinity of the substituted amino acid residue, and provides remarkably high anti-redeposition ability to the resulting mutant alkaline cellulase.

As another preferred example, the mutant alkaline cellulase according to the present invention includes, for example, a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting one or more amino acid residues selected from the non-charged amino acid residues at the 20 positions corresponding to positions 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, 175, 176, 178, 179, 181, 193, 194, 195, 196, 197, 199, 202, 203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 312, 318, 324, 345, 354, 356, 357, 360, 363, and 368 of 25 the amino acid sequence of SEQ ID NO: 2 and of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2, with charged amino acid residues, and having an amino acid sequence that is obtained by further removing a signal sequence (corresponding to a 30 sequence including the positions 1 to 30 of the sequence of SEQ ID NO: 2) from the foregoing sequence.

Another preferred example includes a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting one or more amino acid residues selected from 35 the non-charged amino acid residues at the positions corresponding to positions 16, 23, 27, 31, 35, 37, 42, 74, 90, 93, 94, 95, 96, 98, 101, 111, 132, 135, 146, 147, 149, 150, 152, 164, 165, 166, 167, 168, 170, 173, 174, 188, 196, 198, 199, 222, 238, 243, 247, 248, 251, 253, 268, 281, 283, 289, 295, 316, 40 325, 327, 328, 331, 334, and 339 of the amino acid sequence of SEQ ID NO: 4 and of the amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 4, with charged amino acid residues, and having an amino acid sequence that is obtained by further removing the 45 signal sequence from the foregoing sequence.

Another preferred example includes a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting one or more amino acid residues selected from the non-charged amino acid residues at the positions corresponding to positions 45, 52, 56, 60, 64, 66, 71, 103, 119, 122, 123, 124, 125, 127, 130, 140, 161, 164, 175, 176, 178, 179, 181, 193, 194, 195, 196, 197, 199, 202, 203, 217, 225, 227, 228, 251, 267, 272, 276, 277, 280, 282, 297, 310, 312, 318, 324, 345, 354, 356, 357, 360, 363, and 368 of the amino acid sequence of SEQ ID NO: 6 and of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 6, with charged amino acid residues, and having an amino acid sequence that is obtained by further removing the signal sequence from the foregoing sequence.

Another preferred example includes a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting one or more amino acid residues selected from the non-charged amino acid residues at the positions corresponding to positions 44, 51, 55, 59, 63, 65, 70, 102, 118, 121, 65 122, 123, 124, 126, 129, 139, 160, 163, 174, 175, 177, 178, 180, 192, 193, 194, 195, 196, 198, 201, 202, 216, 224, 226,

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227, 250, 266, 271, 275, 276, 279, 281, 296, 309, 311, 317, 323, 344, 353, 355, 356, 359, 362, and 367 of the amino acid sequence of SEQ ID NO: 8 and of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 8, with charged amino acid residues, and having an amino acid sequence that is obtained by further removing the signal sequence from the foregoing sequence.

Another preferred example includes a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting one or more amino acid residues selected from the non-charged amino acid residues at the positions corresponding to positions 226, 233, 237, 241, 245, 247, 284, 300, 303, 304, 305, 306, 320, 344, 361, 376, 377, 378, 379, 380, 382, 385, 386, 400, 431, 447, 456, 460, 462, 477, 490, 492, 498, 504, 525, 534, 536, 537, and 548 of the amino acid sequence of SEQ ID NO: 10 and of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 10, with charged amino acid residues, and having an amino acid sequence that is obtained by further removing the signal sequence from the foregoing sequence.

Another preferred example includes a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting one or more amino acid residues selected from the non-charged amino acid residues at the positions corresponding to positions 63, 71, 75, 79, 83, 85, 90, 122, 138, 142, 143, 144, 146, 193, 207, 208, 210, 222, 223, 224, 225, 226, 228, 231, 232, 246, 254, 256, 257, 281, 297, 302, 306, 310, 348, 354, 375, 384, 386, and 387 of the amino acid sequence of SEQ ID NO: 12 and of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 12, with charged amino acid residues, and having an amino acid sequence that is obtained by further removing the signal sequence from the foregoing sequence.

According to an even more preferred exemplary embodiment, the mutant alkaline cellulase according to the present invention includes, for example, a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting the amino acid residues in the region that directly participates in cellulose binding, and a region adjacent thereto, in the cellulose binding module in an alkaline cellulase having the amino acid sequence set forth in SEQ ID NO: 2 and an amino acid sequence having at least 70% identity with the amino acid sequence of SEQ ID NO: 2, with other amino acid residues, and having an amino acid sequence that is obtained by further removing a signal sequence (corresponding to a sequence including the positions 1 to 30 of SEQ ID NO: 2) from the foregoing sequence.

The mutant alkaline cellulase according to this exemplary embodiment exhibits anti-redeposition ability based on a decrease in the cellulose binding properties at the cellulose binding module of the cellulase. Therefore, in the case of using the mutant alkaline cellulase simply for the purpose of obtaining an anti-redeposition effect, it makes no difference whether the mutant alkaline cellulase may have cellulase activity or not. However, from the viewpoint that a glycolytic activity against saccharides including cellulose can also be simultaneously obtained, it is more preferable that the mutant alkaline cellulase related to the present invention have cellulase activity. The mutant alkaline cellulase according to the present invention may have a signal sequence at the N-terminal, or may be in the form of a mature protein having the signal sequence removed therefrom.

In the mutant alkaline cellulase according to the exemplary embodiment described above, the substitution of an amino acid residue as an object by other amino acid residue may be, for example, a substitution of tryptophan (non-polar amino acid) with tyrosine (polar amino acid), or may be a substitu-

tion of an amino acid residue other than tryptophan [that is, as non-polar amino acids, valine, leucine, isoleucine, methionine, phenylalanine and proline; as polar amino acids, glycine, asparagine, cysteine, glutamine, serine, threonine and tyrosine; as acidic amino acids, aspartic acid and glutamic 5 acid; and as basic amino acids, arginine, histidine and lysine] with alanine (non-polar amino acid). Such a substitution provides remarkably high anti-redeposition ability to the mutant alkaline cellulase according to the present invention.

A preferred example of the mutant alkaline cellulase of the present invention in the exemplary embodiment described above is a mutant alkaline cellulase in which one or more amino acid residues selected from the amino acid residues at the positions 419, 421, 454, and 501 of the amino acid sequence set forth in SEQ ID NO: 2 (these are the positions of 15 amino acid residues that directly participate in cellulose binding and their suitable adjacent site (position 640)) are substituted with other amino acid residues. Such a substitution decreases the cellulose binding property in the vicinity of the substituted amino acid residues, and as a result, the substitution brings about a partial decrease in the binding property to cellulose of the resulting mutant alkaline cellulase, while provides remarkably high anti-redeposition ability.

Another preferred example includes a mutant alkaline cellulase in which one or more amino acid residues selected from 25 the amino acid residues at the positions 419, 421, 454 and 501 (these are the positions of amino acid residues that directly participate in cellulose binding and their suitable adjacent site (position 640)) in the amino acid sequence of a mutant alkaline cellulase having a mutation of substitution of the 30 glutamine residue at position 242 of the amino acid sequence set forth in SEQ ID NO: 2 by a serine residue, are substituted with other amino acid residues.

As another preferred example, the mutant alkaline cellulase according to the present invention includes, for example, 35 a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting one or more amino acid residues selected from the amino acid residues at the positions corresponding to positions 418, 419, 420, 421, 422, 453, 454, 455, 457, 458, 459, 494, 495, 496, 500, 501, 502, 503, 504, 40 550, 551, 552, 604, 605, 606, 607, 608, 640, 641, 642, 644, 645, 646, 683, 684, 685, 690, 691, 692, 693, 694, 739, 740, and 741 of the amino acid sequence of SEQ ID NO: 2 and of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2, with other amino acid 45 residues, and having an amino acid sequence that is obtained by further removing a signal sequence (corresponding to a sequence including the positions 1 to 30 of SEQ ID NO: 2) from the foregoing sequence.

Another preferred example includes a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting one or more amino acid residues selected from the amino acid residues at the positions corresponding to positions 389, 390, 391, 392, 393, 424, 425, 426, 428, 429, 430, 465, 466, 467, 471, 472, 473, 474, 475, 521, 522, 523, 5523, 575, 576, 577, 578, 579, 611, 612, 613, 615, 616, 617, 654, 655, 656, 661, 662, 664, 665, 710, 711, and 712 of the amino acid sequence of SEQ ID NO: 4 and of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 4, with other amino acid residues, and having an amino acid sequence that is obtained by further removing the signal sequence from the foregoing sequence.

Another preferred example includes a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting one or more amino acid residues selected from 65 the amino acid residues at the positions corresponding to positions 418, 419, 420, 421, 422, 452, 453, 454, 456, 457,

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458, 493, 494, 495, 499, 500, 501, 502, 503, 548, 549, 550, 602, 603, 604, 605, 606, 638, 639, 640, 642, 643, 644, 681, 682, 683, 688, 689, 690, 691, 692, 737, 738, and 739 of the amino acid sequence of SEQ ID NO: 6 and of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 6, with other amino acid residues, and having an amino acid sequence that is obtained by further removing the signal sequence from the foregoing sequence (the strain 1139-derived cellulase contains a signal sequence consisting of 30 amino acids).

Another preferred example includes a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting one or more amino acid residues selected from the amino acid residues at the positions corresponding to positions 417, 418, 419, 420, 421, 451, 452, 453, 455, 456, 457, 492, 493, 494, 498, 499, 500, 501, 502, 547, 548, 549, 601, 602, 603, 604, 605, 637, 638, 639, 641, 642, 643, 680, 681, 682, 687, 688, 689, 690, 691, 736, 737, and 738 of the amino acid sequence of SEQ ID NO: 8 and of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 8, with other amino acid residues, and having an amino acid sequence that is obtained by further removing the signal sequence from the foregoing sequence (the strain KSM-64-derived cellulase contains a signal sequence consisting of 29 amino acids).

Another preferred example includes a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting one or more amino acid residues selected from the amino acid residues at the positions corresponding to positions 598, 599, 600, 601, 602, 633, 634, 635, 637, 638, 639, 674, 675, 676, 680, 681, 682, 683, 684, 729, 730, 731, 783, 784, 785, 786, 787, 819, 820, 821, 823, 824, 825, 862, 863, 864, 869, 870, 871, 872, 873, 919, 920, and 921 of the amino acid sequence of SEQ ID NO: 10 and of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 10, with other amino acid residues, and having an amino acid sequence that is obtained by further removing the signal sequence from the foregoing sequence (the strain KSM-635-derived cellulase contains a signal sequence consisting of 29 amino acids).

Another preferred example includes a mutant alkaline cellulase having an amino acid sequence that is obtained by substituting one or more amino acid residues selected from the amino acid residues at the positions corresponding to positions 451, 452, 453, 454, 455, 486, 487, 488, 490, 491, 492, 527, 528, 529, 533, 534, 535, 536, 537, 583, 584, 585, 639, 640, 641, 642, 643, 675, 676, 677, 679, 680, 681, 720, 721, 722, 727, 728, 729, 730, 731, 775, 776, and 777 of the amino acid sequence of SEQ ID NO: 12 and of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 12, with other amino acid residues, and having an amino acid sequence that is obtained by further removing the signal sequence from the foregoing sequence (the strain N-4-derived cellulase contains a signal sequence consisting of 28 amino acids).

The mutant alkaline cellulase according to the present invention can be produced by using various technologies for introducing mutation that are known in the pertinent art. For example, the mutant alkaline cellulase according to the present invention can be produced by mutating the nucleotide sequence that encodes an amino acid residue to be substituted in the alkaline cellulase gene that encodes a parent amino acid sequence of the mutant alkaline cellulase (parent alkaline cellulase gene), to a nucleotide sequence that encodes an amino acid residue after substitution, and then expressing the mutant alkaline cellulase from the mutant gene.

The intended introduction of mutation into the parent alkaline cellulase gene can be carried out essentially based on PCR amplification in which the parent alkaline cellulase gene is used as a template DNA, or replication reactions using various DNA polymerases, by using various site-directed mutagenesis methods that are well known to those ordinary skilled in the art. The site-directed mutagenesis method can be carried out by, for example, any technique such as an inverse PCR method or an annealing method (Muramatsu, et al., ed., "New Genetic Engineering Handbook, Revised 4th Edition", Yodosha Co., Ltd., p. 82-88). If necessary, various commercially available kits for site-directed mutagenesis, such as QuickChange II Site-Directed Mutagenesis Kit and QuickChange Multi Site-Directed Mutagenesis Kit by Stratagene, Inc. may be used. In the present invention, a method of preparing DNA fragments by respectively amplifying the upstream side and the downstream side of the mutated site by separately using two complementary mutation primers containing the nucleotide mutations to be introduced, and linking 20 the DNA fragments into one by SOE (splicing by overlap extension)-PCR (Horton R. M. at al., Gene (1989) 77(1), p. 61-68) may be used. The procedure for the introduction of mutation using this SOE-PCR method will be described in detail in the Examples described below.

The template DNA containing the parent alkaline cellulase gene can be prepared by extracting the genomic DNA from a organism that produces an alkaline cellulase by a routine method, or by extracting the RNA and synthesizing a cDNA by reverse transcription. As the organism that produces an 30 alkaline cellulase, bacteria including bacteria of the genus Bacillus, such as Bacillus subtilis, bacteria of the genus Clostridium and bacteria of the genus Acidothermus, as well as plants and animals have been reported. However, most advanced studies have been conducted on bacteria of the 35 genus Bacillus, such as Bacillus subtilis, and those organisms are easily available to those ordinary skilled in the art. For example, Bacillus sp. strain KSM-S237 (Accession No. FERM BP-7875), strain KSM-64 (Accession No. FERM BP-1485) have been deposited in the International Patent Organism Depositary, National Institute of Advanced Industrial Science and Technology (Central 6, 1-1-1, Higashi, Tsukuba-shi, Ibaraki-ken, Japan), under the respective indicated accession numbers.

Preparation of the genomic DNA from these bacterial strains of the genus *Bacillus* can be carried out by using, for example, the method described in Pitcher et al., Lett. Appl. Microbiol., 1989, 8:p. 151-156, or the like. The template DNA containing the parent alkaline cellulase gene may be 50 prepared in the form in which a DNA fragment containing the parent alkaline cellulase gene that has been excised from a prepared cDNA or the genomic DNA, is inserted into an available vector. Meanwhile, previously reported DNA sequences (sequences registered in GenBank) including the 55 base sequences encoding Bacillus sp. strain KSM-S237-derived alkaline cellulase (SEQ ID NO: 2), strain DMS12648derived alkaline cellulase (SEQ ID NO: 4), strain 1139-derived alkaline cellulase (SEQ ID NO: 6), strain KSM-64derived alkaline cellulase (SEQ ID NO: 8), strain KSM-635- 60 derived alkaline cellulase (SEQ ID NO: 10) and strain N-4derived alkaline cellulase (SEQ ID NO: 12) are set forth in SEQ ID NO: 1 (GenBank Accession No. AB018420), SEQ ID NO: 3, SEQ ID NO: 5 (GenBank Accession No. D00066), SEQ ID NO: 7 (GenBank Accession No. M84963), SEQ ID 65 NO: 9 (GenBank Accession No. M27420), and SEQ ID NO: 11 (GenBank Accession No. M25500), respectively.

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The site-directed mutagenesis into the parent alkaline cellulase gene can be carried out, most generally by using a mutation primer containing the nucleotide mutation to be introduced. Such a mutation primer may be annealed into a region containing the nucleotide sequence encoding the amino acid residues to be subjected in the parent alkaline cellulase gene, and may be designed to include a base sequence having the nucleotide sequence (codon) that encodes the amino acid residue after substitution, instead of the nucleotide sequence (codon) that encodes the amino acid residue to be substituted. The nucleotide sequences (codons) that encode the amino acid residues to be substituted and the amino acid residues after substitution can be appropriately recognized and selected by a person ordinary skilled in the art, based on conventional textbooks and the like.

For example, in the case of substituting the glutamine residue at position 58 of S237 cellulase (SEQ ID NO: 2) with an arginine residue, a primer containing a sequence in which the codon CAA (positions 744 to 746 of SEQ ID NO: 1) corresponding to the glutamine has been changed into the arginine codon GAA (Q58R-FW; SEQ ID NO: 30) and a primer having a complementary sequence thereof (Q58R-RV; SEQ ID NO: 29) can be used as the mutation primers.

Furthermore, for example, in the case of substituting ala-25 nine at position 56 of S237 cellulase (SEQ ID NO: 2) with aspartic acid, a primer containing a sequence in which the codon GCA (positions 738 to 740 of SEQ ID NO: 1) corresponding to the alanine has been changed to the aspartic acid codon GAT, 5'-TCTGAGGCTGGCGATTTACAATTA-CAAG-3' (A56D-FW; SEQ ID NO: 33) and a primer having a complementary sequence thereof, 5'-CTTGTAATTG-TAAATCGCCAGCCTCAGA-3' (A56D-RV; SEQ ID NO: 34) can be used as the mutation primers.

Furthermore, for example, in the case of substituting asparagine at position 419 of S237 cellulase (SEQ ID NO: 2) with alanine, a primer containing a sequence in which the codon AAT (positions 1827 to 1829 of SEQ ID NO: 1) corresponding to the asparagine has been changed to the alanine codon GCT, 5'-AGGATTTGGAGTGGCTTCGGATTCTCCAAA-BP-2886), and strain KSM-635 (Accession No. FERM 40 3' (N419A-FW; SEQ ID NO: 22) and a primer having a complementary sequence thereof (N419A-RV; SEQ ID NO: 21) can be used as the mutation primers.

The primers used in the present invention can be produced by a well known oligonucleotide synthesis method such as a phosphoroamidite method (Nucleic Acids Research, 17, 7059-7071, 1989). Such synthesis of primers can be carried out by using, for example, a commercially available oligonucleotide synthesizer (manufactured by Applied Biosystems, Inc. or the like). When site-directed mutagenesis such as described above is carried out by using a primer set including mutation primers and using the parent alkaline cellulase gene as a template DNA, a mutant alkaline cellulase gene having the intended mutation introduced therein can be obtained. The present invention relates also to a mutant alkaline cellulase gene that may be obtained as such. Furthermore, the term "mutant alkaline cellulase gene" as used in the present invention means any nucleic acid fragment (including DNA, mRNA, artificial nucleic acid, and the like) encoding the amino acid sequence of a mutant alkaline cellulase. The "gene" according to the present invention may include other base sequences such as an untranslated region (UTR), in addition to an open reading frame.

A recombinant vector can be produced by inserting the mutant alkaline cellulase gene thus obtained into an available vector by a conventional method, and linking the gene to the vector. There are no particular limitations on the vector used in the present invention, and any vector such as a plasmid, a

phage, a phagemid, a cosmid, a virus, a YAC vector, or a shuttle vector may be used. Such a vector is more preferably, but not limited to, a vector which can be amplified in bacterial cells, particularly bacterial cells of the genus Bacillus, and is even more preferably an expression vector capable of induc- 5 ing the expression of a transgene in the bacterial cells of the genus Bacillus. Among others, a shuttle vector, which is a vector capable of replication even in any of organisms other than bacteria of the genus Bacillus, can be particularly suitably used in the recombinant production of a mutant alkaline cellulase. Preferred examples of the vector include, but are not limited to, shuttle vectors such as pHY300PLK (an expression vector capable of transforming both Escherichia coli and Bacillus subtilis; Ishikawa, H. and Shibahara, H., Jpn. J. Genet., (1985) 60, p. 235-243), and pAC₃ (Moriyama, 15 H. et al., Nucleic Acids Res. (1988) 16, p. 8732); plasmids that can be utilized in the transformation of bacteria of the genus Bacillus, such as pUB110 (Gryczan, T. J. et al., J. Bacteriol. (1978) 134, p. 318-329), and pTA10607 (Bron, S. et al., Plasmid, 18 (1987), p. 8-15); and secretion vectors 20 capable of adding secretion signals to recombinant proteins (Yamane, et al., "Fusion Protein Produced by Bacillus subtilis Secretion Vector", Denpun Kagaku (Starch Science), 34 (1987), p. 163-170). Furthermore, Escherichia coli-derived plasmids (for example, pET22b(+), pBR322, pBR325, 25 pUC118, pUC119, pUC18, pUC19, and pBluescript) can also be used.

For the purpose of producing a recombinant mutant alkaline cellulase, the vector is preferably an expression vector. The expression vector may include various elements that are essential to the expression in a host organism (a transcription promoter, a terminator, a ribosome binding site and the like), as well as cis-elements such as a selection marker gene, a polylinker and an enhancer, and useful sequences such as a poly(A) addition signal and a ribosome binding sequence (SD 35 sequence), as necessary.

A transformant can be produced by using a recombinant vector containing the mutant alkaline cellulase gene. In the present invention, when a transformant (transformed cell) is produced by introducing a recombinant vector (specifically, a 40 recombinant expression vector) containing the mutant alkaline cellulase gene according to the present invention into a host cell, and the transformant is cultured under the conditions under which the expression of a recombinant protein is induced, the mutant alkaline cellulase can be produced. The 45 present invention also relates to a transformant produced as such, and a method for producing a mutant alkaline cellulase using the transformant. As the host cell into which a recombinant vector is introduced, microorganisms including bacteria such as Escherichia coli and Bacillus subtilis, and yeast 50 cells, as well as any cells such as insect cells, animal cells (for example, mammalian cells) and plant cells can be used. According to the present invention, it is particularly preferable to use bacteria of the genus Bacillus, such as Bacillus

Transformation can be carried out by applying well known transformation technologies such as a calcium phosphate method, an electroporation method, a lipofection method, a particle gun method, and a PEG method. Examples of the transformation method applicable to bacteria of the genus 60 *Bacillus* include a competent cell transformation method (Bott, K. F. and Wilson, G. A., J. Bacteriol. (1967) 93, 1925), an electroporation method (Brigidi, P. et al., FEMS Microbiol. Lett. (1990) 55, 135), a protoplast transformation method (Chang, S. and Cohen, S. N., Mol. Gen. Genet., 65 (1979) 168, p. 111-115), and a Tris-PEG method (Takahashi W., et al., J. Bacteriol. (1983) 156, p. 1130-1134).

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The transformant for recombinant protein production can be cultured according to methods that are commonly used by those ordinarily skilled in the art. For example, as a medium for culturing a transformant based on a microbial host such as Escherichia coli or a yeast cell, any of a natural medium and a synthetic medium may be used as long as it is a medium which contains carbon sources, nitrogen sources, inorganic salts and the like that can be assimilated by the host organism, and can efficiently carry out the culture of the transformant. Ampicillin, tetracyclin and the like may also be added to the medium, in accordance with the type of the drug selection marker. When a microorganism that has been transformed with an expression vector having an inducible promoter is cultured, an inducer may also be added to the medium as necessary. For example, in the case of culturing a bacterium or the like that has been transformed with an expression vector having a Lac promoter, isopropyl-1-thio-β-D-galactoside (IPTG) and the like can be added to the medium, and in the case of culturing a bacterium that has been transformed with an expression vector having a trp promoter, indole acetic acid (IAA) and the like can be added to the medium. There are no particular limitations on the culture conditions, and preferably, the culture is carried out under the conditions suitable for the transformed host organism. For example, in the culture of a Bacillus subtilis transformant for producing a recombinant protein, for example, LB medium, 2×YT medium, 2×Lmaltose medium, and CSL fermentation medium can be used.

The mutant alkaline cellulase according to the present invention may be expressed from a mutant alkaline cellulase gene or a transcription product thereof, using a cell-free translation system. The "cell-free translation system" is an in vitro transcription/translation system or an in vitro translation system constructed by adding reagents such as amino acids needed for the translation of proteins, to a suspension obtained by mechanically homogenizing the host cells. The mutant alkaline cellulase thus expressed can be acquired from a culture fluid, a cell lysate or a cell-free translation system, by using a general method used for protein purification, for example, centrifugation, ammonium sulfate precipitation, gel chromatography, ion exchange chromatography or affinity chromatography, alone or in appropriate combination. However, a solution such as a culture supernatant or a lysate supernatant separated or concentrated by using centrifugation, an ultrafiltration type filter or the like, can be directly used as a crude enzyme fluid. When the mutant alkaline cellulase thus expressed is not secreted from the cells, the cells may be homogenized, and then separation and purification of the protein may be carried out.

For the mutant alkaline cellulase produced as described above, an enhancement of the anti-redeposition ability can be confirmed by an anti-redeposition ability evaluation method that will be described below.

Furthermore, experiments such as the preparation of the mRNA used in the present invention, production of a cDNA, 55 PCR, RT-PCR, production of a library, ligation into a vector, transformation of cells, determination of the base sequence of DNA, chemical synthesis of nucleic acid, determination of the N-terminal side amino acid sequence of the protein, mutagenesis, and extraction of the protein, can be carried out according to the methods described in conventional manuals for experiment. Examples of such manuals include Molecular Cloning, A laboratory manual (2001) 3rd Ed., Sambrook, J. & Russell, D W. Cold Spring Harbor Laboratory Press. Particularly, for the experiment of genetic recombination of *Bacillus subtilis*, for example, reference can be made to general experiment manuals on the genetic manipulation of *Bacillus subtilis*, such as Yoshikawa, Hirofumi, "7.2 *Bacillus subtilis*

family" "Genetic Research Method II in Lectures on Biochemical Experiment, 2" series", (1986), Vol. 1, Tokyo Kagakudojin (Tokyo), p. 150-169).

2. Evaluation of Properties of Mutant Alkaline Cellulase (2-1) Anti-Redeposition Ability

The mutant alkaline cellulase of the present invention shows enhanced anti-redeposition ability as compared with its parent alkaline cellulase.

The "anti-redeposition ability" of the alkaline cellulase (the mutant alkaline cellulase or the parent alkaline cellulase) 10 according to the present invention means the ability of an alkaline cellulase that is incorporated into an aqueous solution, to prevent hydrophobic (oleophilic) dirty materials that are dispersed in the aqueous solution from re-adhering to substrates such as the clothes present in the aqueous solution. 15 pared to substrates such as the clothes present in the aqueous solution. 15 pared to substrates a clothes in the aqueous solution to substrates such as clothes in the aqueous solution in the presence of an alkaline cellulase (the mutant alkaline cellulase or the parent 20 12.5% alkaline cellulase), and the degree of such an effect.

Preferably, the evaluation of the anti-redeposition ability can be carried out by dispersing carbon black as a model for hydrophobic soot stain in water containing a dissolved cleaning agent composition, adding the mutant alkaline cellulase produced as described above to the dispersion liquid to prepare a washing water, washing a white cotton cloth using the washing water, measuring the reflection ratio at 550 nm of the white cloth after washing, and comparing the reflection ratio with the same reflection ratio of an unwashed white cotton cloth. The details of the procedure of the anti-redeposition ability evaluation method are disclosed in the Examples that are described below. Meanwhile, the water used in the anti-redeposition ability evaluation of the present invention can be prepared by appropriately dissolving CaCl₂ and MgCl₂.6H₂O 35 in deionized water.

In the anti-redeposition ability evaluation, the anti-redeposition ratio obtained in various test systems can be calculated by the following formula based on the reflection ratio of a white cotton cloth before washing and after washing.

Anti-redeposition ratio (%)={(Reflection ratio of white cotton cloth after test)/(reflection ratio of white cotton cloth before test)}×100 [Mathematical formula 1]

A value of this redeposition reflection ratio closer to 100% $_{45}$ indicates that the amount of redeposition by carbon black is smaller

Furthermore, the degree of promotion of anti-redeposition (%) as a result of the addition of a mutant alkaline cellulase can be calculated based on the following formula, and on the anti-redeposition ratio calculated for a test system using a washing water to which a mutant alkaline cellulase is added (enzyme-added group) and a test system using a washing water to which a mutant alkaline cellulase is not added (non-enzyme-added group).

Degree of promotion of anti-redeposition by enzyme addition (%)={(Anti-redeposition ratio of enzyme-added group)-(anti-redeposition ratio of non-enzyme-added group)}/{100-(anti-redeposition ratio of non-enzyme-added group)}×100 [Mathematical formula 2]

An increase in this degree of promotion of anti-redeposition (%) indicates that the effect of preventing redeposition by the addition of a mutant alkaline cellulase has greatly improved.

Furthermore, the degree of anti-redeposition ability enhancement (%) for individual mutant alkaline cellulases is

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calculated by the following formula. Further, the "mutant enzyme" in the following formula refers to the mutant alkaline cellulase, and the control enzyme refers to the parent alkaline cellulase, that is, a wild-type alkaline cellulase or an alkaline cellulase before the introduction of mutation.

Degree of anti-redeposition ability enhancement (%) for mutant alkaline cellulase={(Anti-redeposition ratio of mutant enzyme-added group)-(anti-redeposition ratio of control enzyme-added group)}/
{100-(anti-redeposition ratio of control enzyme-added group)}×100 [Mathematical formula 3]

An increase of this degree of anti-redeposition ability enhancement (%) indicates that the anti-redeposition ability of the mutant alkaline cellulase is greatly improved as compared with before the introduction of mutation.

Although there are no limitations, the mutant alkaline cellulase according to the present invention can exhibit a degree of anti-redeposition ability enhancement (%) of, for example, 1% to 25%, usually 2% to 15%, and more generally 3% to 12.5%.

(2-2) Cellulose Binding Property

Furthermore, the mutant alkaline cellulase of the present invention can show decreased cellulose binding property as compared with the parent alkaline cellulase.

The decrease of the cellulose binding property in the mutant alkaline cellulase according to the present invention can be confirmed according to a method similar to the antiredeposition ability evaluation method that will be described below. Specifically, an evaluation cloth (white cotton cloth) is washed in washing water prepared by adding a predetermined amount of an alkaline cellulase, and sodium chloride in an amount of 5% relative to the total amount of the washing water, to water in which a cleaning agent composition is dissolved, using an agitation-type detergency tester at 20° C. After the washing, the white cloth is taken out and lightly wrung out, and is rapidly introduced into 2000 mL of tap water. Subsequently, the white cloth is taken out and dehydrated without rinsing, and is stained with Coomassie Brilliant Blue. The cloth is lightly wrung out and then immersed 40 in a decolorization solution. The cloth is washed with water and is subjected to finish ironing. Subsequently, the brightness (L value) is measured using a spectrophotometer. A control experiment is carried out by the same procedure, except that a wild-type alkaline cellulase is added to the washing water. A comparison is made for the brightness (L value) of the evaluation clothes after washing thus obtained. This L value decreases along with an increase in the amount of proteins adsorbed to the evaluation cloth. Accordingly, as the L value is higher compared to the L value obtained by adding a wild-type alkaline cellulase, it is shown that the amount of the mutant alkaline cellulase adsorbed to the evaluation cloth is reduced, that is, the cellulose binding property is decreased.

(2-3) Protease Resistance

Furthermore, the mutant alkaline cellulase of the present invention can show enhanced protease resistance as compared with the parent alkaline cellulase.

According to the present invention, the "protease resistance" refers to the stability of the cellulase to protease, which is evaluated by the residual activity of the cellulase in a cleaning agent composition in which various proteases, particularly alkaline proteases, have been incorporated.

Specifically, for example, the protease resistance may be the residual activity (see the following formula) of a cellulase to be evaluated, which is calculated when the cellulase is stored in a cleaning agent composition containing an alkaline protease at 40° C. for 24 hours.

Cellulase residual activity (%)=(Cellulase activity after 24 hours of storage/cellulase activity immediately after preparation)×100 [Mathematical formula 4]

The mutant alkaline cellulase of the present invention exhibits a cellulase residual activity (%) of, for example, 20% to 75%, and usually 50% to 70%, so that the mutant alkaline cellulase exhibits a residual activity enhanced by 15% to 35% as compared with the parent alkaline cellulase.

Therefore, the mutant alkaline cellulase of the present invention having enhanced protease resistance is suitable for use in the co-presence of various proteases. Here, examples of the proteases include commercially available Alcalase, Esperase, Savinase, Everlase, Kannase (registered trademark; Novozymes, Inc.), Properase, Purafect (registered trademark; Genencor, Inc.), and KAP (Kao Corp.).

3. Use of Mutant Alkaline Cellulase

The mutant alkaline cellulase according to the present invention shows high anti-redeposition ability towards hydrophobic dirt materials in an aqueous solution. Therefore, the mutant alkaline cellulase according to the present invention 20 can be advantageously used as an anti-redeposition agent. The anti-redeposition agent may contain any additives such as an inert carrier, a pH adjusting agent, a dispersant, a buffering agent and an antiseptic agent, in addition to the mutant alkaline cellulase. When such an anti-redeposition agent is added to an aqueous solution containing an object to be washed, the redeposition of the object to be washed can be favorably prevented. For example, the anti-redeposition agent according to the present invention can be incorporated in detergents such as detergents for clothing and detergents for 30 domestic use, fabric softeners and the like, and used.

The present invention also provides an enzyme composition containing the mutant alkaline cellulase according to the present invention. The enzyme composition according to the present invention means an enzyme preparation containing 35 the mutant alkaline cellulase as an active ingredient. The enzyme composition according to the present invention may further include, in addition to the mutant alkaline cellulase, hydrolases including proteases, cellulases, β-glucanases, hemicellulases, lipases, peroxidases, laccases, α-amylases, 40 glucoamylases, cutinases, pectinases, reductases, oxidases, phenoloxidases, ligninases, pullulanases, pectate lyases, xyloglucanases, xylanases, pectinacetylesterases, polygalacturonases, rhamnogalacturonases, pectin lyases, other mannanases, pectin methylesterases, cellobiohydrolases, and 45 transglutaminases, as well as mixtures of two or more kinds thereof.

The enzyme composition according to the present invention may also contain other components such as a pH adjusting agent, a buffering agent, an antiseptic, salt, alcohol, sugars, and medium components, in addition to the mutant alkaline cellulase and the other enzymes. The enzyme composition according to the present invention may be in any form such as a powder, granules, or a lyophilized product.

The present invention also provides a cleaning agent composition containing one or more of the mutant alkaline cellulase, anti-redeposition agent, and enzyme composition according to the present invention. The cleaning agent composition according to the present invention may contain known cleaning agent components, for example, a surfactant, 60 a divalent metal ion scavenger, an alkali agent, an anti-redeposition agent, a bleaching agent, a fluorescent agent and the like, in addition to the mutant alkaline cellulase described above, or an anti-redeposition agent and/or enzyme composition containing the mutant alkaline cellulase.

As the surfactant, any surfactants such as anionic surfactants, nonionic surfactants, amphoteric surfactants and cat28

ionic surfactants, can be used singly or in combination of two or more kinds. A more preferred surfactant may be an anionic surfactant or a nonionic surfactant.

Preferred examples of the anionic surfactant include sulfuric acid ester salts of alcohols having 10 to 18 carbon atoms, sulfuric acid ester salts of alkoxylation products of alcohols having 8 to 20 carbon atoms, alkylbenzenesulfonates, alkyl sulfuric acid ester salts, paraffin sulfonates, α -olefin sulfonates, α -sulfo fatty acid salts, α -sulfo fatty acid alkyl ester salts, and fatty acid salts. For example, linear alkylbenzenesulfonates and alkyl sulfates having alkyl chains having 10 to 14 carbon atoms, and more preferably 12 to 14 carbon atoms, can be suitably used as the anionic surfactant in the present invention. As the counter ions of these salts, alkali metal salts or amines are preferred, and particularly, sodium, potassium, monoethanolamine and diethanolamine are preferred.

Preferred examples of the nonionic surfactant include polyoxyalkylene alkyl (C_8 - C_{20}) ethers, alkyl polyglycosides, polyoxyalkylene alkyl (C_8 - C_{20}) phenyl ethers, polyoxyalkylene sorbitan fatty acid (C_8 - C_{22}) esters, polyoxyalkylene glycol fatty acid (C_8 - C_{22}) esters, and polyoxyethylene polyoxypropylene block polymers. For example, polyoxyethylene (average number of added moles of EO: 6) alkyl (C_{12} - C_{14}) ethers can be suitably used as the nonionic surfactant in the present invention.

The total amount of the surfactant in the cleaning agent composition according to the present invention can be appropriately selected by a person having ordinary skill in the art. However, from the viewpoints of detergency and solubility, the total amount of the surfactant is preferably 10 to 60% by mass, more preferably 15 to 50% by mass, and even more preferably 20 to 45% by mass, relative to the mass of the cleaning agent composition. Among others, the content of the anionic surfactant is preferably 1 to 60% by mass, more preferably 1 to 50% by mass, and even more preferably 3 to 40% by mass, relative to the mass of the cleaning agent composition. Furthermore, the content of the nonionic surfactant is preferably 1 to 45% by mass, more preferably 1 to 35% by mass, and even more preferably 4 to 25% by mass, relative to the mass of the cleaning agent composition. The anionic surfactants and the nonionic surfactants can be used singly, and preferably the surfactants are used as mixtures. Further, amphoteric surfactants and cationic surfactants can also be used in combination in accordance with the purpose.

The cleaning agent composition according to the present invention may further include a builder. The builder is a compound which itself has no or only slight detergency, however, when incorporated together with a surfactant, can markedly enhance the detergent ability of the surfactant. Examples of the action of the builder include polyvalent metal cation scavenging action, dirt dispersing action, alkali buffering action, and combinations of two or more kinds thereof. Examples of such a builder include water-soluble inorganic compounds, water-insoluble inorganic compounds, and organic compounds.

Examples of builders as water-soluble inorganic compounds include phosphates (tripolyphosphates, pyrophosphates, metaphosphates, trisodium phosphate, and the like), silicates, carbonates, sulfates, and sulfites. Among them, phosphates are preferred in view of having all of the three types of actions. Examples of builders as water-insoluble inorganic compounds include aluminosilicates (A-type zeolite, P-type zeolite, X-type zeolite, amorphous aluminosilicates, and the like), and crystalline silicates. Examples of builders as organic compounds include carboxylates (aminocarboxylates, hydroxyaminocarboxylates, hydroxycarboxylates, cyclocarboxylates, maleic acid derivatives,

oxalates, and the like), organic carboxylic acid (salt) polymers (acrylic acid polymers and copolymers, polyvalent carboxylic acid (for example, maleic acid and the like) polymers and copolymers, glyoxylic acid polymers, polysaccharides and salts thereof). Among them, organic carboxylic acid (salt) polymers are preferred. With regard to the salt of the builder, alkali metal salts and amines are preferred as the counter ions, and sodium, potassium, monoethanolamine and diethanolamine are more preferred. The builder contained in the cleaning agent composition according to the present invention 10 preferably includes the water-soluble inorganic compounds described above, and is more preferably a combination of the water-soluble inorganic compounds and organic compounds described above. The builder is even more preferably a combination of the water-soluble inorganic compounds, organic 15 compounds and water-insoluble inorganic compounds.

The total amount of the builder in the cleaning agent composition according to the present invention can be appropriately selected by a person ordinary skilled in the art, and the total amount is preferably 20 to 80% by mass, more prefer- 20 ably 30 to 70% by mass, and even more preferably 35 to 60% by mass, relative to the mass of the cleaning agent composition. Among them, the content of the water-soluble inorganic compound builder is preferably 10 to 50% by mass, more preferably 15 to 45% by mass, and even more preferably 20 to 25 40% by mass, relative to the mass of the cleaning agent composition. Among them, the content of the water-insoluble inorganic compound builder is preferably 5 to 50% by mass, more preferably 10 to 45% by mass, and even more preferably 15 to 40% by mass, relative to the mass of the cleaning agent 30 composition. Among them, the content of the organic compound builder is preferably 0.1 to 20% by mass, more preferably 0.3 to 15% by mass, and even more preferably 0.5 to 10% by mass, relative to the mass of the cleaning agent composition.

Specific preferred compositions of the cleaning agent composition according to the present invention include the following compositions A to E.

Composition A: 20% by weight of sodium linear alkyl $(C_{12}-C_{14})$ benzenesulfonate, 4% by weight of a nonionic 40 surfactant (polyoxyethylene alkyl ether having 12 to 16 carbon atoms and an average number of added moles of ethylene oxide of 6.0), 30% by weight of sodium carbonate, 10% by weight of sodium sulfate, 30% by weight of zeolite (4A-type zeolite (manufactured by Tosoh Corp.)), 2% by weight of an 45 acrylate-maleate copolymer, and 4% by weight of crystalline silicate (powder SKS-6 (manufactured by Hoechst Tokuyama, Ltd.)).

Composition B: 24% by weight of sodium linear alkyl $(C_{12}-C_{14})$ benzenesulfonate, 5% of linear alkyl $(C_{10}-C_{13})$ 50 sulfuric acid ester sodium, 6% of a fatty acid $(C_{14}-C_{18})$ sodium salt, 7% by weight of a nonionic surfactant (polyoxyethylene alkyl ether having 12 to 16 carbon atoms and an average number of added moles of ethylene oxide of 6.0), 12% of sodium tripolyphosphate, 12% by weight of sodium 55 carbonate, 6% by weight of sodium sulfate, 14% by weight of zeolite (4A-type zeolite (manufactured by Tosoh Corp.)), 6% by weight of sodium polyacrylate (average molecular weight 10,000), and 8% by weight of crystalline silicate (powder SKS-6 (manufactured by Hoechst Tokuyama, Ltd.)).

Composition C: 12% by weight of sodium linear alkyl $(C_{12}-C_{14})$ benzenesulfonate, 11% by weight of a nonionic surfactant (polyoxyethylene alkyl ether having 12 to 16 carbon atoms and an average number of added moles of ethylene oxide of 6.0), 28% by weight of sodium carbonate, 11% by weight of sodium sulfate, 28% by weight of zeolite (4A-type zeolite (manufactured by Tosoh Corp.)), 8% by weight of

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sodium polyacrylate (average molecular weight 10,000), and 2% by weight of crystalline silicate (powder SKS-6 (manufactured by Hoechst Tokuyama, Ltd.)).

Composition D: 14% by weight of sodium linear alkyl (C_{12} - C_{14}) benzenesulfonate, 2% of a fatty acid (C_{14} - C_{18}) sodium salt, 10% by weight of a nonionic surfactant (polyoxyethylene alkyl ether having 12 to 16 carbon atoms and an average number of added moles of ethylene oxide of 6.0), 23% of sodium tripolyphosphate, 29% by weight of sodium carbonate, 6% by weight of sodium sulfate, 11% by weight of zeolite (4A-type zeolite (manufactured by Tosoh Corp.)), 3% by weight of sodium polyacrylate (average molecular weight 10,000), and 2% by weight of crystalline silicate (powder SKS-6 (manufactured by Hoechst Tokuyama, Ltd.)).

Composition E: 20% by weight of a nonionic surfactant (polyoxyethylene alkyl ether having 12 to 16 carbon atoms and an average number of added moles of ethylene oxide of 12.0), 1% by weight of alkylbenzyldimethylammonium chloride (alkyl group having 8 to 18 carbon atoms), 20% by weight of Softanol 7014 (manufactured by Nippon Shokubai Co., Ltd.), 1.5% by weight of an acrylate-maleate copolymer, 1.5% by weight of monoethanolamine, 1.15% by weight of citric acid, 5% by weight of butyl diglycol, 2% by weight of ethanol, 0.2% by weight of sodium sulfite, and 47.65% by weight of water.

The cleaning agent composition according to the present invention may further include other components such as water, a pH adjusting agent, a buffering agent, a dispersant, an antiseptic, an oxidation inhibitor, an excipient, a dye such as a fluorescent dye, a deodorizer, a deodorant, a fragrance, a softening agent, and a plant extract. The cleaning agent composition according to the present invention may be in any form such as a powder, granules, a compression molded tablet, or a liquid. The cleaning agent composition according to the present invention is such that an amount for one-time use may be packaged in sachets, from the viewpoints of portability or convenience, and in that case, the packaging material is preferably water-soluble.

Although there are no limitations, the cleaning agent composition according to the present invention is preferably intended to be used for garments or for clothing products (sheets, curtains, carpets, wall clothes, and the like). Since the cleaning agent composition according to the present invention contains a mutant alkaline cellulase showing high anti-redeposition ability, the cleaning agent composition can exhibit a satisfactory anti-redeposition effect.

The amount of incorporation of the mutant alkaline cellulase of the present invention into a cleaning agent composition is not particularly limited as long as it is an amount by which the alkaline cellulase exhibits activity. However, the amount of incorporation is preferably 0.1 to 5000 U, more preferably 1 to 3000 U, and even more preferably 10 to 2000 U, per kilogram of the cleaning agent composition.

EXAMPLES

Hereinafter, the present invention is more specifically described by way of Examples. However, the scope of the present invention is not intended to be limited to these Examples.

Meanwhile, the experimental procedure, reagents and the like that were commonly employed are described first in the following descriptions.

1) Amplification of DNA Fragment

The amplification of a DNA fragment was carried out by a polymerase chain reaction (PCR) using a GeneAmp PCR system (Applied Biosystems, Inc.) and using a Pyrobest DNA polymerase (Takara Bio, Inc.) and accessory reagents. The

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reaction liquid composition for PCR was obtained by mixing 1 μL of an appropriately diluted template DNA, 20 pmol each of a sense primer and an antisense primer, and 2.5 U of a Pyrobest DNA polymerase, and adding water to the mixture to adjust the total amount of the reaction liquid to 50 μL . The PCR reaction was carried out under the conditions of repeating 30 cycles of a three-stage temperature change cycle of 10 seconds at 98° C., 30 seconds at 55° C., and 1 to 5 minutes at 72° C. (the time was adjusted in accordance with the target amplification product, but was adjusted on the basis of 1 minute per kb), and then performing the reaction for 5 minutes at 72° C.

The primers used in the DNA fragment amplifications that will be described below are presented in Table 1-1 to Table $_{15}$ 1-3.

TABLE 1-1

Primer name	Primer sequence (5'→3')	SEQ ID NO:
237UB1	TTGCGGATCCAACAGGCTTATATTTAGAGGAAATTTC	13
S237RV	TCGCTACCCTTTTATTATCG	14
Q71E-RV	ATTTTTTCTCCATGTTCATCTACTAATGTC	15
Q71E-FW	GACATTAGTAGATGAACATGGAGAAAAAAT	16
S193R-RV	TCCACCATTATTATTACGACTCGGCTCA	17
S193R-FW	TGAGCCGAGTCGTAATAATAATGGTGGA	18
Q242S-RV	AGTCCGGACGCGAACTCCAGTTTG	19
Q242S-FW	CAAACTGGAGTTCGCGTCCGGACT	20

TABLE 1-2

Primer name	Primer sequence (5'→3')	SEQ ID NO:
N419A-RV	TTTGGAGAATCCGAAGCCACTCCAAATCCT	21
N419A-FW	AGGATTTGGAGTGGCTTCGGATTCTCCAAA	22
D421A-RV	CTTTATTTGGAGAAGCCGAATTCACTCCA	23
D421A-FW	TGGAGTGAATTCGGCTTCTCCAAATAAAG	24
W454Y-RV	AGACGAGCATTAGCATAGAAGTTGCCATCT	25
W454Y-FW	AGATGGCAACTTCTATGCTAATGCTCGTCT	26
W501Y-RV	CTCTGGATTTGCATATCCACTTTTAC	27
W501Y-FW	GTAAAAGTGGATATGCAAATCCAGAG	28

TABLE 1-3

Primer name	Primer sequence (5'→3')	SEQ ID NO:
Q58R-RV	ATCGACTTCTTGTAATTCTAATGCGCCA	29
Q58R-FW	TGGCGCATTAGAATTACAAGAAGTCGAT	30
Q58E-RV	ATCGACTTCTTGTAAACGTAATGCGCCA	31
Q58E-FW	TGGCGCATTACGTTTACAAGAAGTCGAT	32

Furthermore, the primer sets for upstream region amplification and for downstream region amplification, which were used to introduce intended amino acid mutations to S237 cellulase or a mutant S237 cellulase in Examples 3 to 5, are presented in Table 2-1 to Table 2-3 (see the Examples described below for the details).

TABLE 2-1

Amino acid mutation to be introduced	Glutamine at position 71 → glutamic acid	Serine at position 193 → arginine
Primer set for upstream	237UB1	237UB1
region amplification	Q71E-RV	S193R-RV
Primer set for downstream	Q71E-FW	S193R-FW
region amplification	S237RV	S237RV

TABLE 2-2

)	Amino acid mutation to be introduced	Asparagine at position 419 → alanine	Aspartic acid at position 421 → alanine	Tryptophan at position 454 → tyrosine	Tryptophan at position 501 → tyrosine
5	Primer set for upstream region amplification	237UB1 N419A-RV	237UB1 D421A-RV	237UB1 W454Y-RV	237UB1 W501Y-RV
)	Primer set for downstream region amplification	N419A-FW S237RV	D421A-FW S237RV	W454Y-FW S237RV	W501Y-FW S237RV

TABLE 2-3

i	Amino acid mutation to be introduced	Glutamine at position 58 → arginine	Glutamine at position 58 → glutamic acid	Glutamine at position 242 → serine
)	Primer set for upstream region amplification Primer set for downstream region amplification	237UB1 Q58R-RV Q58R-FW S237RV	237UB1 Q58E-RV Q58E-FW S237RV	237UB1 Q242S-RV Q242S-FW S237RV

2) Gene Introduction into Bacillus subtilis

The introduction of a gene encoding S237 cellulase or a mutant S237 cellulase into *Bacillus subtilis* was carried out according to any one of a competent cell transformation method (J. Bacteriol. 93, 1925 (1967)), an electroporation method (FEMS Microbiol. Lett. 55, 135 (1990)), and a protoplast transformation method (Mol. Gen. Genet. 168, 111 (1979)).

In the competent cell transformation method, first, Bacillus subtilis (Bacillus subtilis Marburg No. 168 (Nature, 390, (1997), p. 249)) was cultured by shaking in SPI medium (0.20% ammonium sulfate, 1.40% dipotassium hydrogen phosphate, 0.60% potassium dihydrogen phosphate, 0.10% trisodium citrate dihydrate, 0.50% glucose, 0.02% casamino acid (Difco Laboratories, Inc.), 5 mM magnesium sulfate, 0.25 manganese chloride, and 50 μg/ml tryptophan) at 37° C., until the degree of growth (OD600) reached about 1. After the shaking, a portion of the culture fluid was inoculated into a 9-fold amount of SPII medium (0.20% ammonium sulfate, 1.40% dipotassium hydrogen phosphate, 0.60% potassium dihydrogen phosphate, 0.10% trisodium citrate dihydrate, 0.50% glucose, 0.01% casamino acid (Difco Laboratories, Inc.), 5 mM magnesium sulfate, 0.40 µM manganese chloride, and 5 µg/ml tryptophan), and the cells were further

cultured by shaking until the degree of growth (OD600) reached about 0.4. Thus, *Bacillus subtilis* cells were prepared as competent cells. Subsequently, in $100\,\mu L$ of the competent cell suspension thus prepared (competent cell culture in SPII medium), 2 μL of a solution containing a plasmid vector 5 having a gene encoding S237 cellulase or a gene encoding a mutant S237 cellulase was added, and the mixture was shaked for additional one hour at 37° C. Subsequently, the entire amount was spreaded on LB agar medium (1% tryptone, 0.5% yeast extract, 1% NaCl, and 1.5% agar) containing 10 suitable antibiotics for selection. The cells were statically cultured at 37° C., and then grown colonies were isolated as a transformant.

In the protoplast transformation method, first, a Bacillus subtilis strain (Bacillus subtilis Marburg No. 168 (Nature, 19 390, (1997), p. 249)) was cultured by shaking in 50 mL of LB medium (1% tryptone, 0.5% yeast extract, and 1% NaCl) at 37° C. for about 2 hours, and at the time point at which the absorbance at 600 nm reached 0.4, the bacterial cells were collected by centrifugation (7000 rpm, for 15 minutes) at 20 room temperature. The collected bacterial cells were suspended in 5 mL of SMMP [0.5 M sucrose, 20 mM disodium maleate, 20 mM magnesium chloride hexahydrate, and 35% (w/v) Antibiotic Medium 3 (Difco Laboratories, Inc.)], and then 500 µL of a lysozyme solution (30 mg/mL) dissolved in 25 SMMP solution was added to the suspension. The mixture was incubated at 37° C. for one hour to convert the bacterial cells to protoplasts. After completion of the incubation, the protoplasts were collected by centrifugation (2800 rpm, for 15 minutes) at room temperature and were suspended in 5 mL 30 of SMMP to prepare a protoplast solution. To 0.5 mL of the protoplast solution, 10 µL of a plasmid solution (containing a plasmid vector which included a gene encoding S237 cellulase or a gene encoding a S237 cellulase variant) and 1.5 mL of 40% (w/v) polyethylene glycol (PEG8000, Sigma-Aldrich 35 Co.) were added, and the mixture was gently stirred and left to stand for 2 minutes at room temperature. Immediately, 5 mL of a SMMP solution was mixed into the mixture, and the protoplasts were collected by centrifugation (2800 rpm, for 15 minutes) at room temperature and were resuspended in 1 40 mL of a SMMP solution. The protoplast suspension was shaken (120 rpm) for 90 minutes at 37° C., and then the suspension was applied on DM3 regeneration agar medium [0.8% (w/v) agar (Wako Pure Chemical Industries, Ltd.), 0.5% disodium succinate hexahydrate, 0.5% casamino acid 45 technical (Difco Laboratories, Inc.), 0.5% yeast extract, 0.35% monopotassium phosphate, 0.15% dipotassium phosphate, 0.5% glucose, 0.4% magnesium chloride hexahydrate, 0.01% bovine serum albumin (Sigma-Aldrich Co.), 0.5% carboxymethyl cellulose, 0.005% trypan blue (Merck 50 GmbH), and an amino acid mixture liquid (10 µg/mL each of tryptophan, leucine and methionine)] containing tetracycline (15 μg/mL, Sigma). The protoplasts were cultured for 72 hours at 30° C., and grown colonies were isolated as a trans-

In the culture of the transformants for production of recombinant protein in the Examples described below, LB medium (1% tryptone, 0.5% yeast extract, and 1% NaCl) was used as a medium for seed culture, and 2×YT medium (1.6% tryptone, 1% yeast extract, and 0.5% NaCl) or 2×L-maltose 60 medium (2% tryptone, 1% yeast extract, 1% NaCl, 7.5% maltose, and 7.5 ppm manganese sulfate tetra- or pentahydrate) was used as a medium for main culture.

3) Preparation of Cleaning Agent Composition

In the evaluation of the anti-redeposition ability, a cleaning 65 agent of the preferred composition C or E described above, or IEC-A detergent (composition F) supplied from Wfk

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Testgewebe GmbH (D-41379, Germany) was used as the cleaning agent composition. In the evaluation of an enzyme stability test, a cleaning agent of the composition E was used.

4) Evaluation of Anti-Redeposition Ability

The evaluation of the anti-redeposition ability was carried out according to the detergency evaluation method described in JIS K3362: 1998, as follows. 0.33 g of a cleaning agent composition was dissolved in 50 mL of water (in the case of 4° DH, CaCl₂: 55.42 mg/L, MgCl₂.6H₂O: 43.51 mg/L; in the case of 12° DH, CaCl₂: 166.26 mg/L, MgCl₂.6H₂O: 130.53 mg/L), and 0.125 g of carbon black (Asahi Carbon Black for cleaning, manufactured by Asahi Carbon Co., Ltd., or Carbon Black #4000B, MA100 or #40, manufactured by Mitsubishi Chemical Corp.) as a model for hydrophobic soot stain was added to the solution. 50 mL of water (CaCl₂: 55.42 mg/L, MgCl₂.6H₂O: 43.51 mg/L) was added to the mixture, and then the mixture was exposed with ultrasonic waves at 26 ± 1.5 kHz for 5 minutes to uniformly disperse the components. 400 mL of water (CaCl₂: 55.42 mg/L, MgCl₂.6H₂O: 43.51 mg/L) at 20° C. was further added to the dispersion liquid, and a predetermined amount of alkaline cellulase (S237 cellulase or mutant S237 cellulase) was added to the mixture. This was used as washing water. The washing water thus prepared was transferred into the sample cup of an agitation-type detergency tester, Terg-O-To meter; Ueshima Seisakusho Co., Ltd.) at 20° C. As a cloth for evaluation, 5 sheets of a white cotton cloth (#2003 white woven fabric, 100% cotton, supplied by Tanigashira Shoten (4-11-15, Komatsu, Higashiyodogawa-ku, Osaka-shi, Osaka-fu, Japan)) having a size of 6 cm×6 cm were placed in the sample cup. In order to further adjust the amount of cloth (bath ratio) relative to the solution, an appropriate amount of a white knitted cotton cloth [seared bleached cloth (supplied by Tanigashira Shoten) that had been washed and then sufficiently rinsed] was introduced to the sample cup, and the contents were stirred for 10 minutes at a rotation speed of 80±4 rpm. Subsequently, the white cotton cloth was removed together with the white knitted cotton cloth, and the clothes were lightly wrung out and then rapidly introduced into 2000 mL of tap water. Only the white cotton cloth was taken out therefrom, and then the cloth was rinsed for 3 minutes under flowing tap water and was subjected to dehydration and finish ironing. The reflection ratio at 550 nm of the white cotton cloth was measured using a spectrophotometer, CM-3500d (Konica Minolta Holdings, Inc.) (the samples are referred to as enzyme-added group). For the white cotton cloth, the reflection ratio at 550 nm was measured in advance before the washing test, using a spectrophotometer, CM-3500d (Konica Minolta Holdings, Inc.). A control experiment was carried out by the same procedure, except that no alkaline cellulase was added to the dispersion liquid (non-enzyme-added group).

The anti-redeposition ratio for each of the washing test was calculated by the following formula, based on the reflection ratio thus obtained.

Anti-redeposition ratio (%)={(Reflection ratio of white cotton cloth after test)/(reflection ratio of white cotton cloth before test)}×100 [Mathematical formula 5]

Subsequently, the effect of enzyme addition on the preventing of redeposition, that is, the degree of promotion of antiredeposition as a result of the addition of an alkaline cellulase enzyme, was calculated by the following formula, based on the anti-redeposition ratio thus calculated. In the Examples described below, the anti-redeposition ability of the added alkaline cellulase was evaluated by using this degree of promotion of anti-redeposition (%) as an index.

Degree of promotion of anti-redeposition by enzyme addition (%)={(Anti-redeposition ratio of enzyme-added group)-(anti-redeposition ratio of non-enzyme-added group)}/{100-(anti-redeposition ratio of non-enzyme-added group)}×100 [Mathematical formula 6]

Furthermore, the effect of mutagenesis in the alkaline cellulase on the anti-redeposition ability of the enzyme, that is, the degree of anti-redeposition ability enhancement in the mutant alkaline cellulase, was calculated by the following formula. Meanwhile, the mutant enzyme was a mutant alkaline cellulase into which an intended amino acid substitution had been introduced, and the control enzyme was a parent alkaline cellulase, that is, a wild-type alkaline cellulase or an alkaline cellulase before substitution of the amino acid residue.

Degree of anti-redeposition ability enhancement (%) for mutant alkaline cellulase={(Anti-redeposition ratio of mutant enzyme-added group)-(anti-redeposition ratio of control enzyme-added group)}/{100-(anti-redeposition ratio of control enzyme-added group)}x100 [Mathematical formula 7]

5) Evaluation of Anti-Redeposition Ability of *Bacillus* sp. Strain KSM-S237-Derived Alkaline Cellulase (S237 Cellulase)

(1. Recombinant Production of S237 Cellulase)

A nucleic acid fragment (3.1 kb) including an alkaline cellulase gene that encodes S237 cellulase (SEQ ID NO: 2) derived from *Bacillus* sp. strain KSM-S237 (FERM BP-7875) [hereinafter, also referred to as S237 cellulase gene; the base sequence is available based on GenBank 30 Accession No. AB18420 (SEQ ID NO: 1); Hakamada et al., Biosci. Biotechnol. Biochem., 64(11), (2000) p. 2281-2289; JP-A No. 2000-210081] was amplified according to the procedure of the section "1) Amplification of DNA fragment" as described above, using a primer set consisting of primers 35 237UB1 and S237RV indicated in the above Table 1-1. As a template DNA, the genomic DNA extracted from the strain KSM-S237 by a routine method was used.

The amplified fragment was inserted into the SmaI restriction enzyme cleavage site of a shuttle vector, pHY300PLK 40 (Yakult Honsha Co., Ltd.; Ishiwa, H. & Shibahara, H., Jpn. J. Genet. (1985) 60, p. 235-243), and thus a recombinant plasmid, pHY-S237, was constructed. The sequence of the S237 cellulase gene fragment inserted into the plasmid was determined by using a 3100 DNA Sequencer (Applied Biosys- 45 tems, Inc.), and thereby, confirmed that the fragment had the base sequence set forth in SEO ID NO: 1. Subsequently, Bacillus subtilis (Bacillus subtilis Marburg No. 168 (Nature, 390, (1997) p. 249)) was transformed using the recombinant plasmid, pHY-S237, according to the section "2) Gene intro- 50 duction into Bacillus subtilis" by protoplast transformation method. The transformant thus obtained was cultured overnight by shaking at 30° C. in 10 mL of LB medium (1% tryptone, 0.5% yeast extract, and 1% NaCl), and 0.05 mL of this culture fluid was inoculated into 50 mL of 2×L-maltose 55 medium (2% tryptone, 1% yeast extract, 1% NaCl, 7.5% maltose, 7.5 ppm manganese sulfate tetra- or pentahydrate, and 15 ppm tetracycline) and was cultured by shaking for 3 days at 30° C. A supernatant of the culture fluid, from which bacterial cells had been removed by centrifugation, was 60 diluted 10 times with deionized water, and then the diluted solution was loaded in a DEAE-Toyopearl 650C (Tosoh Corp.) column (1 cm×3 cm) which had been equilibrated with a 20 mM sodium phosphate buffer solution (pH 6.0). The column was washed with 10 mL of a 20 mM sodium phosphate buffer solution (pH 6.0) containing 0.075 M NaCl, and then proteins were eluted from the column using 10 mL of a

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20 mM sodium phosphate buffer solution (pH 6.0) containing 0.4 M NaCl. The target recombinant S237 cellulase was eluted as an electrophoretically almost single component. The eluted sample was dialyzed against a 10 mM Tris hydrochloride buffer solution (pH 7.5) containing 1 mM CaCl₂, and was subjected to desalting treatment. Subsequently, the content of the alkaline cellulase was measured by the following method. That is, 50 μL of 0.4 mM p-nitrophenyl-β-D-cellotrioside (Seikagaku Corp.) was added and mixed into 50 μL of the sample solution which had been appropriately diluted with a 1/7.5 M phosphate buffer solution (pH 7.4, Wako Pure Chemical Industries, Ltd.), and the amount of p-nitrophenol released when the reaction had been carried out at 30° C. was quantified based on the change of absorbance at 420 nm (OD420). The amount of the enzyme that releases 1 µmol of p-nitrophenol for one minute was defined as 1 U. Further, the amount of proteins was measured by using a Protein Assay Rapid Kit (manufactured by Wako Pure Chemical Industries, Ltd.) and the bovine serum albumin included in the kit as a standard, using.

A sample containing the recombinant S237 cellulase thus obtained (wild-type S237 cellulase) was subjected to evaluation of the anti-redeposition ability as described below. The evaluation was carried out by adding an enzyme protein in an amount equivalent to 26.4 mU, 52.8 mU, 106 mU, 211 mU or 264 mU, to 500 mL of the washing system.

(2. Anti-Redeposition Ability Evaluation Using Various Carbon Blacks)

The anti-redeposition ability of S237 cellulase was evaluated by using various carbon blacks. As the carbon blacks, 4 kinds of carbon blacks such as Asahi carbon black for cleaning manufactured by Asahi Carbon Co., Ltd., and Carbon Black #4000B, MA100 and #40 manufactured by Mitsubishi Chemical Corp. were used.

Prior to the anti-redeposition ability evaluation, the characteristics of each of the carbon blacks (hydrophobicity, and the acidic functional group content) were investigated as follows. 100 mL of an aqueous solution of 0.1% (w/v) polyoxyethylene (average number of added moles of EO 6) alkyl (C₁₂-C₁₄) ether was charged into a 100-mL beaker (Iwaki Co., Ltd.), 0.1 g of each carbon black powder was dropped from 10 cm above the surface of the aqueous solution to the surface, and the time necessary for settling the entire amount of the powder was measured. Measurement was repeated three times, and as a result, the time for settling was 4.9 ± 0.6 seconds for #4000B, 14.1±1.0 seconds for MA100, and 60.5±4.6 seconds for #40. For A portion of the Asahi carbon black for cleaning was staying on the surface of the aqueous solution even after a lapse of 10 minutes or longer, and the time necessary for the entire amount to be wetted was 78.8±9.1 seconds. According to the these results, it was found that among the various carbon blacks, hydrophobicity of #4000B was the lowest, followed by MA100 and #40 in this order, and hydrophobicity of the Asahi carbon black for cleaning was the highest. Subsequently, each of carbon black suspensions was exposed to ultrasonication at 26±1.5 kHz for 5 minutes, and while the suspension was thoroughly stirred, pH was measured. The pH of an aqueous solution of 0.1% (w/v) polyoxyethylene (average number of added moles of EO 6) alkyl (C₁₂-C₁₄) ether containing no carbon black was 4.75, and was 4.97 for #4000B, 4.69 for MA100, 4.81 for #40, and 4.86 for Asahi carbon black for cleaning after each of the carbon black was dispersed therein. pH of the dispersion liquid was decreased as a result of addition of MA100, the result suggests that MA100 contains the largest number of acidic functional groups (it may be considered that a majority of the functional groups are carboxyl groups), of the other

hand, #4000B contains the smallest acidic functional group content among the carbon blacks investigated.

Considering the characteristics of various carbon blacks as described above, an evaluation of the anti-redeposition ability was carried out according to the section "4) Evaluation of 5 anti-redeposition ability" described above. The results obtained by using a cleaning agent having the composition C and adding S237 cellulase in an enzyme amount equivalent to 0 mU, 106 mU or 264 mU, are shown in Table 3 (anti-redeposition ratio) and Table 4 (degree of promotion of anti-redeposition).

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When each of Endolase (Novozymes, Inc.) and Carezyme (Novozymes, Inc.) was used, the anti-redeposition ratio gradually increased along with increase of amount of the enzyme addition. However, when compared with S237 cellulase, the degree of the increase was small, and the anti-redeposition promoting effect was small. On the other hand, when S237 cellulase was used, anti-redeposition promoting effect, that was twice or more than that of Endolase, which exhibited a higher anti-redeposition promoting effect than Carezyme, was observed. These results suggest that S237

TABLE 3

		Anti-redeposition ratio (%)						
Enzyme	Asah	i cleaning		#40	N	IA100	#4	4000B
added	Mean	Standard	Mean	Standard	Mean	Standard	Mean	Standard
(mU/500 mL)	value	deviation	value	deviation	value	deviation	value	deviation
0	62.6	2.8	77.6	1.0	74.0	1.0	83.4	0.5
106	76.1	1.1	81.3	0.9	78.8	1.0	85.4	0.7
264	78.2	0.5	81.9	1.3	80.4	0.9	85.6	0.6

TABLE 4

Enzyme added	Degree of promotion of anti- zyme added redeposition due to enzyme addition (%)						
(mU/500 mL)	Asahi cleaning	#40	MA100	#4000B			
106 264	36.2 41.6	16.8 19.1	18.6 24.8	11.7 13.3			

As shown in Table 4, an effect of promoting prevention of the redeposition caused by any of the carbon blacks, as a result of the addition of S237 cellulase, was observed. The most significant anti-redeposition promoting effect was exhibited by Asahi carbon black for cleaning, and followed by MA100, #40 and #4000B in this order, with slight differences. These results of order suggest that the effect of promoting the preventing of redeposition due to the addition of S237 cellulase is greatly affected by degree of hydrophobicity of the carbon black.

Furthermore, for a comparison, an evaluation of the antiredeposition ability was carried out by the same method as described above, except that a protease, an amylase or a lipase as another type of hydrolase, and Endolase (Novozymes, Inc.) and Carezyme (Novozymes, Inc.) as another type of cellulase were used instead of S237 cellulase. Protein amounts of these enzymes were equivalent to concentrations of 26.4 mU, 52.8 mU and 211 mU. As the carbon black, Asahi carbon black for cleaning manufactured by Asahi Carbon Co., Ltd. was used. For a control experiment, an evaluation of the anti-redeposition ability was carried out under the same conditions using S237 cellulase. The results are shown in FIG. 2.

As a result, when a protease, an amylase or a lipase was used, the anti-redeposition ratios of the enzyme-added group and the non-enzyme-added group were all approximately 60 52%, without a significant difference, and the anti-redeposition promoting effect caused by the addition of these enzymes was not observed. On the other hand, when S237 cellulase was used, an increase in the anti-redeposition ratio was observed along with increase of amount of the enzyme addition, and therefore, the anti-redeposition promoting effect was confirmed.

cellulase provides high anti-redeposition promoting effect which is not observed when other enzymes are used.

(3. Investigation of pH Dependency of Anti-Redeposition Promoting Effect)

The effect of pH of washing water on the anti-redeposition promoting effect caused by enzyme addition was investigated as follows. An evaluation was carried out according to the anti-redeposition ability evaluation method described above, by using a cleaning agent of the composition E, and S237 cellulase in an enzyme amount equivalent to 0 mU or 52.8 mU, with the proviso that the pH of the cleaning liquid was adjusted to 8.0, 4.3, 4.1 or 3.6 by adding sulfuric acid.

As a result, as shown in FIG. 3, the anti-redeposition promoting effect was slightly decreased at pH 4.1, and was largely decreased at pH 3.6. Even in consideration of the fact disclosed in Patent Document 1 (which describes an invention related to S237 cellulase) that the pH stability of S237 cellulase greatly decreases near pH 4 (FIG. 3 in Patent Document 1), the above-described results suggest that the antiredeposition ability possessed by S237 cellulase is largely impaired under the conditions where an irreversible structural change will occur, such as pH decrease. Furthermore, S237 cellulase would hardly exhibit any activity under the conditions of pH 5 or lower (FIG. 5 in Patent Document 1), however the anti-redeposition promoting effect at pH 4.3 was recognized to be almost equivalent to the effect at pH 8. These findings suggest that the anti-redeposition promoting effect of S237 cellulase originates from the structure of the enzyme.

According to the evaluation of anti-redeposition ability described above, the promoting effect of less hydrophobic MA100 was found to be equal to or higher than the effect of #40, suggesting that the anti-redeposition ability of S237 cellulase is affected by the content of the acidic functional groups of carbon black, that is, the abundance of negative charge factors. Therefore, it could be considered that the anti-redeposition ability of S237 cellulase is dependent on the physical repulsion between S237 cellulase and hydrophobic substances (repulsion between hydrophilic groups and hydrophobic groups+electrostatic repulsion).

Furthermore, it was speculated, based on the results obtained from the investigation of pH dependency as described above, that when the properties derived from the structure of S237 cellulase are modified, the anti-redeposition ability may be enhanced.

Specifically, it was conceived that when the hydrophilicity of the enzyme surface is further increased while the basic structure of S237 cellulase is maintained, high repulsion is caused between the enzyme surface and hydrophobic carbon black, and consequently, the anti-redeposition ability of S237 5 cellulase can be enhanced (see FIG. 1B).

6) Steric Structure Modeling of S237 Cellulase

In order to increase the hydrophilicity of the enzyme surface of S237 cellulase, a method may be employed wherein a position which has less influence on the overall structure of 10 S237 cellulase, even if the position is substituted, is selected among the positions on the amino acid sequence where non-charged amino acid residues that are exposed to the enzyme surface in S237 cellulase are present, and the non-charged amino acid residue at the position is substituted with a 15 charged amino acid residue. Thus, in order to select suitable positions for substitution, at first, a steric structure model of S237 cellulase was constructed as described below.

Steric structure modeling of the catalytic domain of S237 cellulase was carried out by the following procedure, using 20 steric structures of the catalytic domain of *Bacillus* sp. strain KSM-635-derived alkaline cellulase (635 cellulase) (registered in Protein Data Bank (PDB); 1G01 and 1G0C) as models. IRIS Indigo2 Extreme was used as the computer, and insightII (Ver. 95.5) was used as a graphical user interface 25 program. The structure of S237 cellulase was constructed based on the structure of 635 cellulase, using a module of insight II, modeler 4. That is, the topology of the S237 cellulase sequence was produced by referring to the coordinates of 635 cellulase, and for the atoms for which the coordinates 30 could not be defined, allocation of the coordinates was carried out by referring to a residue topology file (RTF) of a polymer system modeling program, CHARMm (accessible from Chemistry at HARvard Macromolecular Mechanics, http:// www.charmm.org/). Subsequently, the restraint on the struc- 35 ture of S237 cellulase was calculated as a probability density function (PDF), based on the alignment between the amino acid sequence of 635 cellulase and the amino acid sequence of S237 cellulase. The number of models thus produced was previously fixed at 1, for producing models that satisfy as 40 many restraints as possible. Optimization of the models by a variable target function method (VTF) was carried out as follows. First, only those restraints that can be easily optimized (in which corresponding atoms are closely positioned each other) were selected, and energy minimization was car- 45 ried out by a conjugate gradient method. Subsequently, these steps were repeated for other restraint conditions also, and finally, energy minimization according to a conjugate gradient method was performed for all restraints. Next, model optimization by simulated annealing was carried out. Energy 50 minimization according to a conjugate gradient method, simulated annealing (heating conditions (Heating)-cooling conditions (Cooling)), and energy minimization according to a conjugate gradient method were carried out by using an optimization protocol "Low" (based on maximum speed 55 molecular dynamic simulated annealing), under the conditions in which only those restraints that did not satisfy the conditions and the atoms corresponding thereto were selected, while other atoms were fixed. Subsequently, all the restraints and atoms were selected and energy minimization 60 according to the conjugate gradient method was carried out. The degree of restraint violation was calculated, and after confirming that the level was not particulary irregular, the final model was constructed.

The model thus constructed was subjected to a visualization analysis using the program Discovery Studio Visualizer Ver. 1.5 (Accelrys Software, Inc.). The residue solvent acces-

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sibility of each amino acid residue was calculated by performing an analysis using the Solvent Accessibility program of Discovery Studio Visualizer Ver. 1.5, by setting the grid point for each atom at 240, and the probe radius at 1.40.

An amino acid residue having a value of this residue solvent accessibility of 50 or greater was considered as an amino acid residue having a high degree of surface exposure, and the relevant amino acid was appointed as a primary candidate of amino acid residues that can be selected as the objects of substitution in the present invention. From the primary candidates, first, charged amino acid residues were excluded. Next, the 3 amino acid residues in the N-terminal region and the 36 amino acid residues in the C-terminal region, which had low reliability of modeling, were also excluded. Furthermore, in consideration of that the constructed model was a model lacking the cellulose binding module (CBM domain), in the actual steric structure of S237 cellulase, the amino acid residues following the aspartic acid at position 369 (Asp369), which have a possibility of being covered by the CBM domain and not exposed to the surface, were excluded from the candidates for substitution. Furthermore, in the actual steric structure of S237 cellulase, leucine at position 42 (Leu42) to glycine at position 44 (Gly44), which have a possibility of being covered by the amino-terminal region that had been excluded from the constructed model, were excluded from the candidates for substitution. Further, the two tryptophan residues at position 88 and position 240 (Trp88 and Trp240) in the vicinity of the substrate binding pocket of S237 cellulase also have a possibility of participating in the binding with a substrate, and therefore, the tryptophan residues were excluded from the candidates. The asparagine residues at position 250 and position 330 (Asn250 and Asn330), around which both acidic amino acid residues and basic amino acid residues are present, were also excluded from the candidates for substitution. As a result, 55 noncharged amino acid residues were selected as the amino acid residues to be substituted of the present invention. The result suggests that when these non-charged amino acid residues thus selected are substituted with charged amino acid residues, the surface charge of S237 cellulase would be increased, and the anti-redeposition ability of S237 cellulase would be enhanced.

Furthermore, with regard to other cellulases sharing high identity with S237 cellulase, the result suggests that non-charged amino acid residues that are exposed to the enzyme surface may be present at the positions corresponding to these 55 amino acid residues with high possibility, and therefore the anti-redeposition ability of the cellulases would be enhanced by similarly substituting the amino acid residues at those positions with charged amino acid residues.

In the columns for "S237" in the following Table 5, 54 amino acid residues that can be considered as the target for substitution that have been selected as described above in connection with the catalytic domain of S237 cellulase are shown. Furthermore, Table 5 shows amino acid residues of other alkaline cellulases sharing high amino acid sequence identity with S237 cellulase, Bacillus sp. strain DSM12648derived alkaline cellulase (DSM12648 cellulase; SEQ ID NO: 4), Bacillus sp. strain 1139-derived alkaline cellulase (1139 cellulase; SEQ ID NO: 6), Bacillus sp. strain KSM-64derived alkaline cellulase (endo-1,4-β-glucanase) (64 cellulase; SEQ ID NO: 8), Bacillus sp. strain KSM-635-derived alkaline cellulase (KSM-635 cellulase; SEQ ID NO: 10), and Bacillus sp. strain N-4-derived alkaline cellulase (endoglucanase) (N4 cellulase; SEQ ID NO: 12), which are aligned to the 55 amino acid residues of S237 cellulase (that is, present at the positions corresponding to those residues) when an align-

ment of the amino acid sequence of each of the cellulases and the amino acid sequence of S237 cellulase is produced (Table 5). Each of the positions of the amino acid residues is indicated by the number of the amino acid residue in the amino acid sequence of the alkaline cellulase set forth in respective 5 sequence ID number.

								IABLE	J								
	S237			DSM12648			1139			49			KSM635	ĺ		N4	
Amino acid position	Amino acid (3-letter code)	Amino acid (1-letter code)	Amino acid position	Amino Amino acid acid (3-letter oosition code)	Amino acid (1-letter code) 1	Amino acid position	Amino acid (3-letter code)	Armino acid (1-letter code)	Amino acid position	Amino acid (3-letter code)	Amino acid (1-letter code)	Amino acid position	Amino acid (3-letter 1 code)	Amino acid (1-letter code)	Amino acid position	Amino acid (3-letter 1 code)	Amino acid (1-letter code)
45	Asn	z	16	Asn	z	45	Asn	z	44	Asn	z	226	Thr	T	63	Gly	Ü
52	Ser	S	23	Ser	S	52	Ser	S	51	Ser	S	233	Ser	S	71	Ser	S
99	Ala	A	27	Ala	A	99	Ala	Ą	55	Ala	Α	237	Ala	A	75	Ala	A
28	Gln	~	29	Gln	o	28	Gln	~	27	Gln	∼	239	Gln	0	77	Gln	~
99 ;	Gln	0	31	Gln	0	8	Gln	0	59	Gln	0	241	Val	> 1	79	Val	> 1
2	Gly	Ü	35	Gly	Ü	\$	Gly	Ü	63	Gly	Ů	245	Gly	Ü	83	Gly	Ü
99 i	Met	Σ	37	Met	Σ	9 i	Met	Σ	65	Met	Σ	247	Leu	ı,	85	Val	> 1
71	Gli	∵ ;	42	Gln) ;	71	Gli.	∵ ;	0/,	Gln	ς;	252	Glu	ш;	06	Gln	∵ ;
103	Asn	Z;	4 6	Asn	Z ;	103	Asn	Z;	102	Asn	z;	787	Asn	z;	122	Asn	z ;
119	Asn	z, -	9 5	Asn	z.	119	Asn	ζ.	118	Asn	Ζ.	200	Asn	Ζ.	138	Asn	Z, f
122	Ala	₹ 8	3,5	Ala	∢ ≀	771	Ala	₹ 0	171	Ala	∢ :	503	Ala	∢ :	141	Arg	⊻;
123	Thr.	- ;	46	Ser	ν ;	123	Ser	ν ;	122	Ser	ν.;	505 40 5	Thr	- ;	142	lyr	Υ ;
124	Asn	Z	6	Asn	Z	124	Asn	Z	123	Asn	Z	303	Asn	Z	143	Asn	Z
125	Pro	۸,	96	Pro	۱ بد	125	Pro	<u>م</u> ,	124	Pro	۰,	306	Pro	۵,	144	Pro	a, ,
127	Leu	٦	86	Leu	J	127	Leu	J	126	Leu	J				146	Leu	J
130	Gln	0	101	Ser	S	130	Ser	S	129	Ser	S	310	Asp	Ω	148	Glu	ш
140	Ile	I	111	lle	-	140	lle	_	139	Ile	I	320	Phe	Œ,	158	Lys	¥
161	Pro	Ь	132	Pro	Ь	161	Pro	Ь	160	Pro	Ы	341	Asp	О	179	Asp	D
<u>\$</u>	Ala	Ą	135	Ala	Ą	<u>\$</u>	Ala	A	163	Ala	Ą	2 4	Ser	S	193	Leu	Γ
175	Ala	Ą	146	Ala	Ą	175	Ala	A	174	Ala	Α	355	Asp	Ω	204	Glu	ш
176	Leu	l	147	Leu	L	176	Leu	ļ	175	Leu	J	356	His	Η	205	Lys	М
178	Pro	Ь	149	Pro	Ь	178	Pro	Ь	177	Pro	Ы	361	Pro	Ы	207	Pro	Ь
179	Asn	Z	150	Asn	z	179	Asn	Z	178	Asn	Z	362	Lys	¥	208	Asn	z
181	Pro	Ь	152	Pro	Ь	181	Pro	Ь	180	Pro	Д	36	His	Η	210	Pro	Ь
193	Ser	S	164	Ser	Ø	193	Ser	S	192	Ser	S	376	Pro	Д	222	Ser	Ø
194	Asn	Z	165	Asn	z	194	Asn	Z	193	Asn	Z	377	Asn	z	223	Asn	z
195	Asn	Z	166	Asn	Z	195	Asn	z	194	Asn	Z	378	Asn	Z	224	Ser	Ø
196	Asn	Z	167	Asn	Z	196	Asn	Z	195	Asn	Z	379	Asn	Z	225	Ser	ß
197	Gly	Ü	168	Gly	Ü	197	Gly	Ü	196	Gly	Ü	380	Gly	Ü	226	Gly	Ü
199	Ala	Ą	170	Ala	Ą	199	Ala	¥	198	Ala	Ą	382	Pro	Д,	228	Pro	Ы
202	Pro	Д	173	Pro	Ч	202	Pro	Ы	201	Pro	Д	385	Thr	Н	231	Thr	H
203	Asn	Z	174	Asn	Z	203	Asn	Z	202	Asn	Z	386	Asn	Z	232	Asn	Z
217	Pro	Ь	188	Pro	Ь	217	Pro	Ь	216	Pro	Ь	400	Pro	Д	246	Pro	Ь
225	Ser	S	196	Ser	S	225	Ser	S	224	Ser	S	408	Lys	¥	254	Ser	S
227	Asn	Z	198	Asn	Z	227	Asn	Z	226	Asn	Z				256	Asn	Z
228	Ala	Ą	199	Ala	Ą	228	Ala	Ą	227	Ala	Ą		1		257	Ala	Ą
251	Pro	Ь	222	Pro	Ь	251	Pro	Ь	250	Pro	Ы	431	Pro	Ы	281	Pro	Ь
267	Ser	S	238	Ser	S	267	Ser	S	566	Ser	S	447	Ser	S	297	Thr	L
272	Thr	Ι	243	Thr	T	272	Thr	T	271	Thr	T	452	His	Η	302	Asn	Z
276	Pro	Ь	247	Pro	Ь	276	Pro	Ь	275	Pro	Ь	456	Pro	Ы	306	Pro	Ь
277	Ser	S	248	Pro	Ь		Pro	Ь	276	Pro	Ь	457	Glu	ш	307	Glu	П
280	Pro	Ь	251	Pro	Ь		Pro	Ь	279	Pro	Ь	460	Pro	Ь	310	Ser	S
282	Ser	S	253	Ser	S		Ser	S	281	Ser	S	462	Ser	S	312	Glu	ы
297	Asn	Z	268	Asn	Z	297	Asn	Z	296	Asn	Z	477	Asn	Z	327	Ivs	Y
								;			i						

TABLE 5-continued

	Amino acid (1-letter code)	E	Y	> 2	<u>.</u>	Γ	Z	D	D	ы
N4	Amino Amino acid acid (3-letter oosition code)	340 Glu 342 Asp								
	Amino acid (1-letter code) F	σz	Y	> 2	<u> </u>	Γ	Ü	О	О	A
KSM635	Amino Amino acid acid (3-letter position code)	490 Gln 492 Asn								
	Amino acid (1-letter code)	οz	Y	> 2	<u>.</u>	J	Ğ	Z	S	Ь
64	Amino Amino acid acid (3-letter position code)	309 Gln 311 Asn								
	Amino acid (1-letter code)	ΟZ	Y	> >	T H	ļ	Ċ	Z	S	Ь
1139	Amino Amino acid acid (3-letter position code)	310 Gln 312 Asn								
ľ	Amino acid (1-letter code)	οz	Y	> 2	<u>.</u>	Т	Ğ	Z	Z	Ь
DSM12648	Amino Amino acid acid (3-letter position code)	281 Gln 283 Asn								
	Amino acid (1-letter code)	o s	Y	> 2	<u>.</u>	L	ŋ	Z	Z	Ь
S237	Amino acid Amino acid (3-letter position code)	310 Gln 312 Ser	-	324 Val 345 Asn					363 Asn	

In Examples 3 and 4 described below, the mutant S237 cellulases in which the amino acid residues at position 71 and position 193 among the non-charged amino acid residues at the positions selected as described above were substituted with charged amino acid residues, were constructed.

7) Investigation of Relationship Between Adsorption Power of Alkaline Cellulase to Cellulose and Anti-Redeposition Ability, and Determination of Amino Acid Residues to be Substituted that Enhance Anti-Redeposition Ability

Promotion of adsorption of the alkaline cellulases to cellulose by addition of sodium chloride was shown according to a method similar to the method of "4) Evaluation of antiredeposition ability", as follows.

Bacillus sp. strain KSM-635-derived alkaline cellulase (635 cellulase) was produced according to methods reported (in Agric. Bio. Chem., 55, 2387, 1991). Subsequently, 0.33 g of a cleaning agent composition was dissolved in 500 mL of water (CaCl₂: 55.42 mg/L, MgCl₂.6H₂O: 43.51 mg/L), and 2110 mU of 635 cellulase, and sodium chloride in an amount of 5% relative to the total amount of the washing water were added to the solution, to obtain a washing water. The washing water thus prepared was transferred into the sample cup of an agitation type detergency tester, Terg-O-To meter; Ueshima Seisakusho Co., Ltd.) at 20° C. As a cloth for evaluation, 5 sheets of a white cotton cloth (#2003 white woven fabric, 100% cotton, supplied by Tanigashira Shoten) having a size of 6 cm×6 cm were placed in the sample cup, and 40 g of a white knitted cotton cloth (seared bleached cloth (supplied by Tanigashira Shoten) that had been washed and then sufficiently rinsed) was introduced to the sample cup. The contents were stirred for 10 minutes at rotation speed of 80±4 rpm. Subsequently, the white cotton cloth was removed together with the white knitted cotton cloth, and the clothes were lightly wrung out and then rapidly introduced into 2000 mL of tap water. Only the white cotton cloth was taken out therefrom, and the cloth was dehydrated without rinsing and then was introduced into a Coomassie Brilliant Blue G staining solution (prepared by dissolving 2.5 g of Coomassie Brilliant Blue G250 (Merck GmbH), 4 g of methanol, and 90 mL of acetic acid in 910 mL of deionized water). After immersion for 30 minutes, the cloth was lightly wrung out and then was transferred into a decolorization solution (prepared by mixing 50 mL of deionized water, 50 mL of methanol and 10 mL of acetic acid). Immersion of the cloth in the decolorization solution for 30 minutes was repeated two times. Subsequently, the cloth was washed with water and was subjected to finish ironing. Then, the brightness (L value) was measured using a spectrophotometer, CM-3500d (Konica Minolta Holdings, Inc.). A control experiment was carried out by the same procedure, except that sodium chloride was not added to the washing water.

As a result, while the L value in the case of non-addition of sodium chloride was 92, the L value in the case of adding sodium chloride in an amount of 5% was 71. Since the L value decreases with the adsorption of proteins to the cloth, this decrease in the L value means an increase in the amount of adsorption of 635 cellulase to the white cotton cloth. That is, it was shown that the adsorption of the alkaline cellulase to cellulose was promoted by the addition of sodium chloride.

Subsequently, the anti-redeposition ability of 635 cellulase in the presence of sodium chloride was evaluated according to the section "4) Evaluation of anti-redeposition ability", using

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washing water to which sodium chloride had been added in an amount of 5%. A cleaning agent of the composition B, and 1000 mU of 635 cellulase were used. As the carbon black, Asahi carbon black for cleaning was used. As control experiments, the same evaluation with the proviso that sodium chloride was not added to the washing water, and that no cellulase was added, was carried out.

As a result, the anti-redeposition ratio in the case of adding sodium chloride was 47% in the washing water without any added cellulase, and was 30% in the washing water with an added cellulase, in which a decrease of the anti-redeposition ratio in the presence of a cellulase was observed. On the other hand, in the case where sodium chloride was not added, the anti-redeposition ratio was 72% in the washing water without addition of cellulase, and was 82% in the washing water without addition of cellulase, thus, an increase of the anti-redeposition ratio, in the presence of a cellulase, was observed. Taking into consideration of the above experimental results altogether, the result suggests that since the adsorption of the alkaline cellulase to cellulose was promoted in the presence of sodium chloride, a redeposition promoting effect was rather exhibited.

An alkaline cellulase adsorbs to cellulose via the cellulose binding module (CBM) in the interior of the enzyme. Thus, the result suggests that when the cellulose binding property of the alkaline cellulase via the cellulose binding module is decreased, it would be more difficult for the alkaline cellulase to adsorb to cellulose, therefore, higher anti-redeposition effect would be provided even under the condition where, for example, the cellulose adsorption is promoted (see FIG. 1B).

The cellulose binding module (CBM) of 635 cellulase or S237 cellulase consists of two kinds of CBMs that belong to CBM17 family and CBM28 family. Amino acid residues that directly participate in the binding to cellulose, which are included in the members of those CBM17 and CBM28 families, have already been reported (published as Biochem. J., 361, 35, 2002). For example, for the CBM17 of S237 cellulase, asparagine at position 419, aspartic acid at position 421, tryptophan at position 454, arginine at position 458, glutamine at position 495, tryptophan at position 501, asparagine at position 503, and asparagine at position 551 in the amino acid sequence set forth in SEQ ID NO: 2; and for the CBM28, alanine at position 605, glutamic acid at position 607, alanine at position 641, arginine at position 645, glutamine at position 684, tryptophan at position 691, glutamine at position 693, and isoleucine at position 740 participate in cellulose binding. Therefore, it suggests that when these amino acid residues are substituted with other amino acid residues, the binding property of the alkaline cellulase to cellulose may be weakened. Furthermore, it may be considered that amino acid residues adjacent to these amino acid residues (particularly, two amino acid residues adjacent to the relevant amino acid residues) also participate in cellulose binding either directly or indirectly. Therefore, the result suggests that the cellulose binding property may be weakened by substituting or deleting the amino acid residues that are adjacent to the amino acid residues that directly participate in cellulose binding, or by inserting amino acid residues at the positions more adjacent to the relevant residues. Based on such investigation results, 44 amino acid residues that can be considered as target of substitution so as to weaken the cellulose binding property of the alkaline cellulase are summarized in Table 6.

Amino and Options Ami		S237			DSM12648			1139			64			KSM635			N4	
Val Y 389 Val Y 418 Val Y 417 Val Y 418 Val Y 418 Val Y 419 Am Y 539 Val Y 419 Am Y 419 Am Y 450 Am Am 419 Am Y 451 Am Y 451 Am Y 451 Am Y 453 Am Y	Amino acid position		Amino acid (1-letter code)	Aminc acid positio	Amino acid (3-letter n code)	Amino acid (1-letter code)	Amino acid position	Amino acid (3-letter code)	Armino acid (1-letter code)	Amino acid position	Amino acid (3-letter code)	Amino acid (1-letter code)	Aminc acid positio	Amino acid (3-letter 1 code)	Amino acid (1-letter code)	Amino acid positio	Amino acid (3-letter n code)	Amino acid (1-letter code)
Ash N 391 Ash N 410 Ash Ash N 450 Ash Ash<	418	Val	Δ	389	Val	Δ	418	Val	>	417	Val	>	598	Val	>	451	Gln	С
Sker 5 391 Sker 430 Gly Gly Gl 600 Gly Gl 645 Sker Sker Sker 445 Sker Gl 600 Gly Gl 600 Gly Gl 600 Gly Gl 445 Sker 445 Me 451 Me	419	Asn	Z	390	Asn	Z	419	Asn	Z	418	Asn	Z	599	Asn	Z	452	Asn	Z
Asp D 437 Asp D 421 Asp D 421 Asp D 421 Asp D 421 Asp D 431 Asp Asp D 431 Asp	420	Ser	S	391	Ser	S	420	Glv	Ü	419	Glv	Ü	009	Glv	Ü	453	Ser	x
Sign Sign <th< td=""><td>421</td><td>Asp</td><td>Q</td><td>392</td><td>Asp</td><td>О</td><td>421</td><td>Asp</td><td>D</td><td>420</td><td>Asp</td><td>О</td><td>601</td><td>Asp</td><td>О</td><td>454</td><td>Asp</td><td>D</td></th<>	421	Asp	Q	392	Asp	О	421	Asp	D	420	Asp	О	601	Asp	О	454	Asp	D
Phys. F 424 Phys. F 452 Phys. F 452 Phys. F 452 Phys. F 454 Phys. F 452 Phys. A 451 Phys. A 451 Phys. A 455 Phys. A 653 Phys. A 455 Phys. </td <td>422</td> <td>Ser</td> <td>S</td> <td>393</td> <td>Ser</td> <td>S</td> <td>422</td> <td>Ser</td> <td>S</td> <td>421</td> <td>Ser</td> <td>S</td> <td>602</td> <td>Ser</td> <td>S</td> <td>455</td> <td>Ser</td> <td>S</td>	422	Ser	S	393	Ser	S	422	Ser	S	421	Ser	S	602	Ser	S	455	Ser	S
Πη W 4.25 Τη W 4.53 Τη W 4.53 Τη W 6.53 Λη Α 4.53 Πη W 4.55 Λη 4.55 Λη <t< td=""><td>453</td><td>Phe</td><td>ш</td><td>424</td><td>Phe</td><td>щ</td><td>452</td><td>Tyr</td><td>Y</td><td>451</td><td>Tyr</td><td>Υ</td><td>633</td><td>Tyr</td><td>Υ</td><td>486</td><td>Tyr</td><td>Y</td></t<>	453	Phe	ш	424	Phe	щ	452	Tyr	Y	451	Tyr	Υ	633	Tyr	Υ	486	Tyr	Y
Ala A 426 Ala A 453 Ala A 653 Ala B 653 Ala A 653 Ala Ala Ala Ala	454	Ттр	M	425	Ттр	M	453	Trp	M	452	Ттр	M	634	Тгр	W	487	Trp	W
Alg Alg <td>455</td> <td>Ala</td> <td>Ą</td> <td>426</td> <td>Ala</td> <td>Ą</td> <td>454</td> <td>Ala</td> <td>V</td> <td>453</td> <td>Ala</td> <td>Ą</td> <td>635</td> <td>Asp</td> <td>Ω</td> <td>488</td> <td>Ser</td> <td>S</td>	455	Ala	Ą	426	Ala	Ą	454	Ala	V	453	Ala	Ą	635	Asp	Ω	488	Ser	S
Agg R 429 Agg Lea 1 456 Agg Agg R 457 Agg R 456 Agg Agg Agg Lea 1 457 Lea 1 457 Lea 1 455 Lea 1 457 Lea 1	457	Ala	Ą	428	Ala	Ą	456	Ala	Ą	455	Ala	Ą	637	Val	^	490	Val	>
Leat L 430 Leat L 457 Leat L 458 Glad Q 459 Ala N 450	458	Arg	¥	429	Arg	R	457	Arg	¥	456	Arg	Я	638	Arg	R	491	Arg	R
Pro P 465 Pro P 492 Pro P 493 Pro	459	Leu	Γ	430	Leu	Γ	458	Leu	Γ	457	Leu	Γ	639	Leu	Τ	492	Ile	ı
Gli Q 446 Gli Q 493 Gli Q 675 Gli Q 530 Gli Ser S 446 Ser S 495 Gli G 675 Gli Q 533 Gli Gly G 471 Gly G 499 Asn N 498 Asn N 680 Gly G 533 Gli Asn A 471 Gly G 499 Gly N 680 Gly G 533 Gli Asn A 472 Th N 500 Val N 683 Asn N 534 Asn N	494	Pro	Ь	465	Pro	Ь	493	Pro	Ь	492	Pro	Ы	674	Pro	Д	527	Pro	Ь
Ser Ser 447 Ser 459 Gly G 494 Gly G 676 Gly G 529 Ser Gly M 471 I gly G 499 Asn N 499 Th G 499 Asn N 499 Th N 499 Asn N 499 Th N 499 Asn N 499 Th N 499 Asn N 498	495	Gln	0	466	Gln	0	494	Gln	0	493	Gln	0	675	Gln	0	528	Gln	0
Gly G 471 Gly G 499 Asn N 498 Asn N 680 Gly G 533 Gln Ip As 67 Tp W 500 Tp W 681 Tp W 534 Tp Asn As Asn N 502 Asn N 501 Asn N 683 Asn N 534 Ip Asn N 473 Asn N 502 Asn N 501 Asn N 534 Asn Asn Asn Asn Asn Asn Asn Asn	496	Ser	S	467	Ser	S	495	Gly	Ü	494	Gly	Ð	9/9	Gly	Ō	529	Ser	S
Tip W 472 Tip W 499 Tip W 681 Tip W 534 Tip Ala A 473 Ala A 500 Tip V 500 Ara V 683 Ala N 534 Ala Pro P 502 Ara N 502 Ara N 683 Ara N 536 Ara Asa N 502 Ara N 549 Ara N 683 Ara N 536 Ara Asa N 502 Ara N 549 Ara N 683 Ara N 536 Ara Asa N 522 Ara N 549 Ara N 536 Ara N 536 Ara Ara Asa N 549 Ara N 549 Ara N 548 Ara Ara Ara	200	Gly	Ü	471	Gly	Ğ	499	Asn	Z	498	Asn	Z	089	Gly	Ü	533	Glu	П
Ala A 473 Ala A 501 Val V 682 Ala A 535 Ala Asn N 473 Ala Asn V 500 Val V 684 Po 535 Asn Asn N 543 Asn N 544 Asn N 634 Po 536 Asn N 536 Asn Asn N 536 Asn N 536 Asn As	501	Tr	W	472	Trp	W	200	Trp	W	499	Trp	W	681	Ттр	W	534	Trp	W
Ash N 474 Ash N 501 Asn N 683 Asn N 536 Asn Pro 1 475 Pro P 503 Pro P 684 Pro P 537 Asn Asn N 543 Asn N 544 Asn N 730 Ser P 537 Asn He 1 522 Asn N 549 Bro P 684 Asn N 536 Asn N 537 Asn N 549 Asn N 539 Asn N 539 Asn N 539 Asn N 549 Asn N 539 Asn N 540 Asn N 539 Asn N 540 Asn N 540 Asn N 539 Asn N 480 Asn N 480 Asn N 480 Asn	502	Ala	Ą	473	Ala	Ą	501	Val	>	500	Val	^	682	Ala	A	535	Ala	Ą
Pro P 415 Pro P 502 Pro P 664 Pro P 537 Ala Asin N 521 Asin N 547 Asin N 729 Ser S 533 Ala Asin N 521 Asin N 544 Asin N 729 Ser S 83 Asin Asin N 731 Ile 1 536 Asin N 546 Asin N 547 Asin N 731 Ile 1 731 Ile 1 536 Ile 1 549 Ile 1 731 Ile 1 536 Ile 1 536 Ile 1 536 Ile 1 536 Ile 1 531 Ile	503	Asn	Z	474	Asn	Z	502	Asn	Z	501	Asn	Z	683	Asn	Z	536	Asn	Z
Asn N 51 Asn N 54 Asn N 58 Asn N 78 Th N 78 II N 58 II N 64 Asn A 60 Asn Asn Asn Asn Asn Asn </td <td>504</td> <td>Pro</td> <td>Ь</td> <td>475</td> <td>Pro</td> <td>Ь</td> <td>503</td> <td>Pro</td> <td>Ь</td> <td>502</td> <td>Pro</td> <td>Ы</td> <td>684</td> <td>Pro</td> <td>Д</td> <td>537</td> <td>Ala</td> <td>A</td>	504	Pro	Ь	475	Pro	Ь	503	Pro	Ь	502	Pro	Ы	684	Pro	Д	537	Ala	A
Asn N 522 Asn N 548 Asn N 558 Ile 1 550 Ile 4 602 Ala A 78 Ile 1 640 Ala Ala Ala Ala Ala Ala Ala Ala Ala Ala </td <td>550</td> <td>Asn</td> <td>Z</td> <td>521</td> <td>Asn</td> <td>Z</td> <td>548</td> <td>Asn</td> <td>Z</td> <td>547</td> <td>Asn</td> <td>Z</td> <td>729</td> <td>Ser</td> <td>S</td> <td>583</td> <td>Asn</td> <td>z</td>	550	Asn	Z	521	Asn	Z	548	Asn	Z	547	Asn	Z	729	Ser	S	583	Asn	z
He I 523 He I 549 He I 731 He 731 He 733 He 733 He 633 Th Ala A 602 Tip W 601 Tip W 783 Tip W 639 Tip Glu G 573 Glu E 604 Glu	551	Asn	Z	522	Asn	Z	549	Asn	Z	248	Asn	Z	730	Asn	Z	584	Asn	z
Tip W 575 Tip W 601 Tip W 601 Tip W 601 Tip W 783 Tip W 639 Tip Ala A 602 Ala A 602 Ala A 784 His H 640 Asp Glu G 604 Gly G Gly G 787 Tip T 641 Asp Ser S 579 Ser S 605 Ser S 605 Ser Ser Ser Ser 787 Ser Ser G <td< td=""><td>552</td><td>lle</td><td>_</td><td>523</td><td>lle</td><td>П</td><td>550</td><td>Ile</td><td>_</td><td>549</td><td>Ile</td><td>_</td><td>731</td><td>Ile</td><td>_</td><td>585</td><td>lle</td><td>1</td></td<>	552	lle	_	523	lle	П	550	Ile	_	549	Ile	_	731	Ile	_	585	lle	1
Ala A sold Ala Ala Ala A sold Ala	604	Тrр	×	575	Trp	×	602	Trp	×	601	Ттр	M	783	Trp	M	639	Trp	M
Gly G 577 Gly G 64 Gly G 604 Gly G 6 Gly G 785 Thr T 641 Ser Ser S 579 Ser S 605 Gly E 604 Gly E 786 Gly E 642 Glu E 642 Glu Ser S 579 Ser S 605 Gly S 67 S 67 S 67 S 67 Ala A 612 Ala A 612 Ala A 639 Ala A 638 Ala A 638 Ala A 638 Ala A 638 Ala A 639 Thr A 638 Ala A 638 Al	905	Ala	Ą	276	Ala	Ą	603	Ala	A	602	Ala	Ą	784	His	Η	<u>\$</u>	Asp	D
Glu E 578 Glu E 604 Glu E 604 Glu E 604 Glu E 786 Glu E 642 Glu Ser	909	Gly	ŋ	277	Gly	ŋ	604	Gly	ŋ	603	Gly	Ð	785	Thr	L	<u>\$</u>	Ser	ß
Ser S 579 Ser S 665 Ser S 665 Ser S 643 Ser Trp W 611 Trp W 631 Trp W 819 Trp W 675 Trp Arb A 612 Arb A 638 Trp W 819 Trp W 675 Trp Pro A 613 Arb A 639 Trp A 673 Trp W 673 Trp W 673 Trp A 673 Trp	209	Glu	П	578	Glu	ш	605	Glu	ш	604	Glu	Щ	786	Glu	ш	42	Glu	ш
Trp W 611 Trp W 638 Trp W 637 Trp W 631 Trp W 637 Trp W 631 Trp W 675 Trp M 675 Trp M 675 Trp Free 61	809	Ser	S	579	Ser	S	909	Ser	S	905	Ser	S	787	Ser	S	23	Ser	S
Ala A 612 Ala A 639 Ala A 638 Ala A 639 A B A 639 A B	640	Ттр	M	611	Тр	M	638	Trp	M	637	Тгр	Μ	819	Тгр	M	675	Trp	M
Thr T 613 Thr T 640 Thr T 639 Thr T 821 Thr T 677 Thr Pro P 642 Pro P 641 Pro P 823 Pro P 679 Pro Arg R 643 Arg R 642 Arg R 824 Arg R 680 Arg Pro F 641 Pro L 642 Arg R 824 Arg R 680 Arg Pro F 654 Pro L 643 Leu L 643 Pro	641	Ala	K	612	Ala	¥	639	Ala	Ą	859	Ala	Ą	820	Ala	Α	9/9	Ala	Ą
Pro P 615 Pro P 641 Pro P 642 Arg R 824 Arg R 680 Arg R 824 Arg R 680 Arg P 681 Pro F 680 Pro F 681 Pro Pro R 824 Pro P 681 Pro P 682 Pro P 683 Pro P 683 Pro P 864 Pro P 722 Pro Iyr Y 661 Iyr Y 681 Iyr Y 863 Iyr X 127 Iyr Y	642	Thr	L	613	Thr	Τ	640	Thr	Τ	639	Thr	Τ	821	Thr	L	<i>LL</i> 9	Thr	L
Arg R 616 Arg R 643 Arg R 644 Arg R 643 Arg R 824 Arg R 680 Arg Len L 617 Len L 644 Len L 643 Len L 681 Len Phe F 654 Phe F 680 Phe F 862 Phe F 671 Phe Gln Q 652 Gln Q 681 Gln Q 863 Gln Q 721 Gln Phe Pro 656 Pro P 683 Pro P 864 Pro P 722 Pro Iyr Y 661 Iyr Y 869 Iyr Y 727 Iyr	644	Pro	Ь	615	Pro	Ь	642	Pro	Ь	24	Pro	Ь	823	Pro	Ы	619	Pro	Ь
Len L 617 Leu L 644 Leu L 643 Leu L 825 Leu L 681 Leu Phe F 654 Phe F 681 Phe F 680 Phe F 862 Phe F 720 Phe Gln Q 655 Gln Q 682 Gln Q 681 Gln Q 863 Gln Q 721 Gln Pro P 656 Pro P 683 Pro P 682 Pro P 864 Pro P 722 Pro Tyr Y 661 Tyr Y 688 Tyr Y 687 Tyr Y 869 Tyr Y 727 Tyr	645	Arg	ĸ	919	Arg	R	643	Arg	R	642	Arg	R	824	Arg	~	089	Arg	R
Phe F 654 Phe F 680 Phe F 682 Phe F 720 Phe Gln Q 655 Gln Q 681 Gln Q 863 Gln Q 721 Gln Pro P 656 Pro P 683 Pro P 864 Pro P 722 Pro Tyr Y 661 Tyr Y 687 Tyr Y 889 Tyr Y 727 Tyr	646	Leu	L	617	Leu	Γ	\$	Leu	Γ	£3	Leu	J	825	Leu	J	681	Leu	Γ
Gla Q 655 Gla Q 682 Gla Q 681 Gla Q 863 Gla Q 721 Pro P 656 Pro P 683 Pro P 682 Pro P 864 Pro P 722 Tyr Y 661 Tyr Y 688 Tyr Y 687 Tyr Y 869 Tyr Y 727	683	Phe	ш	654	Phe	щ	681	Phe	ш	089	Phe	щ	862	Phe	щ	720	Phe	Ŧ
Pro P 656 Pro P 683 Pro P 682 Pro P 722 Tyr Y 661 Tyr Y 687 Tyr Y 869 Tyr Y 727	684	Gln	0	655	Gln	o	682	Gln	ø	681	Gln	0	863	Gln	0	721	Gln	0
Tyr Y 661 Tyr Y 688 Tyr Y 687 Tyr Y 869 Tyr Y 727 .	685	Pro	Ь	959	Pro	Ь	683	Pro	Ь	682	Pro	Ы	864	Pro	Ы	722	Pro	Ь
	069	Tyr	Y	961	Tyr	Υ	889	Tyr	Y	289	Tyr	Υ	698	Tyr	Υ	727	Tyr	Y

TABLE 6

TABLE 6-continued

	Amino acid (1-letter code)	W	Ą	0	Ą	Γ	I	щ
N4	Amino Amino acid acid (3-letter position code)			730 Gln				
	Amino acid An 1-letter ac code) posi	W 7.	7	E 7.	V 7.	L 7	L 7	. L
KSM635	Amino Amino acid acid (3-letter (1 position code)			872 Glu				
	Amino acid (1-letter code) p	W	>	0	Ą	Σ	I	I
64	Amino Amino acid acid (3-letter position code)			690 Gln				
	Amino acid (1-letter code)	W	^	0	¥	Σ	_	_
1139	Amino Amino acid acid (3-letter position code)			691 Gln				
İ	Amino acid (1-letter code)	W	Λ	0	¥	M	I	Ι
DSM12648	Amino Amino acid acid (3-letter position code)	662 Trp		664 Gln				712 Ile
	Amino acid (1-letter code)	W	^	0	٧	Σ	ı	I
S237	Amino acid Amino acid (3-letter position code)	Тгр	Val	Gln	Ala	Met	Ile	Ile
	Amino acid position	691	692	693	694	739	740	741

In the column for "S237" in Table 6, 44 amino acid residues that may be considered as targets of substitution in the cellulose binding module of S237 cellulase, are shown. Furthermore, Table 6 shows amino acid residues, of other alkaline cellulases sharing high amino acid sequence identity with 5 S237 cellulase, Bacillus sp. strain DSM12648-derived alkaline cellulase (DSM12648 cellulase; SEQID NO: 4), Bacillus sp. strain 1139-derived alkaline cellulase (1139 cellulase; SEQ ID NO: 6), Bacillus sp. strain KSM-64-derived alkaline cellulase (endo-1,4- β -glucanase) (64 cellulase; SEQ ID NO: 10 8), Bacillus sp. strain KSM-635-derived alkaline cellulase (KSM-635 cellulase; SEQ ID NO: 10), and Bacillus sp. strain N-4-derived alkaline cellulase (endoglucanase) (N4 cellulase; SEQ ID NO: 12), which are aligned to the 44 amino acid residues of S237 cellulase (that is, present at the positions 15 corresponding to those residues) when an alignment of the amino acid sequence of each of the cellulases and the amino acid sequence of S237 cellulase is produced. Each of the positions of the amino acid residues is indicated with the number of the amino acid residue in the amino acid sequence 20 of the alkaline cellulase set forth in the corresponding sequence ID number.

In Examples 5 and 6 described below, mutant S237 cellulases in which some amino acid residues among such amino acid residues that participate in cellulose binding (in Table 6, 25 underlined residues, namely, the amino acid residues at the positions 419, 421, 454 and 501) were substituted with other amino acid residues, were constructed.

Example 1

Production of Mutant S237 Cellulase-1

A mutant S237 cellulase in which glutamine at position 58 of S237 cellulase (SEQ ID NO: 2) was substituted with arginine (S237-Q58R) was produced by introducing a nucleotide mutation to the S237 cellulase gene and recombinantly expressing the mutant, as follows.

PCR amplification was carried out according to the section "1) Amplification of DNA fragment", using, as a template, the 40 genomic DNA extracted from Bacillus sp. strain KSM-S237 (FERM BP-7875) by a routine method, and using a primer set consisting of primers 237UB1 and Q58R-RV and a primer set consisting of primers Q58R-FW and S237RV indicated in the above Table 2-3. As a result, a 0.7-kb amplified DNA frag- 45 ment which includes a nucleotide mutation introduced at the position corresponding to the amino acid residue at position 58 on the S237 cellulase gene (SEQ ID NO: 1) in the vicinity of the 3'-terminus and a region that is mainly in the upstream of the mutation position, and a 2.5-kb amplified DNA frag- 50 ment which includes the nucleotide mutation in the vicinity of the 5'-terminus and a region that is mainly in the downstream of the mutation position, were obtained. The base sequence of the primer Q58R-RV thus used was designed based on the base sequence of the S237 cellulase gene, wherein the nucleotide mutation for substituting glutamine at position 58 of S237 cellulase with arginine is included. The base sequence of the primer Q58R-FW is a complementary sequence of the primer Q58R-RV.

Subsequently, the two fragments thus obtained were mixed 60 to be used as templates, and SOE (splicing by Overlap Extension)-PCR (Horton R. M. et al., Gene (1989) 77(1), p. 61-68) was carried out according to the section "1) Amplification of DNA fragment", using a primer set consisting of 237UB1 and S237RV indicated in Table 1. Thus, a 3.2-kb DNA fragment 65 in which those two fragments are linked through a complementary sequence was obtained.

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The 3.2-kb DNA fragment thus obtained (mutant S237 cellulase gene) was inserted at the SmaI restriction enzyme cleavage point of a shuttle vector pHY300PLK, and thus a recombinant plasmid pHY-S237 Q58R was constructed. The base sequence of the mutant S237 cellulase gene inserted in the plasmid was confirmed by determining the sequence using a 3100 DNA Sequencer (Applied Biosystems, Inc.). Subsequently, a transformant obtained by introducing the recombinant plasmid pHY-S237_Q58R into Bacillus subtilis by the method according to the section "2) Gene introduction into Bacillus subtilis", was cultured. From the culture thus obtained, a recombinantly produced protein was isolated and purified by the same method as in the section "5) 1. Recombinant production of S237 cellulase", and quantification was carried out. This recombinant protein is a mutant S237 cellulase in which glutamine at position 58 of the amino acid sequence of S237 cellulase (SEQ ID NO: 2) has been substituted with arginine (hereinafter, also referred to as S237_Q58R). An enzyme sample containing the mutant S237 cellulase thus obtained was used in the evaluation of antiredeposition ability in Test Example 1 described below.

Example 2

Production of Mutant S237 Cellulase-2

A mutant S237 cellulase, S237_Q242S, in which glutamine at position 242 of S237 cellulase (SEQ ID NO: 2) was substituted with serine, was produced by the same 30 method as described in Example 1. That is, the upstream region of the S237 cellulase gene (SEQ ID NO: 1) containing the intended nucleotide mutation (region in the upstream of the vicinity of the mutation position), and the downstream region of the S237 cellulase gene (SEQ ID NO: 1) containing the intended nucleotide mutation (region in the downstream of the vicinity of the mutation position) were amplified by PCR, using a primer set consisting of 237UB1 and Q242S-RV and a primer set consisting of Q242S-FW and S237RV indicated in Table 2-3. Two DNA fragments thus obtained were used as templates, and thus a 3.2-kb DNA fragment containing the intended mutant S237 cellulase gene was amplified. The DNA fragment was inserted into a shuttle vector pHY300PLK, and thus a recombinant plasmid pHY-S237_Q242S was constructed. The base sequence of the mutant S237 cellulase gene inserted into the plasmid was confirmed, and a transformant obtained by introducing the recombinant plasmid into Bacillus subtilis by the method according to the section "2) Gene introduction into Bacillus subtilis" was cultured. A recombinant protein (that is, mutant S237 cellulase S237_Q242S) was isolated and purified from the culture thus obtained, and quantification of the recombinant protein was carried out.

Based on this mutant S237 cellulase S237_Q242S, two kinds of S237 cellulase double mutant, in which glutamine at position 58 was substituted with arginine or glutamic acid (QS_Q58R and QS_Q58E, respectively), were produced basically in the same manner as described in Example 1. First, two DNA fragments were obtained by performing PCR amplification of the upstream region (region in the upstream of the vicinity of the mutation position) of the mutant S237 cellulase S237_Q242S gene containing the intended nucleotide mutation in the vicinity of the 3'-terminal, and the downstream region (region in the downstream of the vicinity of the mutant S237 cellulase S237_Q242S gene containing the intended nucleotide mutation position) of the mutant S237 cellulase S237_Q242S gene containing the intended nucleotide mutation, using the pHY-S237_Q2428 produced as described above as a template DNA, and respectively using two pairs of primer sets for

introducing the intended mutations indicated in the above Table 2-3. Using the two DNA fragments thus obtained as templates, a DNA fragment including the intended mutant S237 cellulase gene was amplified. The DNA fragment was inserted into a shuttle vector pHY300PLK to construct a 5 recombinant plasmid, and the base sequence of the mutant S237 cellulase gene inserted in the plasmid was confirmed. A transformant obtained by introducing the recombinant plasmid into Bacillus subtilis by the method according to the section "2) Gene introduction into Bacillus subtilis", was 10 cultured. From the culture thus obtained, a recombinant protein (mutant S237 cellulase) was isolated and purified by the same method as in the section "5) 1. Recombinant production of S237 cellulase", and quantification of the recombinant protein was carried out. Enzyme samples containing the 15 respective mutant S237 cellulases thus obtained were used in an evaluation of the anti-redeposition ability and an evaluation of stability in Test Examples 2 and 3 described below.

Example 3

Production of Mutant S237 Cellulase-3

A mutant S237 cellulase in which serine at position 193 of S237 cellulase (SEQ ID NO: 2) was substituted with arginine, 25 was produced by introducing a nucleotide mutation into the S237 cellulase gene, and recombinantly expressing the mutant, as follows.

PCR amplification was carried out according to the section "1) Amplification of DNA fragment", using the genomic 30 DNA extracted from *Bacillus* sp. strain KSM-S237 (FERM BP-7875) by a routine method as a template, and using a primer set consisting of primers 237UB1 and S193R-RV and a primer set consisting of primers S193R-FW and S237RV indicated in the above Table 2-1. As a result, a 0.7-kb ampli- 35 fied DNA fragment which includes a nucleotide mutation introduced at the position corresponding to the amino acid residue at position 193 on the S237 cellulase gene (SEQ ID NO: 1) in the vicinity of the 3'-terminus and a region that is mainly in the upstream of the mutation position, and a 2.5-kb 40 amplified DNA fragment which includes the nucleotide mutation in the vicinity of the 5'-terminus and a region that is mainly in the downstream of the mutation position, were obtained. The base sequence of the primer S193R-RV thus used was designed based on the base sequence of S237 cel- 45 lulase gene, wherein a nucleotide mutation for substituting serine at position 193 of S237 cellulase with arginine is included. The base sequence of the primer S193R-FW is a complementary sequence of the primer S193R-RV.

Subsequently, the two fragments thus obtained were mixed 50 to be used as templates, and SOE (splicing by Overlap Extension)-PCR (Horton R. M. et al., Gene (1989) 77(1), p. 61-68) was carried out according to the section "1) Amplification of DNA fragment", using a primer set consisting of 237UB1 and S237RV indicated in Table 1. Thus, a 3.2-kb DNA fragment 55 in which those two fragments are linked through a complementary sequence was obtained.

The 3.2-kb DNA fragment thus obtained (mutant S237 cellulase gene) was inserted at the SmaI restriction enzyme cleavage point of a shuttle vector pHY300PLK, and thus a 60 recombinant plasmid pHY-S237(S193R) was constructed. The base sequence of the mutant S237 cellulase gene inserted in the plasmid was confirmed by determining the sequence using a 3100 DNA Sequencer (Applied Biosystems, Inc.). Subsequently, a transformant obtained by introducing the 65 recombinant plasmid pHY-S237(S193R) into *Bacillus subtilis* by the method according to the section "2) Gene introduc-

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tion into *Bacillus subtilis*", was cultured. From the culture thus obtained, a recombinantly produced protein was isolated and purified by the same method as in the section "5) 1. Recombinant production of S237 cellulase", and quantification was carried out. This recombinant protein is a mutant S237 cellulase in which serine at position 193 of the amino acid sequence of S237 cellulase (SEQ ID NO: 2) had been substituted with arginine (hereinafter, also referred to as S237_S193R). An enzyme sample containing the mutant S237 cellulase thus obtained, S237 S193R, was used in the evaluation of anti-redeposition ability in Test Example 4 described below.

Example 4

Production of Mutant S237 Cellulase-4

A mutant S237 cellulase in which the non-charged amino acid residues selected as described above were substituted 20 with charged amino acid residues in the mutant S237 cellulase described in Example 2, S237_Q242S, was produced. Specifically, a S237 cellulase double mutant (QS_Q71E) in which glutamine at position 71 of the amino acid sequence of the mutant S237 cellulase S237_Q242S had been substituted with glutamic acid, was produced basically in the same manner as described in Example 3. First, two DNA fragments were obtained by performing PCR amplification of the upstream region (region in the upstream of the vicinity of the mutation position) of the mutant S237 cellulase S237_Q242S gene containing the intended nucleotide mutation in the vicinity of the 3'-terminal, and the downstream region (region in the downstream of the vicinity of the mutation position) of the mutant S237 cellulase S237_Q242S gene containing the intended nucleotide mutation, using the pHY-S237 (Q242S) produced as described above as a template DNA, and respectively using two pairs of primer sets for introducing the intended mutations indicated in the above Table 2-1. Using the two DNA fragments thus obtained as templates, a DNA fragment including the intended mutant S237 cellulase gene was amplified. That DNA fragment was inserted into a shuttle vector pHY300PLK to construct a recombinant plasmid, and the base sequence of the mutant S237 cellulase gene inserted in the plasmid was confirmed. A transformant obtained by introducing the recombinant plasmid into Bacillus subtilis by the method according to the section "2) Gene introduction into Bacillus subtilis", was cultured. From the culture thus obtained, a recombinant protein (mutant S237 cellulase) was isolated and purified by the same method as in the section "5) 1. Recombinant production of S237 cellulase", and quantification of the recombinant protein was carried out. Enzyme samples containing the respective mutant S237 cellulases thus obtained were used in an evaluation of the anti-redeposition ability and an evaluation of stability in Test Example 5 described below.

Example 5

Production of Mutant S237 Cellulase-5

Three kinds of mutant S237 cellulases, in which the amino acid residues participating in cellulose binding were substituted with introducing nucleotide mutations that cause intended amino acid substitutions into the S237 cellulase gene, and recombinantly expressing the mutants, were produced. Specifically, mutant S237 cellulases in which aspartic acid at position 421 of S237 cellulase (SEQ ID NO: 2) was substituted with alanine, tryptophan at position 454 substi-

tuted with tyrosine, and tryptophan at position 501 substituted with tyrosine (S237_D421A, S237_W454Y, and S237_W501Y, respectively) were produced.

For each of the mutant cellulases, two DNA fragments were obtained by performing PCR amplification of the 5 upstream region (region in the upstream of the vicinity of the mutation position) of the mutant S237 cellulase gene containing the intended nucleotide mutations, and the downstream region (region in the downstream of the vicinity of the mutation position) of the mutant S237 cellulase gene containing the intended nucleotide mutations, respectively using two pairs of primer sets for introducing the intended mutations (see Table 2-2). Using the two DNA fragments thus obtained as templates, a DNA fragment including the intended mutant S237 cellulase gene was amplified. That DNA fragment was inserted into a shuttle vector pHY300PLK to construct a recombinant plasmid, and the base sequence of the mutant S237 cellulase gene inserted in the plasmid was confirmed. A transformant obtained by introducing the recombinant plasmid into Bacillus subtilis by the method according to the 20 section "2) Gene introduction into Bacillus subtilis", was cultured. From the culture thus obtained, a recombinant protein (that is, each mutant S237 cellulase) was extracted and purified by the same method as in the section "5) 1. Recombinant production of S237 cellulase", and quantification of 25 cellulase was observed. the recombinant protein was carried out. Enzyme samples containing the respective mutant S237 cellulases thus obtained were used in an evaluation of the anti-redeposition ability in Test Example 6 described below.

Example 6

Production of Mutant S237 Cellulase-6

A mutant S237 cellulase, in which the amino acid residues 35 participating in cellulose binding were further substituted in the mutant S237 cellulase S237 Q242S described in Example 2, was produced. Specifically, a S237 cellulase double variant (QS_N419A) in which asparagine at position 419 of the amino acid sequence of the mutant S237 cellulase 40 S237_Q242S was substituted with alanine, was produced basically in the same manner as described in Example 5. That is, two DNA fragments were obtained by performing PCR amplification of the upstream region (region in the upstream of the vicinity of the mutation position) of the mutant S237 45 cellulase S237_Q242S gene containing the intended nucleotide mutation, and the downstream region (region in the downstream of the vicinity of the mutation position) of the mutant S237 cellulase S237_Q242S gene containing the intended nucleotide mutation, using the pHY-S237(Q242S) 50 produced as described above as a template DNA and respectively using two pairs of primer sets for introducing the intended mutations (see Table 2-2). Using the two DNA fragments thus obtained as templates, a DNA fragment including the intended mutant S237 cellulase gene was amplified. That 55 DNA fragment was inserted into a shuttle vector pHY300PLK to construct a recombinant plasmid, and the base sequence of the mutant S237 cellulase gene inserted in the plasmid was confirmed. A transformant obtained by introducing the recombinant plasmid into Bacillus subtilis by the 60 method according to the section "2) Gene introduction into Bacillus subtilis", was cultured. From the culture thus obtained, a recombinant protein (that is, each mutant S237 cellulase) was isolated and purified by the same method as in the section "5) 1. Recombinant production of S237 cellulase", and quantification of the recombinant protein was carried out. Enzyme samples containing the respective mutant

S237 cellulases thus obtained were used in an evaluation of the anti-redeposition ability in Test Example 7 described below.

Test Example 1

Evaluation of Anti-Redeposition Ability of Mutant S237 Cellulase-1

The anti-redeposition ability of mutant S237 cellulase S237_Q58R produced in Example 1 was evaluated according to the section "4) Evaluation of anti-redeposition ability" described above. Furthermore, for the evaluation, 50 g of a white knitted cotton cloth [seared bleached cloth (supplied by Tanigashira Shoten) that had been washed and then sufficiently rinsed] was introduced, such that the amount of cloth with respect to the solution (bath ratio) would be 10 L/kg. The detergent composition B described above was used, and an amount of enzyme equivalent to the amount of protein exhibiting 52.8 mU of the wild-type S237 cellulase activity with respect to the S237_Q58R, was used. The results obtained are shown in Table 7. Enhanced anti-redeposition ability of the mutant S237 cellulase S237_Q58R as compared with S237 cellulase was observed.

TABLE 7

	Enzyme	Effect of mutation on anti-redeposition (%)
30	Control S237_Q58R	0 2.77

Test Example 2

Evaluation of Anti-Redeposition Ability of Mutant S237 Cellulase-2

The anti-redeposition ability of the mutant S237 cellulases QS_Q58R and QS_Q58E that had been produced in Example 2 using S237_Q242S as the parent cellulase, was evaluated according to the section "4) Evaluation of anti-redeposition ability" described above. Furthermore, upon the evaluation, assuming the co-presence of sebum dirt components, three sheets of a stained cloth wfk10D (Wfk Testgewebe GmbH (D41379, Germany)) each having a size of 6 cm×6 cm were added. A cleaning agent of the composition C was used, and an amount of enzyme equivalent to the amount of protein exhibiting 52.8 mU of the S237_Q242S cellulase activity was used. In the present evaluation, S237_Q242S was used as a control enzyme. The results thus obtained are presented in Table 8. For any of the mutant S237 cellulases evaluated and used, a high anti-redeposition effect was obtained as compared with the control enzyme (S237_Q242S), and the enhanced anti-redeposition ability by mutagenesis was observed.

TABLE 8

Enzyme	Effect of mutation on anti-redeposition (%)
Control	0
QS_Q58E	3.03
Control	0
QS_Q58R	8.92

Test Example 3

Stability Test of Mutant S237 Cellulase in Liquid Cleaning Agent

Stability was evaluated for the case where the mutant S237 cellulase QS_Q58R produced by using S237_Q242S as a parent cellulase in Example 3 was stored in a cleaning agent of the composition E to which various alkaline proteases were added.

As the alkaline proteases, alkaline proteases KP43 (Japanese Patent No. 3479509), KannaseTM (Novozymes, Inc.) and ProperaseTM (Danisco AS) were used. These three kinds of alkaline proteases are all subtilisin-like alkaline proteases suitable to be incorporated into cleaning agents.

To 450 μ L of the cleaning agent of the composition E, 460 U/L of a cellulase (S237QS or S237QS_Q58R) and 0.012 g each, as an amount of protein, of the three kinds of alkaline proteases described above (an amount of protein equivalent to 20 the addition of 29 U/L for KP43) were added, and liquid amount of the samples were adjusted to 500 μ L, and were stored at 40° C. The residual cellulase activity in the cleaning agent after 24 hours was measured.

The cellulase activity was measured by the method of using $\,^{25}$ p-nitrophenyl- β -D-cellotrioside described in Example 1, and the residual cellulase activity was calculated by the following formula.

Residual cellulase activity (%)=(Cellulase activity after 24 hours of storage/cellulase activity immediately after preparation)×100 [Mathematical formula 8]

The experimental results are presented in FIG. 4. The relative residual activity of QS_Q58R is shown by defining the residual activity value of S237_QS after storage for 24 hours 35 at 40° C. as 100. For all of the systems to which the three kinds of proteases were added, QS_Q58R exhibited higher residual activity as compared with S237QS. From these results, enhancements of the anti-redeposition ability and the protease resistance by substituting glutamine at position 58 of 40 S237 cellulase with arginine were observed.

Test Example 4

Evaluation of Anti-Redeposition Ability of Mutant S237 Cellulase-3

The anti-redeposition ability of the mutant S237 cellulase S237_S193R produced in Example 3 was evaluated according to the section "4) Evaluation of anti-redeposition ability" 50 described above. Furthermore, for the evaluation, 50 g of a white knitted cotton cloth [seared bleached cloth (supplied by Tanigashira Shoten) that had been washed and then sufficiently rinsed] was introduced, such that the amount of cloth with respect to the solution (bath ratio) would be 10 L/kg. A 55 cleaning agent of the composition F described above was used, and an amount of enzyme equivalent to the amount of protein exhibiting 52.8 mU of the wild-type S237 cellulase activity with respect to the S237_S193R, was used. Asahi carbon black for cleaning was used as the carbon black, and 60 the hardness of the used water was adjusted to 12° DH. The results thus obtained are shown in Table 9. Furthermore, the value of solvent accessibility determined in Example 2 is also presented. With S237_S193R, a higher anti-redeposition effect as compared with the control enzyme (S237_Q242S) 65 was provided, and enhancement of the anti-redeposition ability by the mutagenesis was observed.

60 TABLE 9

Mutant S237 Degree of anti-redeposition ability enhancement in mutant cellulase (%) Solvent accessibility of residue at mutation position S237_S193R 3.26 76.08

Test Example 5

Evaluation of Anti-Redeposition Ability of Mutant S237 Cellulase-4

The anti-redeposition ability of the mutant S237 cellulase QS_Q71E produced in Example 4 using S237_Q242S as a base, was evaluated according to the section "4) Evaluation of anti-redeposition ability" described above.

A cleaning agent of the composition C described above was used, and an amount of enzyme equivalent to the amount of protein exhibiting 52.8 mU of the wild-type S237 cellulase activity with respect to QS Q71E, was used. For the present evaluation, S237 Q242S was used as a control enzyme. Asahi carbon black for cleaning was used as the carbon black, and the hardness of the used water was adjusted to 4° DH. Furthermore, upon the evaluation, assuming the co-presence of sebum dirt components, three sheets of a stained cloth wfk10D (Wfk Testgewebe GmbH (D41379, Germany)) each having a size of 6 cm×6 cm were added. The results thus obtained are shown in Table 10. Further, the value of the solvent accessibility of residues determined in the section "6) Steric structure modeling of S237 cellulase" described above is also shown. Higher anti-redeposition effect as compared with the control enzyme (S237_Q242S) was provided in QS_Q71E, and enhancement of the anti-redeposition ability by the mutagenesis was observed. From the results of the present Test Example and the Test Example 4, it was demonstrated that the anti-redeposition ability of the mutant S237 cellulases can be enhanced by substituting the non-charged amino acid residues selected as described above, with charged amino acid residues. The result suggests that such amino acid substitution caused high repulsion between the enzyme surface and hydrophobic carbon black by further increasing the hydrophilicity of the enzyme surface, as specu-45 lated in the above, and thereby a higher anti-redeposition effect was provided.

TABLE 10

Mutant S237 cellulase	Degree of anti-redeposition ability enhancement in mutant cellulase (%)	Solvent accessibility of residue at mutation position
QS_Q71E	6.06	116.3

Test Example 6

Evaluation of Anti-Redeposition Ability of Mutant S237 Cellulase-5

The anti-redeposition ability of the mutant S237 cellulases produced in Example 5, S237_D421A, S237_W454Y and S237_W501Y, was evaluated according to the section "4) Evaluation of anti-redeposition ability" described above. For the evaluation, 50 g of a white knitted cotton cloth [seared bleached cloth (supplied by Tanigashira Shoten) that had been washed and then sufficiently rinsed] was introduced,

such that the amount of cloth with respect to the solution (bath ratio) would be 10 L/kg. A cleaning agent of the composition B described above was used, and an amount of enzyme equivalent to the amount of protein exhibiting 52.8 mU of the wild-type S237 cellulase activity with respect to each of the 5 mutant S237 cellulases, was used. In the present evaluation, the wild-type S237 cellulase was used as a control enzyme. Asahi carbon black for cleaning was used as the carbon black, and the hardness of the used water was adjusted to 12° DH. The results thus obtained are shown in Table 11. For all of the 10 mutant S237 cellulases, an enhancement of the anti-redeposition ability as compared with S237 cellulase was observed. Therefore, it was demonstrated that when the amino acid residues that participate in cellulose binding in the cellulose binding module of an alkaline cellulase are substituted with 15 other amino acid residues, the anti-redeposition ability of a mutant S237 cellulase can be enhanced. The result suggests that such amino acid substitution decreased the cellulose binding property of the alkaline cellulase as speculated in the above, and thereby a higher anti-redeposition effect was pro- 20

TABLE 11

vided.

Mutant S237 cellulase	Degree of anti-redeposition ability enhancement in mutant cellulase (%)
S237_D421A	3.29
S237_W454Y	4.21
S237_W501Y	10.5

Test Example 7

Evaluation of Anti-Redeposition Ability of Mutant S237 Cellulase-6

The anti-redeposition ability of the mutant S237 cellulase QS_N419A produced in Example 6 using S237_Q242S as a

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base, was evaluated according to the section "4) Evaluation of anti-redeposition ability" described above. For the evaluation, 45 g of a white knitted cotton cloth [seared bleached cloth (supplied by Tanigashira Shoten) that had been washed and then sufficiently rinsed] was introduced, such that the amount of cloth with respect to the solution (bath ratio) would be 11 L/kg. A cleaning agent of the composition C described above was used, and an amount of enzyme equivalent to the amount of protein exhibiting 52.8 mU of the wild-type S237 cellulase activity with respect to QS_N419A, was used. In the present evaluation, S237_Q242S was used as a control enzyme. Asahi carbon black for cleaning was used as the carbon black, and the hardness of the used water was adjusted to 4° DH. Furthermore, upon the evaluation, assuming the co-presence of sebum dirt components, three sheets of a stained cloth wfk10D (Wfk Testgewebe GmbH (D41379, Germany)) each having a size of 6 cm×6 cm were added. The results thus obtained are shown in Table 12. Higher antiredeposition effect as compared with the control enzyme (S237 Q242S) was provided in QS N419A, and enhancement of the anti-redeposition ability by the mutagenesis was observed. That is, it was demonstrated that when the amino acid residues that participate in cellulose binding in the cellulose binding module of an alkaline cellulase are substituted with other amino acid residues, the anti-redeposition ability of a mutant S237 cellulase can be enhanced. The result suggests that such amino acid substitution decreased the cellulose binding property of the alkaline cellulase as speculated in the above, and thereby a higher anti-redeposition effect was 30 provided.

TABLE 12

5	Mutant S237 cellulase	Degree of anti-redeposition ability enhancement in mutant cellulase (%)
	QS_N419A	4.62

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Lys Arg Pro Sei 50	r Glu Ala Gly 55	y Ala Leu Gln	Leu Gln Glu 60	Val Asp Gly	
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D	7	m	a -	a1.	7	D	7	Ŧ	77.	7.7	7	7	D	T 1	7	

Pro Asn Trp Ser Gln Arg Pro Asp Leu Ala Ala Asp Asn Pro Ile Asn

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_			_		_	_			ttt Phe 740	_	-	-	_	_	_	2440	
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Lys Arg Pro Ser Glu Ala Gly Ala Leu Gln Leu Gln Glu Val Asp Gly 50 55 60	
Gln Met Thr Leu Val Asp Gln His Gly Glu Lys Ile Gln Leu Arg Gly 65 70 75 80	
Met Ser Thr His Gly Leu Gln Trp Phe Pro Glu Ile Leu Asn Asp Asn 85 90 95	
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Lys Ser Arg Val Ile Lys Gly Ile Asp Leu Ala Ile Glu Asn Asp Met 130 135 140	
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Pro Val Tyr Ala Gly Ala Glu Asp Phe Phe Arg Asp Ile Ala Ala Leu 165 170 175	
Tyr Pro Asn Asn Pro His Ile Ile Tyr Glu Leu Ala Asn Glu Pro Ser 180 185 190	
Ser Asn Asn Asn Gly Gly Ala Gly Ile Pro Asn Asn Glu Glu Gly Trp 195 200 205	
Asn Ala Val Lys Glu Tyr Ala Asp Pro Ile Val Glu Met Leu Arg Asp 210 215 220	
Ser Gly Asn Ala Asp Asp Asn Ile Ile Ile Val Gly Ser Pro Asn Trp 225 230 235 240	
Ser Gln Arg Pro Asp Leu Ala Ala Asp Asn Pro Ile Asp Asp His His	
245 250 255	

Thr Met Tyr Thr Val His Phe Tyr Thr Gly Ser His Ala Ala Ser Thr

Ser Tyr Pro Pro Glu Thr Pro Asn Ser Glu Arg Gly Asn V	la Thr he Asp 320 er Trp 35
275 280 285 Ser Asn Thr Arg Tyr Ala Leu Glu Asn Gly Val Ala Val Phe A 290 300 Glu Trp Gly Thr Ser Gln Ala Asn Gly Asp Gly Gly Pro Tyr P 305 Glu Ala Asp Val Trp Ile Glu Phe Leu Asn Glu Asn Asn Ile S	la Thr he Asp 320 er Trp 35
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305 310 315 Glu Ala Asp Val Trp Ile Glu Phe Leu Asn Glu Asn Asn Ile S	320 er Trp 35
	35
	la o Tilano
Ala Asn Trp Ser Leu Thr Asn Lys Asn Glu Val Ser Gly Ala P	ne mi
Pro Phe Glu Leu Gly Lys Ser Asn Ala Thr Ser Leu Asp Pro G 355 360 365	ly Pro
Asp Gln Val Trp Val Pro Glu Glu Leu Ser Leu Ser Gly Glu T 370 375 380	yr Val
Arg Ala Arg Ile Lys Gly Val Asn Tyr Glu Pro Ile Asp Arg T 385 390 395	hr Lys 400
Tyr Thr Lys Val Leu Trp Asp Phe Asn Asp Gly Thr Lys Gln G 405 410 4	ly Phe 15
Gly Val Asn Gly Asp Ser Pro Val Glu Asp Val Val Ile Glu A	sn Glu
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Glu Gly Asn Tyr Trp Ala Asn Ala Arg Leu Ser Ala Asp Gly T 450 455 460	rp Gly
Lys Ser Val Asp Ile Leu Gly Ala Glu Lys Leu Thr Met Asp V 465 470 475	al Ile 480
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Ser Ala Asn Trp Val Asn Pro Asn Arg Ala Ile Lys Val Glu P 500 505 510	ro Thr
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Ser Ala Asp Ser Pro Ser Leu Glu Ala Ile Ala Met His Ala G 530 535 540	lu Asn
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Val Ile Tyr Leu Asp Asn Ile Lys Val Ile Gly Thr Glu Val G 565 570 5	lu Ile 75
Pro Val Val His Asp Pro Lys Gly Glu Ala Val Leu Pro Ser V 580 585 590	al Phe
Glu Asp Gly Thr Arg Gln Gly Trp Asp Trp Ala Gly Glu Ser G 595 600 605	ly Val
Lys Thr Ala Leu Thr Ile Glu Glu Ala Asn Gly Ser Asn Ala L 610 615 620	eu Ser
Trp Glu Phe Gly Tyr Pro Glu Val Lys Pro Ser Asp Asn Trp A	la Thr 640
Ala Pro Arg Leu Asp Phe Trp Lys Ser Asp Leu Val Arg Gly G	lu Asn 55
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tta ggt aat gac aat gtt aaa cgc cct tct gag gct ggc gca tta caa 1577 Leu Gly Asn Asp Asn Val Lys Arg Pro Ser Glu Ala Gly Ala Leu Gln 45 50 55	
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gct att gaa aat gac atg tat gtc atc gtt gat tgg cat gta cat gca 1865 Ala Ile Glu Asn Asp Met Tyr Val Ile Val Asp Trp His Val His Ala 140 145 150	
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cca att gat gat cac cat aca atg tat act gtt cac ttc tac act ggt 2201 Pro Ile Asp Asp His His Thr Met Tyr Thr Val His Phe Tyr Thr Gly 250 255 260 265	
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gta gca gta ttt gca aca gag tgg gga act agc caa gca aat gga gat 2345 Val Ala Val Phe Ala Thr Glu Trp Gly Thr Ser Gln Ala Asn Gly Asp 300 305 310	

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					tgg Trp 335									2441
					aca Thr									2489
					cca Pro									2537
					gta Val									2585
					aaa Lys									2633
					ttt Phe 415									2681
					gaa Glu									2729
					tct Ser									2777
					gga Gly									2825
					att Ile									2873
					cca Pro 495									2921
					act Thr									2969
					act Thr				Ser					3017
	_		_	_	aat Asn								_	3065
					gat Asp									3113
		_	_	_	att Ile 575		-	-		-		 _	_	3161
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					tca Ser									3305

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Arg Pro Ser Glu Ala Gly Ala Leu Gln Leu Gln Glu Val Asp Gly Gln 55

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Ser	Thr	His	Gly	Leu 85	Gln	Trp	Phe	Pro	Glu 90	Ile	Leu	Asn	Asp	Asn 95	Ala
Tyr	Lys	Ala	Leu 100	Ala	Asn	Asp	Trp	Glu 105	Ser	Asn	Met	Ile	Arg 110	Leu	Ala
Met	Tyr	Val 115	Gly	Glu	Asn	Gly	Tyr 120	Ala	Ser	Asn	Pro	Glu 125	Leu	Ile	ГÀа
Ser	Arg 130	Val	Ile	Lys	Gly	Ile 135	Asp	Leu	Ala	Ile	Glu 140	Asn	Asp	Met	Tyr
Val 145	Ile	Val	Asp	Trp	His 150	Val	His	Ala	Pro	Gly 155	Asp	Pro	Arg	Asp	Pro 160
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Gln	Arg	Pro	Asp	Leu 245	Ala	Ala	Asp	Asn	Pro 250	Ile	Asp	Asp	His	His 255	Thr
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Ser	Tyr	Pro 275	Pro	Glu	Thr	Pro	Asn 280	Ser	Glu	Arg	Gly	Asn 285	Val	Met	Ser
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Trp 305	Gly	Thr	Ser	Gln	Ala 310	Asn	Gly	Asp	Gly	Gly 315	Pro	Tyr	Phe	Asp	Glu 320
Ala	Asp	Val	Trp	Ile 325	Glu	Phe	Leu	Asn	Glu 330	Asn	Asn	Ile	Ser	Trp 335	Ala
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Phe	Glu	Leu 355	Gly	ГÀа	Ser	Asn	Ala 360	Thr	Ser	Leu	Asp	Pro 365	Gly	Pro	Asp
Gln	Val 370	Trp	Val	Pro	Glu	Glu 375	Leu	Ser	Leu	Ser	Gly 380	Glu	Tyr	Val	Arg
Ala 385	Arg	Ile	ГÀа	Gly	Val 390	Asn	Tyr	Glu	Pro	Ile 395	Asp	Arg	Thr	ГÀа	Tyr 400
Thr	Lys	Val	Leu	Trp 405	Asp	Phe	Asn	Asp	Gly 410	Thr	Lys	Gln	Gly	Phe 415	Gly
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Gly	Ala	Leu 435	Lys	Leu	Ser	Gly	Leu 440	Asp	Ala	Ser	Asn	Asp 445	Val	Ser	Glu
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Ala Asp Ser Pro Ser Leu Glu Ala Ile Ala Met His Ala Glu Asn Asn
Asn Ile Asn Asn Ile Ile Leu Phe Val Gly Thr Glu Gly Ala Asp Val
Ile Tyr Leu Asp Asn Ile Lys Val Ile Gly Thr Glu Val Glu Ile Pro
Val Val His Asp Pro Lys Gly Glu Ala Val Leu Pro Ser Val Phe Glu
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Thr Ala Leu Thr Ile Glu Glu Ala Asn Gly Ser Asn Ala Leu Ser Trp
Glu Phe Gly Tyr Pro Glu Val Lys Pro Ser Asp Asn Trp Ala Thr Ala
                   630
Pro Arg Leu Asp Phe Trp Lys Ser Asp Leu Val Arg Gly Glu Asn Asp
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Tyr Val Thr Phe Asp Phe Tyr Leu Asp Pro Val Arg Ala Thr Glu Gly
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Arg Asp Ile Thr Asn Ile Gln Asp Asp Thr Leu Leu Arg Asn Met Met
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Ile Ile Phe Ala Asp Val Glu Ser Asp Phe Ala Gly Arg Val Phe Val
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atc aca ctc att atg tca cta ttt gtt cct atg gct tca gca aac aca Ile Thr Leu Ile Met Ser Leu Phe Val Pro Met Ala Ser Ala Asn Thr 20 25 30	697
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		gta Val														1465
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	Glu	cac His														1609
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	_	gca Ala	_			_			_							1945
		gct Ala 450														1993
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	Val	ttt Phe														2089
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		gca Ala 530								_		_	_		_	2233
		cca Pro														2281

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									gtt Val 585							2377
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									ccg Pro 665							2617
									tgg Trp							2665
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_		_					_	_	tca Ser				_			2761
_		_		_	_			_	agc Ser						_	2809
									tta Leu 745							2857
									cac His							2905
						_	_		act Thr	_				_		2953
		_			_	_		_	ctt Leu			_	_	_		3001
			-				-		gcg Ala			-	_			3049
_	_			_		_		_	cta Leu 825	_				_	_	3097
	_	_			_	_			agt Ser		_				_	3145
-	_	_	_		_		_		tca Ser			_	_			3193
		-						-	gtt Val					-		3241

Amp Leu Thr Glu Leu Asp Ser Ala Thr Val Thr Ser Amp Glu Leu Tyr 8880 885 890 891 895 391 10 Leu Tyr 8885 895 895 895 895 394 396 396 395 395 394 394 394 395 395 395 395 394 394 394 394 394 394 395 395 395 394 394 395 <																	
His Tyr Glu Val Lys Ile Asn Ile Arg Asp Ile Glu Ala Ile Thr Asp 900 900 900 910 910 910 910 910 910 910	Asp					Asp					Thr					Tyr	3289
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Asp Phe Ala Gly Arg Val Phe Val Asp Asn Val Arg Phe Glu 930 taatttaaaa acagtagata gagagactct ctatctacct gtttattgct tactattcgt : cttccacttt t : <pre></pre>	_			Leu	_				Leu			_	_	Glu	_	_	3385
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1									_	_	_	_	_	_			
Ser Lys Ser Asn Ala Phe Pro Phe Ser Asp Val Lys Lys Thr Ser Ser Phe Pro Tyr Ile Lys Asp Leu Tyr Glu Gln Glu Val Ile Thr Glu Far		Lys	Ile	Lys		Ile	Lys	Gln	Ser		Ser	Leu	Leu	Leu		Ile	
Ser Phe Pro Tyr I le Lys Asp Leu Tyr Glu Gln Glu Val I le Thr Ser Phe Pro Tyr I le Lys Asp Leu Tyr Glu Gln Glu Val I le Thr Gly Thr Ser Ala Thr Thr Phe Ser Pro Thr Asp Ser Val Thr Arg Ala 80	Thr	Leu	Ile		Ser	Leu	Phe	Val		Met	Ala	Ser	Ala		Thr	Asn	
So	Glu			Ser	Asn	Ala	Phe		Phe	Ser	Asp	Val	_	Lys	Thr	Ser	
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S		Thr	Ser	Ala	Thr		Phe	Ser	Pro	Thr		Ser	Val	Thr	Arg		
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Phe Ala Pro Asn Glu Asn Ile Thr Arg Glu Gln Met Ala Ala Met Ala Ala Arg Arg Arg Arg Arg Arg Glu Tyr Asn Asp Ser Ser Glu Glu Ser Thr Phe Ala Gln Asp Asp Arg	ГÀа	Asp	Tyr		Phe	Lys	Asp	Arg		Asn	Trp	Ala	Tyr		Glu	Ile	
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145 150 155 160 Gln Arg Glu Tyr Asn Asp Ser Ser Ser Ser Ile Ser Thr Phe Ala Gln Asp 165 Ser Ser Ile Ser Thr Phe Ala Gln Asp 175 Asp 175 Ala Val Gln Lys Ala Tyr Val Leu Glu Leu Glu Leu Glu Glu Gly Asn Thr Asp 190 Asp 200 Thr Arg Glu Gln Ser Ala Lys 205 Gly Tyr Phe Gln Pro Lys Arg Asn Ser Thr Arg Glu Gln Ser Ala Lys 200 200 Thr Arg Glu Asp Tyr Leu Tyr 220 Val Ile Ser Thr Leu Leu Trp 215 Lys Val Ala Ser His Asp Tyr Leu Tyr 220 His Thr Glu Ala Val Lys Ser Pro Ser Glu Ala Gly Ala Leu Gln Leu 240 Val Glu Leu Asn Gly Gln Leu Thr Leu Ala Gly Glu Asp Gly Thr Pro 255 Val Gln Leu Arg Gly Met Ser Thr His Gly Leu Gln Trp Phe Gly Glu	Phe		Pro	Asn	Glu	Asn		Thr	Arg	Glu	Gln		Ala	Ala	Met	Ala	
Ala Val Gln Lys Ala Tyr Val Leu Glu Leu Met Glu Gly Asn Thr Asp 180 Tyr Phe Gln Pro Lys Arg Asn Ser Thr Arg Glu Gln Ser Ala Lys 205 Thr Leu Tyr 210 Thr Glu Ala Val Lys 230 Ser Pro Ser Glu Ala Gly Asn Tyr Leu Tyr 240 Tyr Glu Leu Asn Gly Gln Leu Thr Leu Ala Gly Gln Asp Gly Thr Pro 255 Thr Leu Arg Glu Leu Arg Gly Met Ser Thr His Gly Leu Gln Trp Phe Gly Glu		Arg .	Ala	Tyr	Glu	-	Leu	Glu	Asn	Glu		Ser	Leu	Pro	Glu		
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Thr Ser Pro Glu	Asp Asn Thr Met Ser Asn Ile 725 730	Ile Leu Phe Val Gly 735	
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Leu Pro Ser Thr 770	Phe Glu Asp Gly Thr Arg Gln 775	Gly Trp Asp Trp His 780	
Thr Glu Ser Gly 785	Val Lys Thr Ala Leu Thr Ile 790 795	Glu Glu Ala Asn Gly 800	
Ser Asn Ala Leu	Ser Trp Glu Tyr Ala Tyr Pro 805 810	Glu Val Lys Pro Ser 815	
Asp Gly Trp Ala 820	Thr Ala Pro Arg Leu Asp Phe 825	Trp Lys Asp Glu Leu 830	
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Pro Ala Asn Gly 865	Tyr Trp Gln Glu Val Pro Thr 870 875	Thr Phe Glu Ile Asp 880	
Leu Thr Glu Leu	Asp Ser Ala Thr Val Thr Ser 885 890	Asp Glu Leu Tyr His 895	
Tyr Glu Val Lys 900	Ile Asn Ile Arg Asp Ile Glu 905	Ala Ile Thr Asp Asp	
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	gca cca atg gtg agt gca gaa Ala Pro Met Val Ser Ala Glu 25		332
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	У									ttt Phe							572
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	.a									gaa Glu							860
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Gln	Pro	Pro	Ala	Ala 725	Gly	Tyr	Trp	Ala	Gln 730	Ala	Ser	Glu	Thr	Phe 735	Glu
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Tyr	His	Tyr 755	Glu	Val	Glu	Ile	Asn 760	Ile	Glu	Asp	Ile	Glu 765	Asn	Asp	Ile
Glu	Leu 770	Arg	Asn	Leu	Met	Leu 775	Ile	Phe	Ala	Asp	Asp 780	Glu	Ser	Asp	Phe
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Thr	Lys	Val	Glu	Val 805	Leu	Glu	Arg	Asn	Ile 810	Asn	Glu	Leu	Gln	Glu 815	Gln
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What is claimed is:

- 1. A method for enhancing anti-redeposition ability of an alkaline cellulase, or method for both enhancing anti-redeposition ability and protease resistance of an alkaline cellulase, the method comprising substituting a glutamine residue at the position corresponding to position 58 of the amino acid sequence of an alkaline cellulase as set forth in SEQ ID NO: 2 or of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2, with a glutamic acid or arginine residue.
- 2. The method of claim 1, wherein the glutamine residue is substituted with a glutamic acid residue.
- 3. The method of claim 1, wherein the glutamine residue is substituted with an arginine residue.
- 4. The method of claim 1, wherein the alkaline cellulase 50 that has at least 90% identity with the amino acid sequence set forth in SEQ ID NO: 2 lacks the signal sequence that corresponds to amino acids 1 to 30 of SEQ ID NO: 2.
- 5. The method of claim 1, wherein the method further comprises substituting a glutamine residue at the position 55 corresponding to position 71 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2, with a glutamic acid residue.
- 6. The method of claim 1, wherein the method further $_{60}$ comprises substituting a serine residue at the position corresponding to position 193 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2, with an arginine residue.

7. The method of claim 1, wherein the method further comprises substituting a glutamine residue at the position corresponding to position 242 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2, with a serine residue.

- 8. The method of claim 1, wherein the method further comprises substituting an asparagine residue at the position corresponding to position 419 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2, with an alanine residue.
- 9. The method of claim 1, wherein the method further comprises substituting an aspartic acid residue at the position corresponding to position 421 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2, with an alanine residue.
- 10. The method of claim 1, wherein the method further comprises substituting a tryptophan residue at the position corresponding to position 454 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2, with a tyrosine residue.
- 11. The method of claim 1, wherein the method further comprises substituting a tryptophan residue at the position corresponding to position 501 of the amino acid sequence of an alkaline cellulase set forth in SEQ ID NO: 2 or of an amino acid sequence having at least 90% identity with the amino acid sequence of SEQ ID NO: 2, with a tyrosine residue.